EFFECT OF INITIAL POPULATION DENSITY OF MELOIDOGYNE INCOGNITA RACE 3 ON THE GROWTH OF KENAF (HIBISCUS CANNABINUS L.)[†]

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ABSTRACT

Barillas, J. R., G. W. Lawrence, and K. S. McLean. 1993. Effect of initial population density of *Meloidogyne incognita* race 3 on the growth of kenaf (*Hibiscus cannabinus*). Nematropica 23:15–19.

In a greenhouse experiment, seedlings of kenaf (*Hibiscus cannabinus*) cv. Tainung I were inoculated with *Meloidogyne incognita* race 3 at initial population densities (Pi) of 0, 100, 500, 1000, 2500, and 5000 eggs + second-stage juveniles/500 cm³ soil. After 60 days, plant height, stalk diameter, and root and stem weights were significantly reduced at Pi = 5000 but not at lower Pi's. Shoot dry weights were reduced 46% compared with the untreated control. The total nematode population (eggs and second-stage juveniles recovered from roots and soil) increased with increasing inoculum levels and the reproductive factor (R = final population/initial population) decreased as the initial inoculum level increased.

Key words: Hibiscus cannabinus, inoculum, kenaf, Meloidogyne incognita, nematode reproduction, population dynamics, southern root-knot nematode.

RESUMEN

Barillas, J. R., G. W. Lawrence y K. S. McLean. 1993. Efecto de la densidad inicial de la población de *Meloidogyne incognita* raza 3 sobre el crecimiento de kenaf (*Hibiscus cannabinus*). Nematrópica 23:15–19.

En un experimento de invernadero, se inocularon plántulas de kenaf (*Hisbiscus cannabinus* cv. Tainung I) con poblaciones iniciales (Pi) de 0, 100, 500, 1 000, 2 500 y 5 000 huevos y segundos estadíos juveniles de *Meloidogyne incognita* raza 1 por 500 cm³ de suelo. A los 60 días después de la inoculación, el nematodo redujo significativamente la altura de la planta, el diámetro del tallo y los pesos del tallo y las raíces con Pi de 5 000, pero no en aquellas plantas con Pi menores. El peso del follaje se redujo en un 4% en relación con el testigo. La población total de nematodos (huevos y segundos estadíos juveniles recuperados de las raíces y del suelo) aumentó de acuerdo con la densidad inicial, mientras que la tasa de incremento poblacional (Pf/Pi) disminuyó.

Palabras clave: dinámica de poblaciones, Hibiscus cannabinus, kenaf, Meloidogyne incognita, nematodo agallador, reproducción del nematodo.

INTRODUCTION

Kenaf (*Hibiscus cannabinus* L.) is a short-day, annual herbaceous plant, originally cultivated for the soft bast fiber in the stem. It is a member of the Malvaceae family and is closely related to okra, jute, and cotton.

Kenaf is believed to have originated in Africa, where *H. cannabinus* and closely related species are found growing in many countries (6). Although kenaf has been cultivated for several thousand years, it was not grown commercially for fiber extraction until around 1900. Kenaf

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is used for cordage, fishing nets, floor matting, backing for rugs, sacking materials, twine, ropes, and cables (5,8).

In 1952, the U.S. Department of Agriculture cited kenaf as a source of paper pulp in India (13). In 1962, Clark *et al.* examined blending kenaf with wood pulp. The resulting blend produced a paper with a desirable stensile strength, burst strength, and brightness (4).

A potential limitation to the successful cultivation of kenaf in the United States is its susceptibility to *Meloidogyne incognita* Kofoid & White (Chitwood) (17). The pathogenicity of *M. incognita* to kenaf was first described in Florida in 1944 (15). More recently, kenaf was found to be a host for populations of *M. incognita* representing the four host races (16).

Meloidogyne incognita is the predominant species of root-knot nematode in the southern United States and it is a major pathogen of cotton and soybean (7). This nematode has the potential to become a serious problem in the cultivation of kenaf in Mississippi since both soybean and cotton are widely grown and are hosts.

