On the Pterophoridae (Lepidoptera) of Colombia

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Abstract: Twelve species of Pterophoridae were collected in the Department of Cundinamarca, Colombia during three days in July 2018. From this sample, the new species *Postplatyptilia chicaquensis* Landry & Gielis, **sp. n.** and *Hellinsia danielae* Landry & Gielis, **sp. n.** are described and five other species are reported as new for Colombia. Adults of all 12 species are illustrated, as well as the genitalia of the sex(es) collected. Additionally, a list of all of the 53 species reported from Colombia is presented.

Key words: *Adaina*, *Exelastis*, *Lioptilodes*, *Oidaematophorus*, plume-moths, Pterophoroidea.

Resumen: En el departamento de Cundinamarca, Colombia, se hicieron recolectas de la familia Pterophoridae durante tres días en julio del 2018. Se recolectaron 12 especies de las cuáles cinco son reportadas por primera vez para el país. Así mismo se describen las nuevas especies *Postplatyptilia chicaquensis* Landry & Gielis, **sp. n.** y *Hellinsia danielae* Landry & Gielis, **sp. n.** Los adultos son ilustrados, así como las genitalias de los sexos recolectados. Adicionalmente, se presenta una lista de todas las 53 especies reportadas para Colombia.

Palabras claves: *Adaina*, *Exelastis*, *Lioptilodes*, *Oidaematophorus*, polillas plumas, Pterophoroidea.

INTRODUCTION

Pterophoridae, or plume-moths, are small to medium-sized gracile moths found on all continents except Antarctica. The world fauna of Pterophoridae comprises 1554 species, that of the Neotropical region 467 species, and that of Colombia 46 species (Hobern, 2022 and pers. comm.). The Pterophoridae fauna of Colombia has never been treated specifically in a taxonomic or faunistic publication. Publications with species records and descriptions of Pterophoridae of Colombia had a broader geographical scope, from either a few countries (Gielis, 2014) to a continental scale (Gielis, 2006, 2011) to an even larger scale (Zeller, 1877). The Andean mountains provide a unique continuum of great ecological variation in temperature, humidity, altitude, and precipitation, resulting in numerous ecological niches that support the development of local species. However, small moths are poorly sampled in the Neotropical region and, consequently, the true diversity and range of the species are mostly unknown.

Thanks to Helber Adrian Arévalo Maldonado and Prof. Francisco Javier Serna Cardona of the University of Colombia, the first author had the opportunity to collect moth specimens in Colombia in 2018. Twenty-six specimens of Pterophoridae were collected at light in a private nature park a few kilometers away from Bogotá, the capital of the country. Here we illustrate with photos the adults and genitalia of the 12 species collected, including a new species in the genus *Postplatyptilia* Gielis and another in the genus *Hellinsia* Tutt.

MATERIAL AND METHODS

The specimens of Pterophoridae discussed here were collected at light with the use of a LepiLED (Brehm, 2017) placed inside a 2-meter-high tower of white gauze. The collecting locality, chosen for convenience and quality of its variety of habitats, is Parque Natural Chicaque (www.chicaque.com), San Antonio de Tequendama, department of Cundinamarca. Most specimens were collected around the accommodation structure of the park known as the 'Refugio', located at 2150 m and 04°36'53"N, 74°18'40"W. In addition, one specimen came to light at 2250 m in the park's oak (*Quercus humboldtii* Bonpland) forest (04°36'22"N, 74°18'17"W). Also collecting with B. Landry were H. Arévalo and Jurate De Prins.

The specimens collected are mostly deposited in the insect collection of the Universidad Nacional Agronomía Bogotá, Museo Entomológico de la Facultad de Ciencias Agrarias de la Universidad Nacional de Colombia (UNAB), except for a few deposited in the Muséum d'histoire naturelle, Geneva, Switzerland (MHNG).

The online catalogue maintained by Hobern (2022) was used to obtain lists of species known for each of the genera found at Chicaque and to find out which species were already known from Colombia. Based on the world catalogue of Gielis (2003), the Holbern catalogue continues to be fed by the identification database and distribution data of C. Gielis. The generic classification adopted here and order of species presented below reflect those of Gielis (2006, 2011).

The data on the labels of the holotypes are recorded verbatim, with missing information in square brackets and vertical bars representing changes of lines. The term wingspan is abbreviated 'ws' in the figure legends. Morphological terms follow Gielis (2006, 2011).

RESULTS

Postplatyptilia parana Gielis, 1996

Postplatyptilia parana Gielis, 1996: 92.

Figs. 1A, 3a, b

Type locality. Brazil, Paraná, Castro.

Distribution. Argentina (Jujuy), Brazil (Paraná, Rio de Janeiro; Santa Catarina) and Costa Rica (Cartago) (Gielis, 2006). This species is reported from Colombia for the first time.

Remarks. One male specimen was collected at 2250 m in Chicaque's oak forest. The female genitalia are illustrated by Gielis (1996, 2006).

