# EFFECTS OF MELOIDOGYNE INCOGNITA ON GROWTH AND NUTRIENT CONTENT OF AMARANTHUS VIRIDIS AND TWO CULTIVARS OF HIBISCUS SABDARIFFA

Terry-Ann P. Heffes,<sup>1</sup> Phyllis L. Coates-Beckford,<sup>1</sup> and Hilary Robotham<sup>2</sup>

Department of Botany<sup>1</sup> and Center for Nuclear Sciences<sup>2</sup>, University of the West Indies, Mona Campus, Kingston 7, Jamaica.

Accepted:

19.VI.1990

Aceptado:

#### ABSTRACT

Heffes, T. P., P. L. Coates-Beckford, and H. Robotham. 1991. Effects of *Meloidogyne incognita* on growth and nutrient content of *Amaranthus viridis* and two cultivars of *Hibiscus sabdariffa*. Nematrópica 21:7–18.

Pots with seedlings of callaloo (Amaranthus viridis) and the 'Red' and 'White' cultivars of sorrel (Hibiscus sabdariffa) were inoculated with 0, 100, 1000, and 10000 eggs of Meloidogyne incognita race 1 and the plants were grown for 6 weeks in a greenhouse. Nematode reproduction was good on Red sorrel, poor on callaloo, and was inhibited by White sorrel. Dry weights of infested callaloo roots were greater than weights of noninfested roots. Shoot heights, leaf areas, and dry shoot and root weights of infested plants of both sorrel cultivars were less than those of noninfested plants. The concentrations of the nutrients varied with the host, plant organ, and initial nematode density. The shoot/root ratios of the elements were similar at all inoculum levels (Pi) except those of callaloo roots at Pi = 10000, which had significantly lower and higher ratios of sodium and manganese, respectively, than those of noninfested plants.

Key words: Amaranthus viridis, callaloo, Hibiscus sabdariffa cvs. White and Red, host resistance, macronutrients, Meloidogyne incognita race 1, micronutrients, pathogenicity, sorrel.

## RESUMEN

Heffes, T. P., P. L. Coates-Beckford, y H. Robotham. 1991. Efectos de *Meloidogyne incognita* sobre el crecimiento y contenido de nutrientes en *Amaranthus viridis* y dos cultivares de *Hibiscus sabdariffa*. Nematrópica 21:7–18.

Plántulas de amaranto (Amaranthus viridis) y los cultivares de sorrel (Hibiscus sabdariffa) 'Red' y 'White' fueron sembradas en maceteros e inoculados con 0, 100, 1 000, y 10 000 huevos de Meloidogyne incognita raza 1 y mantenidas por 6 semanas en un invernadero. El nematodo se reprodujo bien en hibisco Red, pobremente en amaranto, y fue inhibido por el hibisco White. Pesos secos de raíces infestadas de amaranto fueron mayores que los no inoculados. La altura de los tallos, superficie foliar, y peso seco de los tallos y pesos radiculares de plantas infestadas de ambos cultivares de hibisco fueron más bajos que los no inoculados. La concentración de nutrientes varió con el hospedador, parte de la planta muestreada, y el inóculo inicial. Las relaciones tallo-raíz de los elementos fueron similares en todos, excepto en aquellas raíces de amaranto con una Pi = 10 000 que presentaron niveles de sodio y magnesio significativamente bajos y altos, respectivamente en relación a quellas plantas no inoculadas.

Palabras clave: amaranto, Amarantus viridis, Hibiscus sabdariffa, hibisco, macroelementos, Meloidogyne incognita raza 1, microelementos, patogenicidad, resistencia.

### INTRODUCTION

The root-knot nematode, Meloidogyne incognita (Kofoid & White) Chitwood, is one of the most important pests in Jamaica. Earlier work by Hutton et al. (11) showed that callaloo (Amaranthus viridis L.) and sorrel (Hibiscus sabdariffa L. cv. White) inhibited the reproduction of Meloidogyne spp. in field and greenhouse tests whereas populations increased on 'Red' sorrel. The following studies were designed to investigate, in greater detail, the relationships between these three cultivars and M. incognita race 1. They also included investigations of the effects of parasitism on the nutrient status of hosts that show varying degrees of susceptibility to the nematode.