Although some research has been conducted toward determining damage thresholds for *Hoplolaimus aegypti* on kenaf fiber yield (9), the effect of *M. incognita* on kenaf yield has received little attention (1,10,11,12). The objective of this research was to examine the relationship between the initial population density of *M. incognita* race 3 and the growth of kenaf under greenhouse conditions. Net nematode reproduction was also measured.

MATERIALS AND METHODS

The experiment was conducted in the Plant Pathology/Nematology greenhouse at Mississippi State University during 1990.

A population of *M. incognita* (race 3) was obtained from infected cotton plants (*Gossypium hirsutum*) and increased on tomato (*Lycopersicon esculentum*). Eggs and second-stage juveniles (J2) were extracted by cutting the roots into 0.5-cm pieces and placing them in 0.525% sodium hypochlorite for 4 min (3). The resulting suspension was passed through a 50-μmpore sieve (200 mesh) nested over a 28-μm-pore sieve (500 mesh). Eggs and J2 were rinsed on the 28-μm-pore sieve and collected for use as inoculum.

Seeds of kenaf cv. Tainung 1 were germinated for 72 hr and transplanted into 12-cm-diam clay pots containing a 1:1 mixture (v:v) of steam-sterilized sand and soil. *Meloidogyne incognita* eggs and J2 were delivered in 10 ml total volume of water into two 1.5-cm-diam × 3-cm-deep depressions in each pot. Treatments consisted of initial population densities (Pi) of 0, 100, 500, 1 000, 2 500, and 5 000 eggs + J2/500 cm³ soil.

Treatments were arranged in a randomized complete block design with eight replications. Plants were grown in the greenhouse at a temperature ranging from 25 to 32 °C and were harvested after 60 days. The experiment was repeated.

At harvest, plant height, stem diameter, and weights of shoots and roots were recorded. Second-stage juveniles were extracted from soil by gravity screening and sucrose centrifugation (3). The eggs in a 1-g random sample of roots were extracted by the sodium hypochlorite method and counted (3). Root galling was rated using a 0 to 5 scale, where 0 = no galling, 1 = 1-2 galls, 2 = 3-10 galls, 3 = 11-30 galls, 4 = 31-100 galls, and 5 = more than 100 galls per root system (14). Data were analyzed and means were compared using Fisher's protected least significant difference test. Quadratic equations

were fit by least squares to relationships between initial nematode densities and plant growth (as measured by stem weights and heights).

RESULTS AND DISCUSSION

Results of the two replicated experiments were similar. The data presented are from the first run.

Effects on kenaf: At harvest, plant height in untreated pots averaged 84.5 cm. All plant growth parameters were significantly reduced in pots inoculated with 5 000 eggs + J2/500 cm³ soil when compared with the untreated control. Plant heights were reduced 24.6% and stalk diameter at the base of the plant was reduced 21.7% (Table 1). Fresh and dry shoot weights in the same pots were reduced 48% and 46%, respectively. The quadratic models that best described the relationships between Pi and plant growth parameters (plant height, stem diameter, and shoot weight) suggested that growth may have been stimulated at low inoculum densities. However, growth stimulation was not observed in the second run of the test.

Fresh root weights of inoculated plants were consistently increased when compared with the untreated plants, and dry root weights were consistently reduced. Dry root weights were significantly reduced even at Pi = 100 and 1000 (Table 1). The increase in fresh root weight in the infected plants must be due to increased water content. This phenomenon has been reported for roots of various other plants infected by root-knot nematodes.

Nematode reproduction: As initial inoculum levels increased, M. incognita population densities at harvest (Pf) also increased. The Pf ranged from $18\,400$ at Pi = 100to 148700 at Pi = 5000 (Table 2). The reproductive factor (R = final population/initial population) decreased from 184.4 at Pi = 100 to 29.7 at Pi = 5000(Table 2). The number of egg masses recovered per gram of root increased as initial inoculum levels increased, from 1.0 at Pi = 100 to 16.6 at Pi = 5000 (Table 2). The average number of eggs per egg mass at Pi = 5000 (733) was about half that at Pi = 2500 (1320) (Table 2), suggesting a crowding effect due to the increased number of galls in the roots. All

Table 1. Effect of *Meloidogyne incognita* race 3 on plant growth of kenaf cv. Tainung 1 in greenhouse pots, 60 days after inoculation.