Postplatyptilia chicaquensis Landry & Gielis, **sp. nov.**

Figs. 1B, 1C, 3c, d, 5a

Description. MALE (n=3) (Figs. 1B, 1C). Head blackish brown on vertex and above slightly produced fronto-clypeus medially; paler brown laterally above on fronto-clypeus; paler, grayish white between antennae and below fronto-clypeus. Antennae with alternating blackish brown and grayish white rows of scales. Labial palpus porrect, with second palpomere reaching tip of fronto-clypeus, without scale tuft, with palpomere I dark blackish brown laterally, white dorsally and ventrally, palpomere II blackish brown laterally with white at apex, palpomere III blackish brown at base, pale chestnut brown on distal half. Thorax coloration dorsally (Fig. 1B, 1C) mostly brown of various shades, ochraceous on tegulae and black on metathorax lined with white apically along lateral margins. Forewing 7.0-8.0 mm in length, wingspan 15.0-17.5 mm (holotype: 17.5 mm); coloration and markings similar to congeners with blackish-brown costal triangle, subterminal thin cream lines across lobes, and two dark brown scale teeth along anal margin. Hindwing uniformly blackish brown except for small medial white patch on third lobe; third lobe fringes interspersed with dark spatulate scales including apical scale tooth on anal margin, white at apex. Foreleg coxa and femur blackish brown with few dispersed white scales; tibia blackish brown with longitudinal white stripes and spots of variable sizes; tarsomere I pale golden-white with long longitudinal blackish brown stripes dorsally and shorter one laterally on distal half; tarsomere II and III pale golden-white with longitudinal blackish brown stripe dorsally, that on tarsomere III shorter; tarsomeres IV and V pale golden-white. Midleg coxa white with blackish brown mostly at base and apex; femur mostly blackish brown with few white scales mostly at apex; tibia blackish brown with pair of white stripes interrupted at submedian and apical scale tufts, tibial spurs blackish brown and white, reaching just beyond middle of tarsomere I; tarsomere I and II striped blackish brown and pale golden-white; tarsomeres III and IV mostly pale golden-white with few blackish brown scales dorsally, more so on tarsomere III; tarsomere V uniformly pale golden-white. Hindleg coxa not visible; femur blackish brown with small white spots; tibia with four alternating pale golden-white and blackish brown areas, darker blackish brown at base of spurs, with few white scales apically, spurs blackish brown at their base and apex, pale golden-white medially, with median tibial spurs slightly longer, about as long as second tarsomere; tarsomere I blackish brown at base and beyond 1/3, pale golden-white in between; tarsomere II mostly pale golden-white, blackish brown at tip; tarsomeres III-V missing on holotype, III and IV as 2nd, V entirely pale golden-white. Abdomen dorsally (Fig. 1B, 1C) with mixed blackish brown, paler brown and white scales, also with blackish brown spots near middle and on other segments; ventrally on basal half mostly dark blackish brown, with few white scales, on distal half paler, grayish brown, with lateral white spots on apical margins.

Variation. One of the males collected is much darker than the others, especially on the forewing, with markings therefore less contrasted (Fig. 1C).

Male genitalia (Figs. 3c, d, n=2). Uncus straight, positioned ventrally on tegumen slightly beyond middle of tegumen along mid length, barely reaching apical margin of tegumen, of medium girth, slightly narrowing from base to apex, with few setae of medium length. Tegumen moderate in size, apically truncate. Valva rather wide, distinctly enlarged at 2/3, with broadly rounded margin dorsally; sacculus simple, extended to mid length of valva, with thick setae on basal half along dorsal margin; cucullus with ventral margin incorporating strong longitudinal sclerotized bar; apex shortly pointed. Saccus triangular, apically bifid with narrow cleft almost reaching mid length, not clearly visible on image because of overlap with juxta/anellus complex, with pointed arms of medium width. Juxta about as long as wide, with converging sclerotized bars laterally, more thickly sclerotized at rounded base of anellus arms; anellus arms of medium length, narrow along whole length, with few short setae. Phallus of medium girth; shaft slightly enlarged dorsally at base and at 2/3; coecum penis short, straight; vesica spiculose.

FEMALE (n=1). As male except for slightly longer labial palpus. Wingspan 18.0 mm; forewing length: 9.0 mm.

Female genitalia (Fig. 5a, n=1). Papillae anales short, weakly sclerotized, with few short to medium long setae. Apophyses posteriores thin, long, reaching anterior margin of tergum VIII. Tergum VIII with lateral oval windows medially at lateral margins. Tergum VII not modified. Sternite VII about one third longer than tergum VII, medially shortly concave. Lamella postvaginalis about half as wide as tergum VIII, laterally joining thin, pointed apophyses anteriores slightly shorter than papillae anales, apically reaching about ¾ length of tergum VIII and forming pair of rounded bumps separated by short, narrow median cleft. Antrum wide and long, funnel-shaped, wrinkled, slanted to left. Ductus seminalis about twice as long as antrum, thin, sclerotized towards antrum. Corpus bursae oval, large, slightly longer than ductus seminalis plus antrum, with pair of rather short, slightly curved signa with large, rounded area around their bases with lightly sclerotized pentagons.

Etymology. Named for the collecting locality, Chicaque Natural Park.

Specimens examined. Holotype ♂: 1- "COL[OMBIA]., Cundinamarca | S[an]. Antonio de Tequendama | P[arque]N[atural] Chicaque, n[ea]r Refugio, at | lights, 4°36'53"N 74°18'40"W | 2150 m, 19-20.vii.2018, *leg[it]*. B. | Landry, H. Arévalo, J. De Prins"; 2- "if selected as holotype | return to Museo | Entomológico, UNAB | Bogotá, Colombia"; 3- "Export permit | ANLA, Colombia | n° 01292, 2018"; 4- "HOLOTYPE | Postplatyptilia | chicaquensis Landry & Gielis". First three labels white, printed in black, fourth red, handwritten in black. Deposited in UNAB.

Paratypes: 2 ♂♂, same data as holotype, with dissected parts on slides BL 1895 and 1902, 1 ♀ with same data as holotype and dissected parts on slide BL 1898. Deposited in UNAB and MHNG.

