

**QUARANTINE DETECTION OF NEMATODES AND PROCEDURES
FOR THEIR ERADICATION FROM VEGETATIVELY PROPAGATED MATERIALS
IMPORTED BY BRAZIL**

R. C. V. Tenente, E. S. C. Manso, M. A. S. Mendes, A. S. dos A. Marques, and E. Figueira Filho
EMBRAPA/CENARGEN, Cx. P 2372, 70849-970, Brasilia, Brazil.

RESUMEN

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El Centro Nacional para las Investigaciones de los Recursos Genéticos y Biotecnología (CENARGEN) es responsable de la introducción, cuarentena de pos-entrada e intercambio de germoplasmas vegetales para los programas de investigaciones brasileños que incluyen instituciones de investigaciones y universidades. El Laboratorio de Nematología del CENARGEN tiene facilidades para la cuarentena de pos-entrada y la inspección de todos los germoplasmas propagativos vegetales. Resultados de la inspección entre 1981-94 son presentados teniendo en cuenta la incidencia, identificación y la irradiación de los nematodos asociados con materiales propagativos vegetales. Algunos de los mas importantes generos detectados durante este periodo fueron *Globodera*, *Ditylenchus*, *Pratylenchus* y *Xiphinema*. Aproximadamente el 64% de los germoplasmas vegetales infectados fueron satisfactoriamente saneados utilizando métodos de cultivo de tejidos. Algunos nematodos no fueron erradicados de aproximadamente el 24% de las asepsiones inrecladas las que fueron posteriormente incineradas. El resto de los germoplasmas infectados (aproximadamente el 12%) fue saneado mediante el tratamiento con agua caliente o con el tratamiento con el nematocida carbofuran.

Palabras clave: Brasil, cuarentena, nematodos erradicados.

The principal objective of the Centro Nacional de Recursos Genéticos e Biotecnologia (CENARGEN) is to provide the research system in Brazil with germplasm of sufficient variability for research programs. To accomplish its mandate, CENARGEN participates in botanic exploration to collect, classify and conserve germplasm, and is responsible for importation and introduction of exotic material. CENARGEN has imported germplasm into Brazil since its inception in 1974 and has experienced a significant increase in its accessions in the past several years.

While the importation of plant germplasm is a fundamental aspect of agricultural development in Brazil, the introduction of plant materials increases the risks of introducing exotic pathogens and pests at a rate

commensurate with the increasing accession levels. In response to those risks, CENARGEN established the Section of Plant Germplasm Exchange, with the purpose of safely managing plant importation through a process of inspection and quarantine.

Fewer accessions of vegetative propagative material are received by CENARGEN than seed; however, vegetative plant material has the capacity to carry more pathogens and pests than seed. Therefore, detection and eradication of pathogens and pests carried on vegetative propagative material imported into Brazil is considered to be a high priority objective.

The purpose of this communication is to review results of the plant inspection program during the period 1981 to 1994 with regard to the detection of plant para-

sitic nematodes in vegetative plant material. In addition, we describe the efficacy of three methods to eradicate nematodes from vegetative plant material prior to its release to plant breeders.

Several methods were used to detect nematodes in plant germplasm. Microscopic examination of plant tissue was used to detect symptoms (galls, lesions) or signs (females, egg masses) of sedentary plant-parasitic nematodes. Plant material also was processed ($20^{\circ}\text{C} \pm 2^{\circ}\text{C}$) using a modified tray technique to recover motile nematodes (Whitehead and Hemming, 1965). Our modification included the use of a smaller tray size ($53 \times 21 \times 5$ cm), and the use of one layer of plastic net (2-mm aperture) to support Kimwipe tissue. Distilled water was added in sufficient quantity to wet the tissue. After 4-6 hrs of extraction, the water was passed through a $25 \mu\text{m}$ sieve. The contents of the sieve were backwashed into a beaker prior to observation for the presence of nematodes.

To detect nematode cysts, pieces of propagating plant tissue were placed in a flat-bottomed glass flask with 500 ml of distilled water and automatically agitated for 10 min. A sucrose solution (400 g/L water + 12.5 ppm of Separan) was added, and the flask was agitated for an additional 10 min. The suspension was poured into a flask and left for 10 min before passing 1/3 of the suspension through a Whatman filter paper. The material that was retained on the filter paper was examined with a dissecting microscope (Carvalho *et al.*, 1953).