## MATERIALS AND METHODS

Populations of *M. incognita*, each initiated from a single egg mass, were identified as race 1 using the North Carolina Differential Host Test (9) and maintained on *Lycopersicon esculentum* Mill. cv. Rutgers in the greenhouse. Eggs for inoculum were obtained by the method of McClure et al. (16).

For White and Red sorrel, two seeds were sown 1.5 cm deep and. for callaloo, two seedlings were transplanted into 12.5-cm-diam plastic pots containing 1 L of steam-sterilized sand. All pots were thinned to leave one seedling per pot just before inoculation of the soil with eggs. Ten pots with 33-day-old callaloo and 10 with 18-day-old White or Red sorrel seedlings were inoculated with 100, 1 000, or 10 000 eggs of M. incognita race 1 by pouring 20-ml suspensions in preformed 2-cm-deep cavities adjacent to the seedlings. Ten pots of each cultivar, serving as the controls, were inoculated with supernatant from centrifuged egg suspensions. The cavities were filled with sterile sand. For each test host, the pots were separated randomly into a group with six and another with four of each initial nematode population (Pi). Both groups of each host were arranged in a completely randomized design in a greenhouse with a mean daytime temperature of 28 C. At the time of inoculation, plant heights were recorded and each pot was given 25 ml of Long Ashton nutrient solution (10). Plants were fertilized similarly once each week thereafter.

The plants were harvested at 6 weeks after inoculation. For each plant in those groups with six replicates of each treatment, records were made of the number of leaves produced, total leaf area, shoot height, dry shoot and root weights after drying at 70 C for 5 days, and the number of root galls present. The dried organs were kept for nutrient determinations. The soil in each pot was suspended in 6 L water and the nematodes were extracted from 3 L for 24 hours by the Christie and

Perry technique (3). The nematode extracts were made up to 100 ml with tap water and the populations were estimated from either three or five 1-ml aliquots for high (≥10/ml) and low (<10/ml) population densities, respectively, using Peter's counting slide. The number of egg masses produced by the roots of each plant and those groups with four replicates of each treatment were counted after staining with Phloxine B (4). Egg mass and gall production were rated by the indices 0, 1, 2, 3, 4, and 5 representing 0, 1-2, 3-10, 11-30, 31-100, and >100 egg masses or galls per root system, respectively.

Nutrient analyses were performed on the organs of each of the six replicates of callaloo. For each sorrel cultivar, the determinations were made on the combined pairs of randomly selected shoots or roots of each treatment. The dried organs of the replicates were ground in a stainless steel mill fitted with a 60-mesh sieve to retain the unwanted fragments. Samples of 200 mg of each organ were acid-digested by Wolf's method (24) modified by Rees and Sidrak (20) and made up to 100 ml with distilled water when cool. Nitrogen concentrations in callaloo were determined as nitrates by Taras' method (23) at a wave length of 470 nm, 30 min after addition of the reagents. Nitrogen in sorrel was determined as ammonium by the method of Fawcett and Stoughton (7) at 450 nm. Phosphorus concentrations were determined from the acid-digested samples by the method of Rees and Sidrak (2). For both elements, the standards ranged from 0 to 50 ppm.

The concentrations of the other essential macroelements (potassium, magnesium and calcium) and seven microelements (manganese, sodium, cobalt, iron, zinc, aluminum, and vanadium) were determined for each plant speices by instrumental neutron activation analysis (1). Each finely ground, homogenized, plant sample was enclosed in a heat-sealed, 1.7-cc polyethylene capsule. Then the samples were irradiated for 1 min at a neutron flux of 10<sup>12</sup> neutrons cm<sup>-2</sup> sec<sup>-1</sup> in a SLOWPOKE-II reactor (1), and allowed to decay for 10 min before counting for 5 min on a Canberra reverse electrode germanium semi-conductor detector. The samples were irradiated again for 2 hours, allowed to decay for 3 weeks, and counted for 3 hours. The resulting spectra were analyzed using the EG & G Ortec Geligam software package and the concentration of each element determined by comparison with similarly treated standards. All elements were quantified from the first irradiation, decay and count scheme except cobalt, iron, and zinc, which were quantified from the second.