Diagnosis. In external characters this species is similar to many others of the genus, but the white patch medially on the third lobe of the hind wing is possibly diagnostic. In any case, the genitalia need to be studied for a reliable determination. In male genitalia the small uncus barely projecting beyond the distal margin of the tegumen will separate this species from all others, the most similar in this respect being: *Postplatyptilia oxapampa* Gielis, 2014 described from Peru, for which the uncus is very short and knob-like and the anellus arms much shorter; *P. transversus* Gielis, 2006, described from Colombia, Cundinamarca, for which the small, thin uncus does not reach the elongate tegumen's apical margin, the anellus arms are longer and apically spatulate, and the saccus is simple, not bifid as in *P. chicaquensis*; and *P. parana* Gielis, mentioned above (Fig. 3a), in which the uncus is thin from base to apex, longer, and not reaching the apical margin of the pointed tegumen, the saccus is not so deeply cleft (the cleft is more rounded and the arms are narrower), and the valva is thinner. In female genitalia, the position and shape of the lamella postvaginalis along with the long funnel-shaped 'antrum' and the short ductus bursae collectively serve to identify this species. The most similar species is *P. zongoensis* Gielis, 2006 (fig. 352), described from Bolivia, for which the lamella postvaginalis is similar in shape

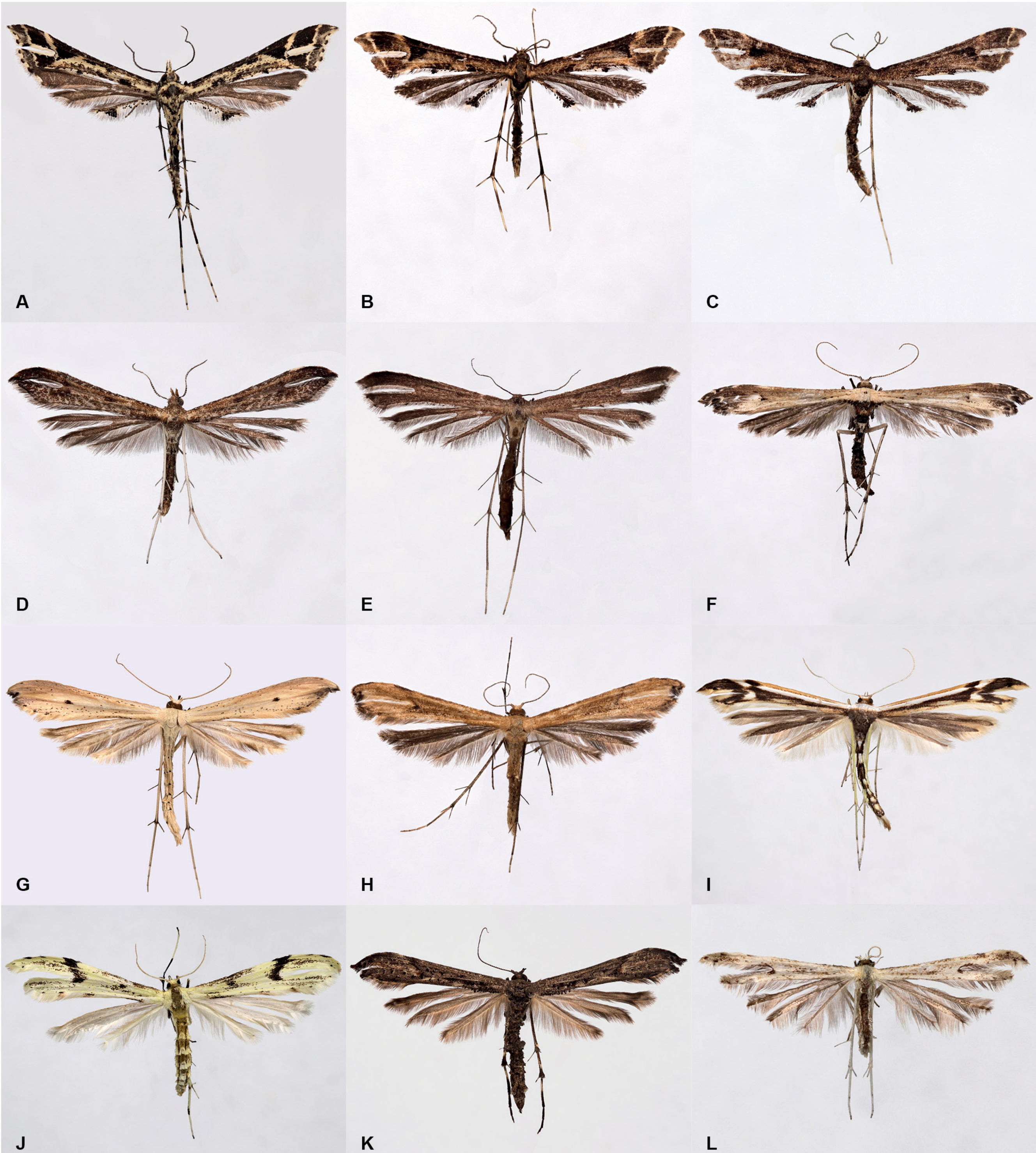

Figure 1. Pterophoridae species collected at Parque Natural Chicaque in 2018. **A.** *Postplatyptilia parana* Gielis, male, ws: 19.5 mm; **B.** *Postplatyptilia chicaquensis* **sp. n.**, holotype male, ws: 17.5 mm; **C.** *Postplatyptilia chicaquensis* **sp. n.**, paratype male, ws: 17.0 mm; **D.** *Lioptilodes cocodrilo* Gielis, male, ws: 17.0 mm; **E.** *Exelastis* sp., female, ws: 17.0 mm; **F.** *Hellinsia barbatus* (Gielis), male, ws: 23.0 mm; **G.** *Hellinsia danielae* **sp. n.**, holotype male, ws: 23.5 mm; **H.** *Hellinsia obscuricilia* Arenberger & Wojtusiak, male, ws: 19.0 mm; **I.** *Hellinsia pelospilus* (Zeller), male, ws: 26.5 mm; **J.** *Hellinsia* sp., female, ws: 26.5 mm; **K.** *Oidaematophorus pseudotrachyphloeus* Gielis, male, ws: 30.0 mm; **L.** *Adaina atahualpa* Gielis, female, ws: 17.0 mm. ws = wingspan.