Inoculum density (Pi) ^z	Plant height (cm)	Basal stem diameter (mm)	Shoot weight (g)		Root weight (g)	
			Fresh	Dry	Fresh	Dry
0	84.5	6.0	31.8	3.9	14.9	1.99
100	95.4	5.9	34.8	4.4	25.2	0.76
500	91.9	6.0	33.9	4.0	26.1	0.86
1 000	97.1	6.3	35.1	4.1	22.6	0.75
2 500	89.1	6.3	33.7	4.0	34.7	1.64
5 000	63.7	4.7	16.5	2.1	19.4	0.93
$\overline{\text{FLSD}} (P \le 0.5)$	14.7	0.8	8.1	1.2	10.3	1.2

Data are means of eight replications. Means compared using Fisher's protected least significant difference test.

²Eggs + second-stage juveniles/500 cm³ soil.

Inoculum density (Pi) ^w	Final population (Pf)*	Reproductive factor (R) ^y	Root gall index ^z	Egg masses/g of root	Eggs/egg mass
0	0	0	0.0	0.0	0
100	18 442	184.4	4.8	1.0	1 079
500	45 943	91.5	4.8	1.6	1 135
1 000	69 717	69.7	4.9	3.3	1 018
2 500	111 212	44.5	4.9	8.5	1 322
5 000	148 739	29.7	5.0	16.6	733
$\overline{\text{FLSD}} (P \le 0.05)$	44 364	***************************************	0.3	5.7	615

Table 2. Reproductive parameters of *Meloidogyne incognita* on kenaf cv. Tainung 1 in greenhouse pots, 60 days after inoculation.

Data are means of eight replications. Means compared using Fisher's protected least significant difference test.

initial population densities (Pi) resulted in a root galling index greater than 4 (Table 2). This high root gall index and the high reproductive factor measured at all inoculum levels confirm that Tainung 1 is a highly suitable host for *M. incognita* (14).

In this 60-day greenhouse study, kenaf plant growth parameters were consistently reduced only at Pi = 5 000 M. incognita eggs + J2/500 cm³ soil. Lower initial population densities did not reduce plant growth but they increased to high levels (18 000–150 000 nematodes/500 cm³ soil) greatly exceeding the highest Pi levels tested, and may have reduced plant growth if the experiments had been extended beyond 60 days. Moreover, relative host sensitivity of a plant characterized under greenhouse conditions, where there is a minimal stress, frequently differs from plant growth responses under field conditions (2). Therefore, additional nematode population development studies on kenaf grown under field conditions are needed to determine damage thresholds for economic injury.

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LITERATURE CITED

- BARILLAS, J. R., G. W. LAWRENCE, and K. S. McLEAN. 1991. Effect of initial population densities of the root-knot nematode Meloidogyne incognita on kenaf. Pp. 1-2 in Proceedings, Third Annual International Kenaf Association Conference, February 28-March 2, 1991, Tulsa, Oklahoma, U.S.A.
- BARKER, K. R., and J. P. NOE. 1987. Establishing and using threshold population levels. Pp. 75–81 in J. A. Veech and D. W. Dickson, eds. Vistas on Nematology: A Commemoration of the Twenty-fifth Anniversary of the Society of Nematologists. Society of Nematologists, Inc: Hyattsville, Maryland, U.S.A.
- BARKER, K. R. 1985. Nematode extraction and bioassays. Pp. 20–35 in K. R. Barker, C. C. Carter, and J. N. Sasser, eds. An Advanced Treatise on Meloidogyne, Vol. II. Methodology. Department of Plant Pathology, North

^wEggs + second-stage juveniles/500 cm³ soil.