All data were subjected to analyses of variance (22) and each treatment mean was compared to that of the control by the least significant difference test (LSD) to determine the significant effects of nematode infestation on host growth and nutrient content.

## RESULTS

Population development: Egg masses developed on roots of all inoculated plants except those of White sorrel growing in pots with Pi = 100 (Table 1). At Pi = 10 000, the respective egg mass indices for callaloo and White and Red sorrel were 4, 2, and 4 and were greater than those for plants grown at lower Pi s. Whereas the egg masses on Red sorrel were very large, those on White sorrel and callaloo were quite small with few eggs. At each Pi, the numbers of nematodes recovered were smallest from pots with White sorrel and were largest from those with Red sorrel.

Nematode pathogenicity: Leaf number, leaf area, shoot lengths, and shoot weights of infested callaloo plants did not differ significantly from those of the controls but the roots had significantly larger dry weights at all Pi s (Table 1). Both sorrel cultivars growing in soil with Pi =  $10\,000$  had significantly shorter shoots, smaller leaf areas, and lower shoot and root dry weights than the controls. Some adverse effects on the growth of each sorrel cultivar were observed at Pi =  $1\,000$  also but not at Pi = 100 (Table 1).

Galls, occurring on the feeder roots only of callaloo and White sorrel, never exceeded 3 mm in length but were ubiquitous and larger on Red sorrel roots. Several formed on callaloo and Red sorrel but few occured on White sorrel roots (Table 1).

Nutrient content: At Pi = 1 000 and 10 000, callaloo shoots had higher concentrations of nitrogen and manganese and, at the highest Pi, shoots had higher concentrations of aluminum than the controls (Table 2). In contrast, there was a significantly lower concentration of sodium at the two higher Pi s and of phosphorus and iron at the highest Pi.

In callaloo roots from pots with  $Pi = 1\,000$  and  $10\,000$ , nitrogen, calcium, magnesium, sodium, and aluminum occurred in higher concentrations than in the controls (Table 2). Nitrogen was also higher at Pi = 100. As in the shoots, phosphorus and iron concentrations were lower at  $Pi = 10\,000$  than at Pi = 0.

In White sorrel shoots of infested plants, the concentrations of only four elements differed from those of the controls. At  $Pi=1\,000$  and  $10\,000$ , nitrogen, manganese, and vanadium concentrations were higher and, at Pi=100, those of manganese and sodium were higher than control concentrations (Table 3).

At all Pi s, White sorrel roots had higher concentrations of nitrogen and manganese than the controls; roots from pots with Pi =  $10\,000$  had higher levels of magnesium, zinc, and vanadium; at Pi =  $1\,000$  and 100, concentrations of calcium were higher also (Tables 3). There were lower levels of potassium in the roots at Pi =  $1\,000$  and  $10\,000$  than at Pi =  $0\,00$  and lower sodium levels at Pi =  $10\,000$ . Nematode infestation did not

Table 1. Population development of Meloidogyne incognita race 1 and response of Amaranthus viridis and two cultivars of Hibiscus sabdariffa grown in soil infested for 6 weeks, at four initial nematode densities.

0 50** 350**	0 2* 4**	A. viridis 25			wt (mg)	wt (mg)	
0 50** 350**	0 2* 4**	25	dis				
50**	5* 4* 4*		11	291	2 544	1 160	0
350**	2* 4**	24	11	569	2 495	1453*	<b>5</b> **
	4**	24	12	276	2 862	1 580**	3**
3 600**		23	12	252	2 677	1 538**	4*
		H. sabdariffa cv. White	cv. White				
0	0	15	10	134	992	289	0
30**	0	15	11	156	773	305	0
20**	*	12	10	134	726	277*	8
330**	2**	11*	10	*86	503**	194**	*
		H. sabdariffa cv. Red	cv. Red				
0	0	25	11	116	869	154	0
150**	2**	20	10	120	775	167	*
1 250**	3**	22*	10	105	584*	168	3**
5 200**	4**	16**	10	*06	266*	*061	4**

Data are means of six replications except for egg mass and root-knot indices, which are means of four replications.