but smaller and extending only to about 1/3 of the length of segment VIII and the antrum is also funnel-shaped and medially situated, but it is straight, narrower, and shorter. *Platyptilia palmeri* Gielis, 1996 (see also Gielis, 2006, fig. 355), described from Mexico, is also similar in female genitalia, but the papillae anales are long, there is no lamella postvaginalis but a lamella antevaginalis forming a pair of bumps apically and extending slightly beyond the basal margin of segment VIII, and the antrum is narrower, short, and not slanted to the left.

Figure 2. Pterophoridae species collected at Parque Natural Chicaque in 2018; *Adaina* sp., female, ws: 21.0 mm.

Distribution. This species presently is known from the type locality only.

Remarks. *Postplatyptilia* Gielis, 1991 includes 40 species distributed in the Neotropical region (Hobern, 2022). Only one of these, *P. pusillus* (Philippi, 1860), cannot be recognized as its description is not sufficiently diagnostic and the type specimen is lost (Gielis, 2006). Another one, *P. paraglyptis* (Meyrick, 1908) described from Argentina, is represented by a unique type specimen without abdomen, but its habitus (see Gielis, 2006) allows us to recognize it as different from the species described here based on the forewing markings on the first lobe. The fact that one of the sexes is unknown for many species of *Postplatyptilia* means that the new species diagnosis is *de facto* incomplete.

Lioptilodes cocodrilo Gielis, 2006
Lioptilodes cocodrilo Gielis, 2006: 152.
Figs. 1D, 3e, f

Type locality. Ecuador, Napo Province, 15 km SE Cosanga, Cocodrilo.
Distribution. Previously known from Ecuador and Peru (Gielis, 2006), the species is reported from Colombia for the first time.
Remarks. Two male specimens were collected around the Refugio of Parque Natural Chicaque. The female genitalia are illustrated by Gielis (2006, fig. 396).

Exelastis **sp.**
Figs. 1E, 5b

Remarks. The unique specimen collected around the Refugio of Parque Natural Chicaque is a female. In female genitalia this specimen is similar to *Exelastis pumilio* (Zeller, 1873) in the absence of a signum on the corpus bursae. However, the central position of the ostium bursae on abdominal segment VII in this specimen does not agree with that shown for *E. pumilio* by Gielis (2006, fig. 408), Landry (1993, fig. 7), and Matthews *et al.* (2019, fig. 46) in which the ostium bursae is positioned at the terminal margin of abdominal segment VII. Also, in the Chicaque specimen the shape of the antrum, with distinctly parallel sides and being not much wider than the ductus bursae, and the posterior part of the ostium subtended by an elongate sclerite, differ from the shorter antrum with divergent sides and the absence of a posterior sclerite in *E. pumilio*. The wingspan indicated by Gielis (2006: 195) for *E. pumilio* is 12-15 mm while the Chicaque specimen reaches 17 mm; this female also possesses spatulate scales in the fringe of the third hindwing lobe, which are absent in *E. pumilio* based on Gielis (2006), Landry (1993), and Matthews *et al.* (2019). We conclude that this species is probably new, but male specimens would need to be available for study to conclude with certainty and provide a description.

Oidaematophorus pseudotrachyphloeus Gielis, 2011
Oidaematophorus pseudotrachyphloeus Gielis, 2011: 697.
Figs. 1K, 4h, i, 5d

Type locality. Peru, Lima, 'Reserva Nacional de Lachay'.
Distribution. Argentina, Ecuador, Peru (Gielis, 2011) and Chile (Vargas, 2021). It is reported for Colombia for the first time.
Remarks. Six specimens, three males and three females, were collected around the Refugio of Parque Natural Chicaque. The rather large tufts of raised scales on the forewing and abdomen are remarkable.

Hellinsia barbatus **(Gielis, 1996)**
Oidaematophorus barbatus Gielis, 1996: 97.
Figs. 1F, 3g-i

Type locality. Colombia, Sierra del Libano.
Distribution. Brazil (new record from: Santa Catarina, São Joaquim, 1400 m, 2.ii.1993, *leg.* V.O. Becker, genitalia preparation CG7103, Coll. V.O. Becker), Colombia, Costa Rica, Venezuela (Gielis, 2011, 2014).
Remarks. This species is represented in the Parque Natural Chicaque sample of 2018 by one male collected near the Refugio. The female genitalia are illustrated by Gielis (2011, fig. 345).

Hellinsia danielae Landry & Gielis, **sp. n.**
Figs. 1G, 4a-c

Description. MALE (Fig. 1G). Head wheat-colored between antennae, chocolate-brown on vertex and on flat fronto-clypeus. Antennae wheat-colored on scape except for two chocolate-brown spots at apex; pedicel and basal 8 flagellomeres wheat-colored with small chocolate-brown spots, grayish white on subsequent flagellomeres. Labial palpus porrect, with ventrally projecting scales on first palpomere and short appressed scales on distal two palpomeres; palpomere I white to pale grayish white; palpomere II mostly pale grayish brown, paler ventrally; palpomere III slightly darker grayish brown. Thorax mostly wheat-colored, light ochraceous at base of tegulae. Forewing length: 11 mm; wingspan: 23.5 mm; coloration and markings wheat-colored with dark brown spot at cleft base and scattered dark brown scales along costa and anal margin; first lobe with R_5 terminus marked by black spot and gray and black fringes; second lobe M_3, Cu_1, and Cu_2 terminals also marked with gray scales. Hindwing uniformly wheat-colored, fringes pale drab. Foreleg coxa wheat-colored with thin chocolate-brown stripe, femur and tibia longitudinally striped pale wheat-colored and chocolate brown; tarsomere I wheat-colored with thin chocolate-brown stripe not reaching apex; tarsomeres II-V wheat-colored. Midleg as in foreleg, with tibial tufts blackish brown, with few white scales at apex of apical tuft, tibial spurs dark brown dorsally, paler grayish brown ventrally. Hindleg mostly wheat-colored, with blackish brown at base of tibial spurs, dorsally on spurs, and at very apex of tarsomeres I-IV. Abdomen dorsally as shown; ventrally not recorded.