Generally, we attempted to eradicate nematodes using heat or carbofuran. If those methods were unsuccessful, meristem cultures were tried. Vegetative propagation tissue was treated with hot water (52°C) for 20 min. Bulbs were soaked for 15 min in a solution (2.4 g a.i./L) of carbofuran (Tenente *et al.*, 1986 and 1994) and then planted in sterilized soil and main-

tained in the greenhouse until new bulbs were produced and inspected. However, meristem tissue culturing was the most successful technique for nematode eradication. Meristems were grown ($20\text{-}22^{\circ}\text{C}$) in specific media in an incubator with 12 hr of light. Plantlets were adapted to soil under greenhouse conditions at $18\text{-}25^{\circ}\text{C}$. Plants successfully generated with this technique were checked for nematodes, using the previously described methods, prior to release.

Most nematodes were recovered as juveniles, thereby preventing identification to species (Table 1). For crops such as potato, garlic, peanut and grape, the most economically important genera detected were *Globodera*, *Xiphinema*, *Meloidogyne* and *Ditylenchus*. These and previous results (Tenente, 1985; Tenente *et al.*, 1993) clearly indicate the need for an effective inspection and quarantine service to mitigate the risks of introducing and disseminating exotic pests in the imported plant germplasm (Tenente and Manso, 1987).

The hot water treatment eradicated *Aphelenchoides*, *Cryptaphelenchus*, *Helicotylenchus*, *Pratylenchus* and *Tylenchus* from pineapple and *Aphelenchoides* from mango (Table 1). *Aphelenchoides* and *Ditylenchus dipsaci* were eradicated from infested garlic bulbs by immersion in carbofuran solution. Nematodes were eradicated and plants regenerated from approximately 64% of the infested accessions using meristem tissue culture. Although nematodes were eradicated from tissues of *Arachis hypogaea*, *Amorphophalus* sp., *Humulus* spp. and *Tulipa generiana*, the meristem-derived plantlets did not survive transplantation to the greenhouse. Our nematode eradication methods were ineffective for approximately 24% of the infested germplasm accessions which included *Acassia*, *Agave*, *Brownea*, *Eriobotrya*, *Humulus*, *Lignum*, *Teconua*,

Table 1. Nematodes detected in vegetative propagating material imported into Brazil (1981-1994).

Plant germplasm	Origin	Detected nematodes	Eradication techniques
<i>Acassia cyanophylla</i>	Mexico	<i>Aphelenchus</i> sp.	Not successful ¹
		<i>Ogma</i> sp.	" "
<i>Agave sisalana</i>	Tanzania	<i>Aphelenchoides</i> sp.	Not successful
		<i>Tylenchus</i> sp.	" "
		<i>Xiphinema</i> sp.	" "
<i>Allium sativum</i>	Italy	<i>Aphelenchoides</i> sp.	Tissue culture
		<i>Tylenchus</i> sp.	" "
	Peru	<i>Aphelenchoides</i> sp.	Tissue culture
		<i>Pratylenchus</i> sp.	" "
	Chile	<i>Ditylenchus dipsaci</i>	Tissue culture
USA	<i>Aphelenchoides</i> sp.	Carbofuran 35%	
		<i>Ditylenchus dipsaci</i>	" "
<i>Amorphophalus</i> sp.	Indonesia	<i>Brachydorus</i> sp.	Tissue culture
<i>Ananas comosus</i>	France	<i>Aphelenchoides</i> sp.	52°C/20 min
		<i>Helicotylenchus</i> sp.	" "
		<i>Pratylenchus</i> sp.	" "
	Martinique	<i>Aphelenchoides</i> sp.	52°C/20 min
		<i>Cryptaphelenchus</i> sp.	" "
		<i>Pratylenchus</i> sp.	" "
		<i>Tylenchus</i> sp.	" "
<i>Arachis hypogaea</i>	Colombia	<i>Aphelenchoides</i> sp.	Tissue culture
		<i>Ditylenchus</i> sp.	" "
		<i>Tylenchus</i> sp.	" "
<i>Arracacia xanthorrhiza</i>	Ecuador	<i>Aphelenchoides</i> sp.	Tissue culture
<i>Brachiaria brizantha</i>	USA	<i>Ditylenchus</i> sp.	Tissue culture
<i>Brownea latifolia</i>	Trinidad and Tobago	<i>Tylenchus</i> sp.	Not successful
<i>Eriobotrya japonica</i>	Israel	<i>Aphelenchoides</i> sp.	Not successful
		<i>Nothotylenchus</i> sp.	" "
<i>Humulus</i> spp.	USA	<i>Dorylaimus</i> sp.	Not successful
		<i>Tylenchus</i> sp.	" "
<i>Hyacinthus orientalis</i>	USA	<i>Aphelenchoides</i> sp.	Tissue culture
<i>Hypahrenia</i> spp.	Colombia	<i>Aphelenchoides</i> sp.	Tissue culture
<i>Lignum vitae</i>	Trinidad and Tobago	<i>Aphelenchus</i> sp.	Not successful
		<i>Dorylaimus</i> sp.	" "

¹Not successful = Nematodes not eradicated and materials were incinerated.