 $<sup>\</sup>sqrt[9]{\text{cale of } 0}$  - 5 where 0 = 0, 1 = 1 - 2, 2 = 3 - 10, 3 = 11 - 30, 4 = 31 - 100 egg masses or galls/root system. \*, \*\* = significantly different from the control at  $P \le 0.05$  and 0.01, respectively, by the LSD test.

<sup>&</sup>lt;sup>z</sup>Final height minus height at inoculation.

Table 2. Nutrient content of shoots and roots of Amaranthus wirdis from soil infested for 6 weeks with Meloidogme incognita race 1, at four initial nematode densities.

											Other micro-	nicro-
Initial number of		Essential macroelements (%)	nacroeleı	nents (%)		<b>—</b>	Ssential n	icroelem	Essential microelements (ppm)		elements (ppm)	(mdd)
eggs/pot	Z	Ъ	K	Ca	Mg	Na	Mn	3	Fe	Zn	Al	Λ
					Shoots							
0	0.30	1.20	4.5	1.30	0.90	355	56	0.5	287	35	100	0.7
100	0.35	1.00	4.5	2.20	1.20	388	56	1	1		109	9.0
1 000	0.67	0.90	4.3	1.80	1.10	214*	37*		ı		114	9.0
10 000	**08.0	0.67*	4.7	1.80	1.20	124**	40**	9.0	242*	39	136**	9.0
					Roots							
0	0.29	0.92	2.9	0.85	0.64	4 300	64	1.0	2 400	30	1 900	11.0
100	0.43**	0.83	3.3	0.89	0.64	4 400	54	•	ı	1	2 200	12.0
1 000	0.68**	0.77*	3.0	0.97*	0.84**	2 000	74	•			3 400	10.0
10 000	0.70	0.53**	3.0	1.02*	**88.0	7 700	54	1.0	2 000	33	3 600	11.0

\*, \*\* = significantly different from the control at  $P \le 0.05$  and 0.01, respectively, by the LSD test. Data are means of six replicates.

Table 3. Nutrient content of shoots and roots of Hibiscus sabdariffa cv. White from soil infested with Meloidogyne incognita race 1 for 6 weeks, at four initial nematode densities.

17:7:1		1.000	-	(10)		-					Other micro-	micro-
Initial number of		Essential	Essential macroelements (%)	ments (%)		7	Essentiai m	ncroelem	Essential microelements (ppm)		elements (ppm)	(mdd) s
eggs/pot	Z	Ь	K	Ca	Mg	Na	Mn	3	Fe	Zn	Al	>
					Shoots							
0	0:00	0.17	1.42	1.99	0.42	154	255	1	397	42	145	-
100	1.10	0.18	1.40	2.07	0.40	249**	311		,	1	150	_
1 000	1.34*	0.17	1.40	2.04	0.45	153	335			•	183	** 2**
10 000	1.86**	0.17	1.41	1.96	0.44	143	324*	_	403	47	153	** **
					Roots						į	
0	0.41	0.10	2.54	1.12	1.46	3 100	119	œ	1 600	51	166	20
100	0.61	0.11	2.54	1.26**	1.52	2 700	183*	•		ı	169	20
1 000	0.58*	0.12	2.30*	1.23**	1.51	2 900	201*	•	, 1	ı	170	25
10 000	1.01**	0.11	2.00*	1.12	1.66*	1 800	205*	10	1 800	**49	165	24*
		,		-								

\*, \*\* = significantly different from the control of  $P \le 0.05$  and 0.01, respectively, by the LSD test. Data are means of three replicates, each from two, combined, experimental units.

affect the root concentrations of phosphorus, iron, and aluminum, and like callaloo, cobalt.

The concentrations of all but three elements were similar in Red sorrel shoots at all Pi s. At Pi =  $10\,000$ , nitrogen, manganese, and iron concentrations were significantly lower, and at Pi = 100 that of nitrogen was higher, than concentrations in control plants (Table 4).