Male genitalia (Figs. 4a-c, n=1). Uncus thin, thinning toward apex, pointed, with sparse short setae. Tegumen rather long and narrow, more than twice as long as uncus, with broad triangular anterior projection dorsomedially. Valvae asymmetrical, both with hair pencils of moderate size and density, with apices somewhat pointed; left valva broader, with long, thin saccular spine first directed medially for short distance, then straight anteriorly, then curved back

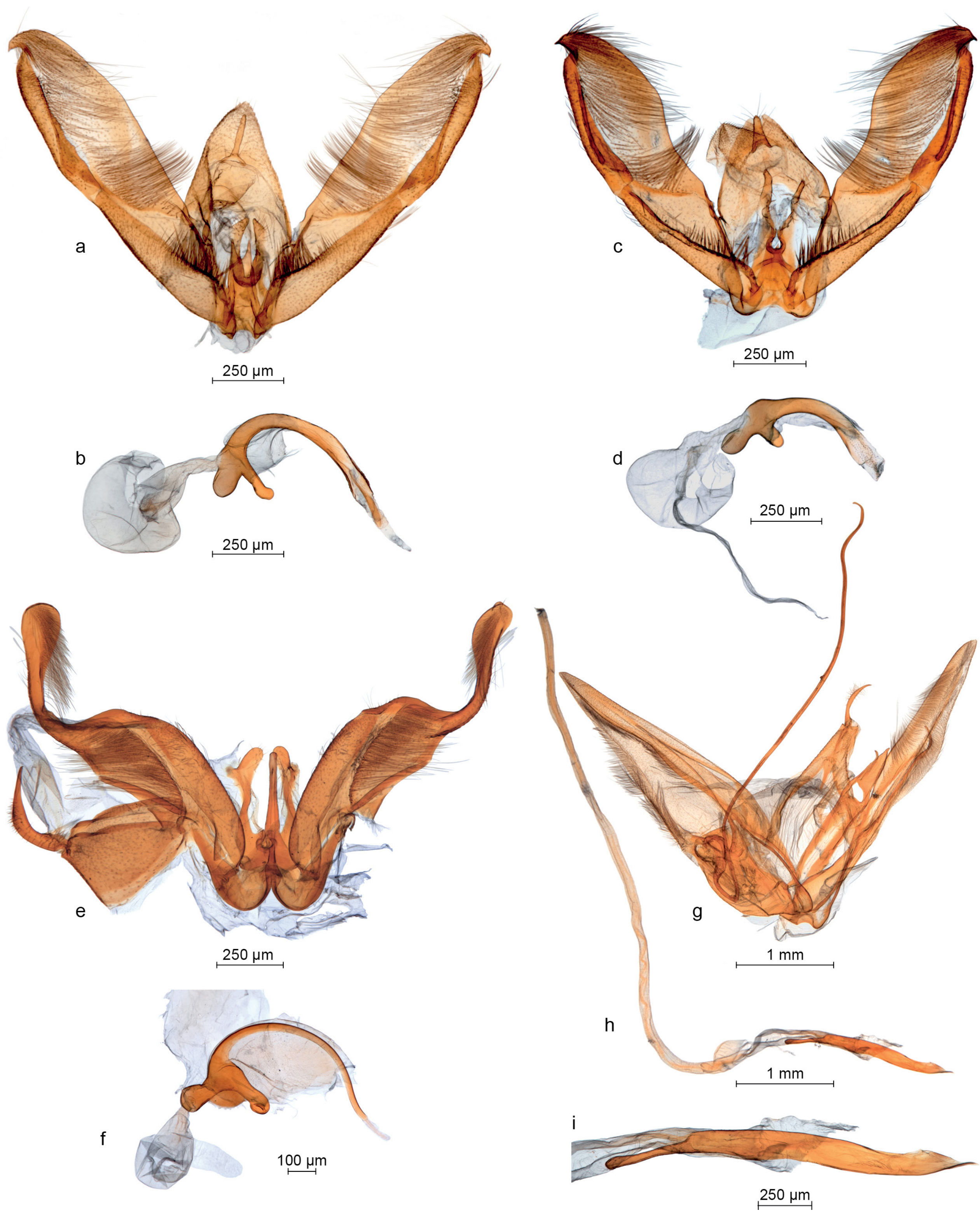

Figure 3. Male genitalia of Pterophoridae species collected at Parque Natural Chicaque in 2018; genital capsule above phallus. **a-b**. *Postplatyptilia parana* Gielis; **c-d**. *Postplatyptilia chicaquensis* **sp. n.**; **e-f.** *Lioptilodes cocodrilo* Gielis; **g-i.** *Hellinsia barbatus* (Gielis).

posteriorly, and reaching tip of valva; saccus of right valva with short, broad triangular process associated with subventral sclerotized bar reaching slightly beyond mid-length. Juxta narrow, elongate, about half as long as valvae, with short asymmetrical anellus arms with sparse short setae, the right arm wider. Phallus thin, slightly down-curved, about half as long as valvae, apically pointed; vesica adorned with thin short plate with basal triangular projection.