^{*}Eggs in roots + juveniles recovered from 500 cm³ soil.

 $^{{}^{}y}R$ = Final population/Initial population.

²Root galling was rated on a 0-5 scale where 0 = no galling, 1 = 1-2 galls, 2 = 3-10 galls, 3 = 11-30 galls, 4 = 31-100 galls, and 5 = more than 100 galls per root system.

- Carolina State University, and the U.S. Agency for International Development, North Carolina State University Graphics, Raleigh, North Carolina.
- 4. CLARK, T. F., G. H. NELSON, H. J. NIE-SCHLAG, and I. A. WOLF. 1962. A research for new fiber crops. V. Pulping studies on kenaf. Journal of the Technical Association of the Pulp and Paper Industry 45:780-786.
- CRANE, D. C. 1947. Kenaf-fiber plant rival of jute. Economical Botany 1:334-350.
- 6. DEMPSEY, S. M. 1975. Fiber Crops. The University Press of Florida, Gainesville, Florida, U.S.A. 457 pp.
- 7. FASSULIOTIS, G. 1980. Plant resistance to root-knot nematodes, Pp. 33-49 in R. D. Riggs, ed. Nematology in the Southern Region of the United States. Southern Cooperative Series Bulletin 276. Arkansas Agricultural Experiment Station, University of Arkansas, Fayetteville, Arkansas, U.S.A.
- 8. FULLER, M. J. 1990. The potential for kenaf in Mississippi: an economic prospective. Pp. 1-2 in Proceedings, Second Annual International Kenaf Association Conference, March 1-3, 1990, Tulsa, Oklahoma, U.S.A.
- 9. HAMIDA, A. D., and K. K. FAIKA. 1985. Effect of population density of Hoplolaimus aegypti on kenaf yield and nematode reproduction. Annals of Agricultural Science 30:649-
- 10. LAWRENCE, G. W., and K. S. McLEAN. 1990. Host suitability of kenaf to three plant parasitic nematodes. Phytopathology 80:436. (abstract)
- 11. LAWRENCE, G. W., K. S. McLEAN, B. S. BRASHER, S. W. NEILL, and M. J. FULLER.

- 1990. Association of plant parasitic nematodes with kenaf in Mississippi. P. 4 in Proceedings, Mississippi Association of Plant Pathologists and Nematologists, March 6-7, 1990, Mississippi State University, Mississippi State, Mississippi, U.S.A. 9:4.
- 12. LAWRENCE, G. W., and K. S. McLEAN. 1991. Reproductive potential of plant parasitic nematodes on kenaf. Mississippi Agricultural and Forestry Experiment Station Research Report, Vol. 16, No. 2. Mississippi State University, Mississippi State, Mississippi, U.S.A.
- McCAM, L. P. 1952. Kenaf (Hibiscus cannabinus), a bibliography survey. U.S. Department of Agriculture Bibliographical Bulletin No. 17. U.S. Government Printing Office, Washington, D.C. 20 p.
- 14. TAYLOR, A. L., and J. N. SASSER. 1978. Biology, Identification and Control of Rootknot Nematodes (Meloidogyne Species). International Meloidogyne Project. Department of Plant Pathology, North Carolina State University, and the U.S. Agency for International Development. North Carolina State University Graphics, Raleigh, North Carolina. 111 p.
- 15. THAMES, W. H. Jr. 1952. Preliminary reports of some of the disease and pest problems of kenaf, Hibiscus cannabinus L. in South Florida. Plant Disease Reporter 36:125.
- 16. VEECH, J. A. 1990. Nematode parasitism of kenaf. P. 5 in Proceedings, Second Annual International Kenaf Association Conference, March 1-3, 1990, Tulsa, Oklahoma, U.S.A.
- 17. WILSON, F. D., and T. E. SUMMERS. 1966. Reaction of kenaf, roselle and related species of Hibiscus to root-knot nematodes. Phytopathology 56:687-690.

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