At the highest Pi, Red sorrel roots had significantly lower concentrations of nitrogen, potassium, magnesium, manganese, cobalt, iron, and zinc and higher concentrations of sodium, aluminum, and vanadium than control roots (Table 4). Similar differences occurred at Pi = 100 for magnesium and sodium. Only phosphorus and calcium concentrations remained unaffected by nematode infestation.

There were no macroscopic symptoms of nutrient deficiency or toxicity in any host. Neither were there differences in the shoot/root ratios of any element except sodium and manganese, which were smaller and greater, respectively, in callaloo at  $Pi = 10\,000$  than at Pi = 0 (Table 5).

## DISCUSSION

The reproduction of *M. incognita* were severely inhibited by White sorrel and the mean egg mass index of 2 at the highest Pi indicated that this host, unlike Red sorrel, was highly resistant to the nematode (9). However, plant productivity was reduced in both cultivars.

The egg mass index of 4 for Red sorrel and callaloo at Pi = 10 000 indicated that both these hosts were more susceptible to the nematode than White sorrel. However, the small size of the egg masses and galls on callaloo roots probably resulted from physiological conditions within the roots being unfavorable to parasitism by the nematode. Also, the lack of growth retardation in callaloo indicated that this host had some degree of tolerance to *M. incognita* as noted for the relative, *Amaranthus hybridus* subsp. *incurvatus* L., which is tolerant to *Meloidogyne javanica* (Treub) Chitwood (2). The poor host status of both plants to *M. incognita* in this study confirms the findings by Hutton et al. (11) for the same hosts and Minton (17) for various lines of *H. sabdariffa*.

The degree of susceptibility of the plants to the nematode was not obviously reflected in the differences in their nutrient status resulting from parasitism. For example, the response of all elements, except phosphorus, potassium, and vanadium in the roots, to nematode infestation in both sorrel cultivars differed markedly without showing a general pattern (Table 3, 4). Also, several elements were less concentrated in the roots of infested Red sorrel than in noninfested ones whereas they have been observed to be more concentrated in the roots of infested cowpeas, another highly susceptible host (unpublished data). Therefore, the concentrations of nutrients in the plants appeared to depend on the host/nematode combination.

Table 4. Nutrient content of shoots and roots of Hibiscus sabdariffa cv. Red from soil infested with Meloidogme incognita race 1 for 6 weeks, at four initial nematode densities.

Initial		Essential	Essential macroelements (%)	nents (%)		I	Essential m	icroelem	Essential microelements (ppm)		Other micro- elements (ppm)	nicro- s (ppm)
number of eggs/pot	z	Ь	K	Ca	Mg	Na	Mn	3	Fe	Zn	Ψ	<b>&gt;</b> 2
					Shoots							
0	1.26	0.16	1.25	1.63	0.47	185	245	_	551	117	124	2
100	1.44*	0.16	1.24	1.68	0.46	188	248	,	,	1	125	2
1 000	1.26*	0.16	1.28	1.71	0.43	162	255	,	ı		123	2
10 000	0.78	0.14	1.22	1.65	0.51	160	186**	-	312**	46	122	2
					Roots							
0	06.0	0.11	2.12	1.04	1.78	2 700	163	12	7 900	98	154	25
100	1.08	0.11	1.77	0.98	1.25**	4 400**	181	,	1		151	56
1 000	0.77	0.13	1.68*	0.98	1.19**	3900**	154	,	•		148	56
10 000	0.56**	0.15	1.73*	1.05	1.18**	4 300**	133**	*6	3 200**	*89	225**	35**
Data are means of three renlicates each from two combined exnerimental units	renlicates e	ach from	two com	hined ex	perimenta	unite						

Data are means of three replicates, each from two, combined, experimental units. \*, \*\* = significantly different from the control at  $P \le 0.05$  and 0.01, respectively, by the LSD test.

Table 5. Shoot/root ratios of nutrients in Amaranthus viridis and two cultivars of Hibiscus sabdariffa from pots infested with Meloidogyne incognita race 1 for 6 weeks, at four initial nematode densities.