FEMALE. Unknown.

Etymology. This species is dedicated to Daniela Cubillos Cañizares, Colombian engineer in agronomy, myrmecologist, and friend, for her most kind and efficient help to BL with fieldwork in the Colombian Amazon.

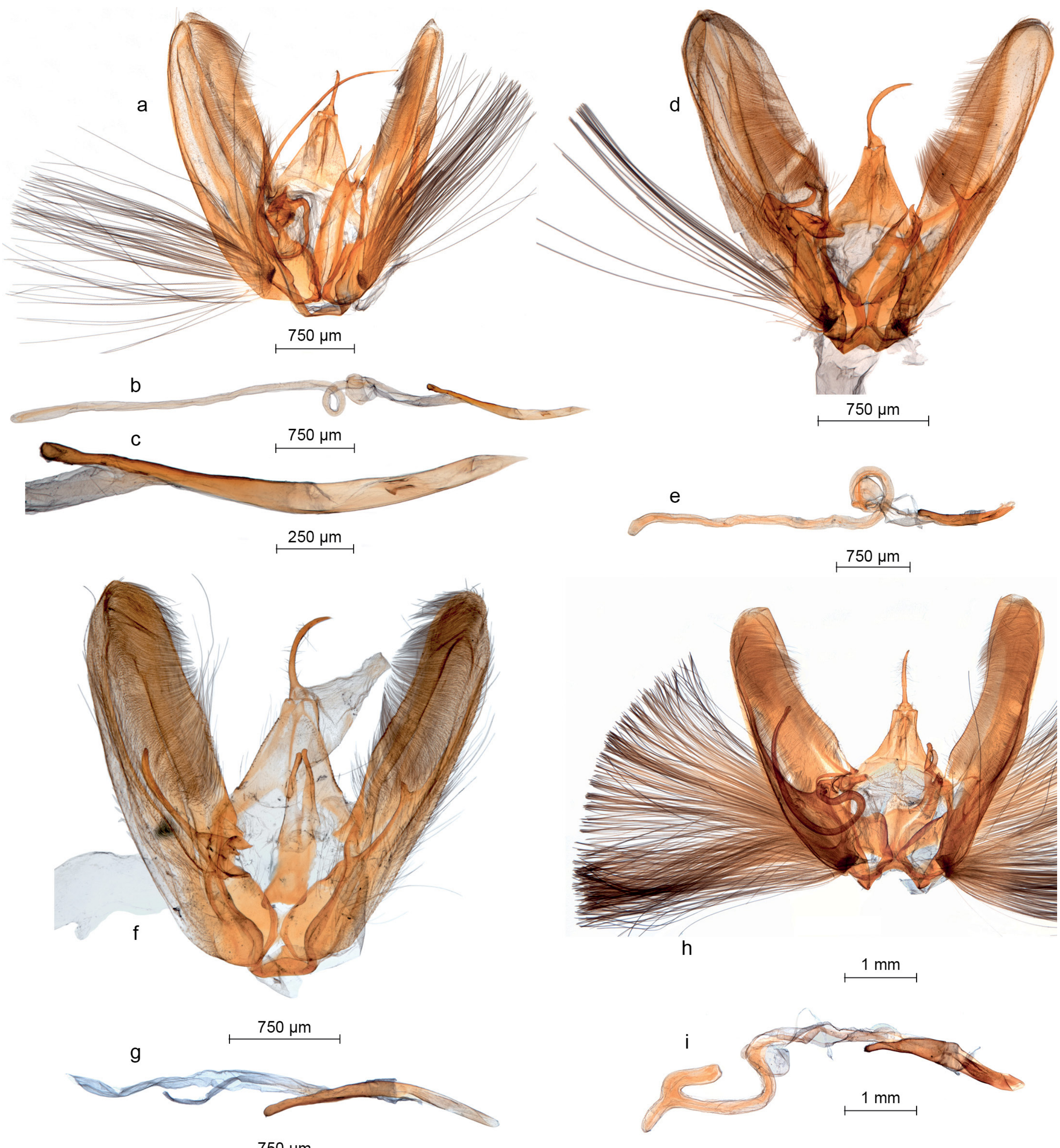

Figure 4. Male genitalia of Pterophoridae species collected at Parque Natural Chicaque in 2018; genital capsule above phallus. **a-c.** *Hellinsia danielae* **sp. n.**; **d-e**. *Hellinsia obscuricilia* Arenberger & Wojtusiak; **f-g.** *Hellinsia pelospilus* (Zeller); **h-i.** *Oidaematophorus pseudotrachyphloeus* Gielis.

Specimen examined. Holotype ♂: 1- "COL[OMBIA]., Cundinamarca | S[an]. Antonio de Teguendama | P[arque]N[atural] Chicaque, n[ea]r Refugio, at | lights, 4°36'53"N 74°18'40"W | 2150 m, 19-20.vii.2018, *leg[it]*. B. | Landry, H. Arévalo, J. De Prins"; 2- "if selected as holotype | return to Museo | Entomológico, UNAB | Bogotá, Colombia"; 3- "Export permit | ANLA, Colombia | n° 01292, 2018"; 4- "Slide | BL 1905♂"; 5- "HOLOTYPE | Hellinsia | danielae | Landry & Gielis". First three labels white, printed in black, fourth green, handwritten in black, fifth red, handwritten in black. Deposited in UNAB.