Table 1. (Continued) Nematodes detected in vegetative propagating material imported into Brazil (1981-1994).

Plant germplasm	Origin	Detected nematodes	Eradication techniques
		<i>Helicotylenchus</i> sp.	" "
		<i>Tylenchus</i> sp.	" "
<i>Mangifera indica</i>	USA	<i>Aphelenchoides</i> sp.	52°C/20 min
<i>Mentha</i> sp.	Peru	<i>Aphelenchoides</i> sp.	Tissue culture
<i>Mentha villosa</i>	Peru	<i>Pratylenchus</i> sp.	Tissue culture
<i>Musa</i> sp.	Ecuador	<i>Aphelenchoides</i> sp.	Tissue culture
		<i>Helicotylenchus</i> sp.	" "
	Martinique	<i>Ditylenchus</i> sp.	Tissue culture
	Phillippines	<i>Meloidogyne</i> sp.	Tissue culture
		<i>Xiphinema</i> sp.	" "
<i>Narcissus</i> sp.	USA	<i>Aphelenchoides</i> sp.	Tissue culture
<i>Opuntia ficus-indica</i>	Mexico	<i>Dorylaimus</i> sp.	Tissue culture
		<i>Tylenchus</i> sp.	" "
<i>Oreganum vulgare</i>	Peru	<i>Aphelenchoides</i> sp.	Tissue culture
<i>Pinus</i> spp.	Mexico	<i>Criconemella</i> sp.	Tissue culture
		<i>Tylenchus</i> sp.	" "
<i>Piper wichmanii</i>	USA	<i>Ditylenchus</i> sp.	Tissue culture
<i>Prosopis</i> sp.	USA	<i>Aphelenchoides</i> sp.	Tissue culture
<i>Saccharum officinallis</i>	USA	<i>Paurodontus gracilis</i>	Tissue culture
<i>Solanum</i> spp.	Holland	<i>Aphelenchus</i> sp.	Tissue culture
		<i>Aphelenchoides</i> sp.	" "
		<i>Globodera</i> sp.	" "
	USA	<i>Aphelenchoides</i> sp.	Tissue culture
		<i>Aphelenchus</i> sp.	" "
		<i>Ditylenchus</i> sp.	" "
		<i>Meloidogyne</i> sp.	" "
	Canada	<i>Ditylenchus dipsaci</i>	Tissue culture
	Peru	<i>Ditylenchus</i> sp.	Tissue culture
		<i>Tylenchus</i> sp.	" "
<i>Teconua</i> sp.	Trinidad and Tobago	<i>Tylenchus</i> sp.	Not successful
<i>Thaumatococcus danielli</i>	Ghana	<i>Aphelenchoides</i> sp.	Not successful
		<i>Ditylenchus</i> sp.	" "
		<i>Tylenchus</i> sp.	" "
<i>Tulipa generiana</i>	USA	<i>Aphelenchoides</i> sp.	Tissue culture

*Not successful = Nematodes not eradicated and materials were incinerated.

Table 1. (Continued) Nematodes detected in vegetative propagating material imported into Brazil (1981-1994).

Plant germplasm	Origin	Detected nematodes	Eradication techniques
<i>Vitis</i> spp.	France	<i>Aphelenchoides bicaudatus</i>	Tissue culture
		<i>Ditylenchus</i> sp.	" "
		<i>Trichodorus</i> sp.	" "
	Africa	<i>Dorylaimida</i> (Order)	Tissue culture
	USA	<i>Aphelenchoides</i> sp.	Tissue culture
		<i>Ditylenchus</i> sp.	" "
<i>Xiphinema</i> sp.		" "	
<i>Zeniber</i> sp.	Italy	<i>Aphelenchoides</i> sp.	Not successful
<i>Xanthosoma sagihifolium</i>	Venezuela	<i>Aphelenchoides</i> sp.	Tissue culture
		<i>Ditylenchus</i> sp.	" "

*Not successful = Nematodes not eradicated and materials were incinerated.

Thaumatooccus, and *Zeniber*. All remaining tissues of the latter genera were incinerated.

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