	Other micro- elements	Al			0.05 0.05					1.89   0.05					0.83   0.06		
		Zn		1.17	•	•	1.18		0.82			0.70		1.36	•	•	1.45
	ements	Fe		0.12			0.10		0.25			0.22		0.02		•	0.09
	Essential microelements	Co		0.50	•	•	0.30		0.12	•	1	0.10		0.08	1	,	0.11
oor ratio	Essenti	Mn		0.41	0.48	0.50	0.74*		2.14	1.70	1.67	1.58		1.50	1.37	1.65	1.40
Shoot/Koot ratio		Na	85.4	0.20	0.18	0.14	*90.0	. White²	0.05	0.09	0.05	80.0	v. Red²	0.44	0.46	0.37	0.47
		Mg	A. viridis <sup>3</sup>	1.41	1.90	1.31	1.36	H. sabdariffa cv. White²	0.28	0.26	0.29	0.27	H. sabdariffa cv. Redz	0.26	0.37	0.36	0.43
	ements	Ca		1.53	2.47	1.86	1.76	H. sa	1.78	1.64	1.66	1.75	H. s	1.57	1.71	1.74	1.57
	Essential macroelements	K		1.55	1.36	1.43	1.56		0.56	0.55	0.61	0.70		0.59	0.70	0.76	0.71
	Essentia	Ъ		1.30	1.20	1.17	1.26		1.70	1.64	1.54	1.54		1.45	1.45	1.23	0.93
		Z		1.03	0.81	0.98	1.14		2.20	1.80	1.84	1.84		1.40	1.33	1.64	1.39
	Initial	eggs/pot		0	100	1 000	10 000		0	100	1 000	10 000		0	100	1 000	10 000

\* = significantly different from the control at  $P \le 0.05$  by the LSD test.

<sup>&</sup>quot;Means of six replicates.

<sup>&</sup>lt;sup>2</sup>Means of three replicates, each from two, combined, experimental units.

Nematode-influenced changes in nutrient concentrations varied with the plant organ. Roots exhibited differences more frequently than shoots. Differences in nutrient concentrations between infested and noninfested plants similar to those noted in these studies have been reported for nitrogen (21), phosphorus (5) potassium (6, 12, 19), calcium (13, 14, 15), magnesium (8), sodium (21), manganese (18), and iron (12).

Translocation of manganese may have been increased in infested plants as the shoot/root ratio was higher at  $Pi=10\,000$  than at Pi=0. At the highest Pi, sodium translocation appeared to be inhibited, as indicated by the significantly low shoot/root ratio. However, as the ratios for all other elements were unaffected by nematode infestation, M. incognita may have caused alterations mainly in the nutrient concentrations of the plants rather than in the translocation of the elements.

### LITERATURE CITED

- 1. ANONYMOUS. 1983. Description and Safety Analyses for the SLOWPOKE-II Reactor. Atomic Energy Agency of Canada, Ltd., Toronto, Canada.
- 2. BAFOKUZARA, N. D. 1983. Reaction of Amaranthus hybridus subsp. incurvatus to varying levels of Meloidogyne javanica inoculum. Nematropica 13:17–26.
- 3. CHRISTIE, J. R., and V. G. PERRY. 1951 Removing nematodes from soil. Proceedings of the Helminthological Society of Washington 18:106–108.
- 4. DICKSON, D. W., and F. B. STRUBLE. 1965. A sieving-staining technique for extraction of egg masses of *Meloidogyne incognita* from soil. Phytopathology 55:497. (Abstr.).
- DROPKIN, V. G., and R. C. KING. 1956. Studies on plant parasitic nematodes homogeneously labelled with radio-phosphorus. Experimental Parasitology 5:469– 480.
- FATEMY, F., and K. EVANS. 1986. Effects of Globodera rostochiensis and water stress on shoot and root growth and nutrient uptake of potatoes. Revue de Nématologie 9:181–184.
- 7. FAWCETT, G. S., and R. H. STOUGHTON. 1944. The Chemical Testing of Plant Nutrient Solutions. The Tintometer, Ltd., Salisbury, U.K.
- 8. GOSWAMI, B. K., S. SINGH, and V. S. VERMA. 1976. Uptake and translocation of calcium and magnesium in tomato plants as influenced by infection with root-knot nematode *Meloidogyne incognita* and tobacco mosaic virus. Nematologica 22:116–117.
- HARTMAN, K. M., and J. N. SASSER. 1985. Identification of Meloidogyne species on the basis of differential host test and perineal-pattern morphology. Pp. 66-77 in K. R. Barker, C. C. Carter, and J. N. Sasser, eds. An Advanced Treatise on Meloidogyne. Vol. 2: Methodology. Department of Plant Pathology, North Carolina State University and U. S. Agency for International Development: Raleigh, North Carolina, U.S.A.
- 10. HEWITT, E. J. 1952. Sand and Water Methods used in the Study of Plant Nutrition. Commonwealth Agriculture Bureau, London, U. K.
- 11. HUTTON, D. G., P. L COATES-BECKFORD, and S. A. E. EASON-HEATH. 1983. Management of *Meloidogyne incognita* populations by crop rotation in a small-scale field trial and nematode pathogenic effects on selected cultivars. Nematropica 13:153–163.
- 12. IBRAHIM, I. K. A., M. A. REZK, and H. A. A. KHALIL. 1984. Effects of Meloidogyne incognita, Fusarium oxysporum and F. vasinfectum on plant growth and mineral content of cotton, (Gossypium barbadense L.). Nematologica 28:298.