Diagnosis. The species has pale forewings with very limited markings, as shown (Fig. 1G). A similar pattern is present also in *Hellinsia capucayae* Gielis, 2012, *H. cosangae* Gielis, 2012 and *H. lenis* (Zeller, 1877). All these species differ in the shape of the male genitalia. The present species has a long saccular spine, which is basally curved back at the base of the valve, consistent with "type E" of Gielis (2011), whereas the other species have a different shape of their saccular processes (*H. capucayae* (type B); *H. cosangae* (type B), and *H. lenis* (type C)).

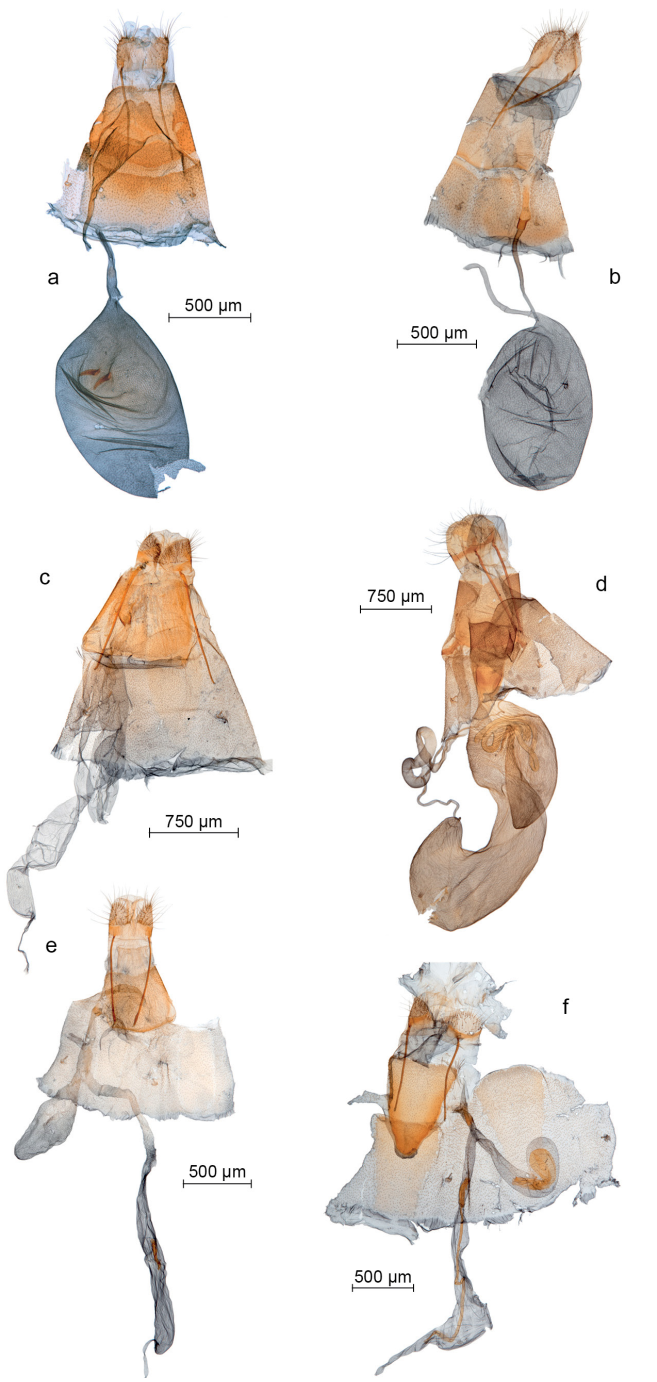

Figure 5. Female genitalia of Pterophoridae species collected at Parque Natural Chicaque in 2018. **a.** *Postplatyptilia chicaquensis* sp. n.; **b.** *Exelastis* sp.; **c.** *Hellinsia* sp.; **d.** *Oidaematophorus pseudotrachyphloeus* Gielis; **e.** *Adaina atahualpa* Gielis; **f.** *Adaina* sp.

Distribution. Presently this species is known only from the type locality, in Colombia.

Remarks. The holotype and unique specimen known is missing the left midleg. The genitalia had been mounted on slide before they were described.

***Hellinsia obscuricilia* Arenberger & Wojtusiak, 2001**

Hellinsia obscuricilia Arenberger & Wojtusiak, 2001: 70.

Figs. 1H, 4d, e

Type locality. Venezuela, Paramo El Batallon, Quebrada de los Pios, 2950 m.

Distribution. Bolivia (Gielis, 2013), Costa Rica (Gielis, 2011), Honduras (Kovtunovich *et al.*, 2018), Panama (Kovtunovich *et al.*, 2019), Venezuela. Reported here from Colombia for the first time.

Remarks. Only one male of this species was collected at Chicaque near the Refugio in 2018. The female is yet unknown.

***Hellinsia pelospilus* (Zeller, 1877)**

Leioptilus pelospilus Zeller, 1877: 481.

Figs. 1I, 4f, g

Type locality. Peru, Junín, Chanchamayo.

Distribution. Previously known from Ecuador and Peru (Gielis, 2011), the species is reported from Colombia for the first time.

Remarks. Five males were collected in the vicinity of the Refugio of Parque Natural Chicaque. The female is still unknown.

***Hellinsia* sp.**

Figs. 1J, 5c

Remarks. Based on our knowledge of the described species of *Hellinsia*, this one appears to be undescribed. However, we refrain from describing it until more specimens, or at least one male can be collected as only the one female specimen figured here was collected in the vicinity of the Refugio of Parque Natural Chicaque. It is similar in color and forewing markings to *H. pallens* Gielis, 2011 and *H. scripta* Gielis, 1999, respectively described from Ecuador, Pichincha and Costa Rica, Puntarenas. However, both of these species are smaller with documented wingspans of 17 mm for *H. pallens* and 19-22 mm for *H. scripta* compared to the 26.5 mm of the Colombian species. On the forewing the blackish-brown markings of this species appear to be unique in the continuous blackish-brown oblique band from the base of the cleft to the costa. In the female genitalia, all three species have the ostium bursae positioned on the left and a funnel-shaped antrum, but differ in details of the length of the enlarged section of the ductus seminalis compared to the length of the corpus bursae.

***Adaina atahualpa* Gielis, 2011**

Adaina atahualpa Gielis, 2011: 707.

Figs. 1L, 5e

Type locality. Ecuador, Napo Province, 15 km SE Cosanga, Cocodrilo.

Distribution. Colombia, Ecuador (Gielis, 2011).

Remarks. Two females were collected at Chicaque in the vicinity of the Refugio.

***Adaina* sp.**

Figs. 2, 5f

Remarks. This species is slightly similar in habitus to *Adaina ambrosiae* (Murtfeldt, 1880) although the paler and darker stripes and spots, especially on the forewing lobes differ significantly. Also, the female genitalia of the unique specimen collected in the vicinity of Chicaque's Refugio differ strongly in the large, elongate, triangular, and thickly sclerotized tergite VIII, and other features of the genitalia as such (see Landry *et al.*, 2004). Although we suspect that it is new, we refrain from describing it pending the availability of male specimens.

DISCUSSION

Far from having been a true inventory of the Pterophoridae at Parque Natural Chicaque, the collecting efforts involved during three days and nights nevertheless allowed us to record a sample of 12 species, including two that could be identified as new, and three that may be new, but for which material is lacking to ascertain their taxonomic status.

Based on Holbern (2022) and the database of the second author, below we provide a complete checklist of the Pterophoridae of Colombia in phylogenetic order. Species marked with * are recorded here for the first time for Colombia.

Quadriptilia obscurodactyla Gielis, 1994
Platyptilia thyellopa Meyrick, 1926
Stenoptilia suprema Meyrick, 1926
Stenoptilia tenuis (Felder & Rogenhofer, 1875)
Amblyptilia scutellaris (Felder & Rogenhofer, 1875)
Anstenoptilia hugoiella Gielis, 1996
Stenoptilodes gilvicolor (Zeller, 1877)
Stenoptilodes posticus (Felder & Rogenhofer, 1875)
Stenoptilodes sordipennis (Zeller, 1877)
Stenoptilodes stigmatica (Felder & Rogenhofer, 1875)
Stenoptilodes thrasydoxa (Meyrick, 1926)
Stenoptilodes umbrigeralis (Walker, 1864)
****Postplatyptilia chicaquensis*** Landry & Gielis, **sp. n.**
Postplatyptilia fuscicornis (Zeller, 1877)
****Postplatyptilia parana*** Gielis, 1996
Postplatyptilia transversus Gielis, 2006
Lioptilodes albistriolatus (Zeller, 1877)
****Lioptilodes cocodrilo*** Gielis, 2006
Cnaemidophorus smithi Gielis, 1992
Exelastis pumilio (Zeller, 1873)
Michaelophorus nubilus (Felder & Rogenhofer, 1875)
Sphenarches languidus (Felder & Rogenhofer, 1875)
Megalorhipida leucodactylus (Fabricius, 1794)
Hellinsia argutus (Meyrick, 1926)
Hellinsia barbatus (Gielis, 1996)
Hellinsia bogotanus (Felder & Rogenhofer, 1875)
Hellinsia conjunctus (Zeller, 1877)
Hellinsia crescens (Meyrick, 1926)
****Hellinsia danielae*** Landry & Gielis, **sp. n.**
Hellinsia estrellae Gielis, 2014
Hellinsia fumiventris (Zeller, 1877)
Hellinsia fusciciliatus (Zeller, 1877)
Hellinsia lenis (Zeller, 1877)
Hellinsia monserrate Arenberger & Bond, 1995
Hellinsia nigricalcarius (Gielis, 1996)
Hellinsia nodipes (Zeller, 1877)
****Hellinsia obscuricilia*** Arenberger & Wojtusiak, 2001
Hellinsia ochracealis (Walker, 1864)
Hellinsia ochricostatus (Zeller, 1877)
****Hellinsia pelospilus*** (Zeller, 1877)
Hellinsia praenigratus (Meyrick, 1921)
Hellinsia schneblei Gielis, 2014
Hellinsia scribarius (Meyrick, 1926)
Hellinsia surinamensis (Sepp, 1855)
Oidaematophorus espeletiae Hernandez, Fuentes, Fajardo & Matthews, 2014
****Oidaematophorus pseudotrachyphloeus*** Gielis, 2011
Adaina atahualpa Gielis, 2011
Adaina desolata Arenberger & Bond, 1995
Emmelina buscki (Barnes & Lindsey, 1921)
Singularia brechlini Kovtunovich & Ustjuzhanin, 2016
Singularia guajiro Kovtunovich & Ustjuzhanin, 2016
Singularia lesya Kovtunovich & Ustjuzhanin, 2016
Singularia tolima Kovtunovich & Ustjuzhanin, 2016

In sum, with 53 species (56 if the three undetermined species treated here from Chicaque are included), the Pterophoridae fauna of Colombia is undoubtedly poorly known, given the 130 species recorded from neighboring Ecuador (Ustjuzhanin *et al.*, 2021). Of the 46 species previously mentioned in the literature from Colombia, 41 were described as new species with a type locality in Colombia. This high proportion suggests an equally high proportion of endemic species, a situation also observed in neighboring countries such as Ecuador and Peru. Much more field work is needed to provide a clearer picture of the diversity of the Colombian Pterophoridae, a situation that is undoubtedly similar in many other groups of micro-moths.

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