- 13. JENKINS, W. R., and R. B. MALEK. 1966. The influence of nematodes on absorption and accumulation of nutrients in vetch. Soil Science 101:46–49.
- 14. MALEKEBERHAN, H., J. M. WEBSTER, R. C. BROOKE, J. M. D'AURIA, and M. CACKETTE. 1987. Effects of *Meloidogyne incognita* on plant nutrient concentration and its influence on the physiology of beans. Journal of Nematology 19:324–330.
- MALEKEBERHAN, H., R. C. BROOKE, J. M. WEBSTER, and J. M. D'AURIA. 1985. The influence of *Meloidogyne incognita* on the growth, physiology and nutrient content of *Phaseolus vulgaris* (cultivar Topnotch Golden Wax). Physiological Plant Pathology 26:259–268.
- McCLURE, M. A., T. H. KRUK, and I. MISAGHI. 1973. A method of obtaining quantities of clean Meloidogyne eggs. Journal of Nematology 5:230.
- 17. MINTON, N. A. 1970. Reaction of kenaf and roselle to three root-knot nematode species. Phytopathology 60:1844–1845.
- 18. NASR, T. A., I. K. A. IBRAHIM, E. M. EL-AZAB, and M. W. A. HASSAN. 1982. The effect of root-knot nematodes on the mineral, amino-acid and carbohydrate concentrations of almond and peach root-stock. Nematologica 26:133–138.
- 19. OTEIFA, B. A. 1952. Potassium nutrition of the host in relation to infection by a root-knot nematode, *Meloidogyne incognita*. Proceedings of the Helminthological Society of Washington 19:99–104.
- REES, W. J., and G. H. SIDRAK. 1956. Plant nutrition of fly-ash. Plant and Soil 8:141-159.
- 21. SHAFIEE, M. F., and W. R. JENKINS. 1963. Host-parasite relationships of Capsicum frutescens and Pratylenchus penetrans, Meloidogyne incognita acrita and Meloidogyne hapla. Phytopathology 53:325-328.
- 22. STEELE, R. G. D., and J. H. TORRIE. 1960. Principles and Procedures of Statistics. McGraw-Hill: New York.
- TARAS, M. J. 1958. Nitrogen. Pp. 75-160 in D. F. Boltz, ed. Chemical Analysis Volume 8. Colorimetric Determination of Nonmetals. Interscience Publishers, Inc.: New York.
- WOLF, B. 1943. Rapid determination of soluble nutrients in soil and plant extract by means of photoelectric colorimeter. Industrial and Engineering Chemistry Analyses 15:948

Received for publication:

20.VIII.1989

Recibido para publicar:

## **ACKNOWLEDGEMENTS**

We wish to thank Professor G. H. Sidrak for his advice on the nutrient determinations, Mr. R. Rattray for his assistance with some of the neutron activation analyses, Mr. W. Fielding for the statistical analyses, and Mr. D. G. Hutton for supplying the sorrel seeds. We also thank the referees for their helpful comments.