

PLANT-PARASITIC NEMATODES ASSOCIATED WITH WEEDS IN POTATO (*SOLANUM TUBEROSUM* L.) FIELDS FROM THE NORTHERN AREA OF CARTAGO, COSTA RICA

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ABSTRACT

Núñez Rodríguez, L. A., D. A. Humphreys-Pereira, A. M. Rodríguez-Ruiz, and L. Flores-Chaves. 2022. Plant-parasitic nematodes associated with weeds in potato (*Solanum tuberosum* L.) fields from the northern area of Cartago, Costa Rica. *Nematropica* 52:33-44.

Potatoes are the main vegetable produced in Costa Rica, however, basic information about the interaction of nematodes and alternative hosts in potato fields is lacking. The objective of this study was to identify the plant-parasitic nematodes associated with weeds in potato fields from the northern area of Cartago, Costa Rica. Weeds from 15 potato fields (3 months after planting) were sampled. To determine the most frequent weed species, 5 sections (approximately 4 m²) were sampled in each field using a 1 m² square. The most frequent weed species (a maximum of five) were selected, and five composite root samples per weed were processed for subsequent nematode identification and quantification. Twenty-seven weed species were identified in the potato fields, with *Brassica campestris*, *Lepidium virginicum*, and *Poa annua* among the six most frequent weed species. Fifteen nematode genera, two families, and a subfamily were identified associated with weed roots. There were statistical differences between endoparasitic nematodes. *Pratylenchus* had the highest frequency of occurrence relative to heteroderids and *Meloidogyne*. *Pratylenchus* also had the highest average population density of the endoparasitic nematodes with 31,056 nematodes/100 g of roots. Our results provide valuable information for the integrated management of nematodes. Alternative hosts such as weeds should be managed after the potato production cycle. Fallow could increase population densities of plant-parasitic nematodes, detrimental to other common crops in the region, such as onions and carrots.

Key words: *Meloidogyne*, *Pratylenchus*, potato, weeds, quantification, roots

RESUMEN

Núñez Rodríguez, L. A., D. A. Humphreys-Pereira, A. M. Rodríguez-Ruiz, and L. Flores-Chaves. 2022. Nematodos fitoparásitos asociados a malezas en el cultivo de papa (*Solanum tuberosum* L.) en la zona norte de Cartago, Costa Rica. *Nematropica* 52:33-44.

La papa es la principal hortaliza cultivada en Costa Rica, sin embargo, falta información básica sobre la interacción de nematodos con hospederos alternativos en plantaciones de papa. El objetivo de este estudio fue identificar los nematodos fitoparásitos asociados a las principales malezas en plantaciones de papa de la zona norte de Cartago, Costa Rica. Se muestrearon malezas en 15 plantaciones de papa (tres meses de edad). En cada finca se muestrearon cinco puntos (aproximadamente 4m²) utilizando un marco de 1m² para determinar las malezas más frecuentes. Se seleccionaron las especies de malezas más frecuentes (un

máximo de cinco) y se recolectaron cinco muestras compuestas por especie para su procesamiento, identificación y cuantificación de nematodos. Se identificaron un total de 27 especies de malezas en las plantaciones de papa, de las cuales, seis especies fueron las más frecuentes, entre ellas *Brassica campestris*, *Lepidium virginicum*, y *Poa annua*. Se identificaron 15 géneros de nematodos, dos familias y una subfamilia. Se observó diferencia estadística significativa entre nematodos endoparásitos, donde *Pratylenchus* fue el más frecuente en comparación a heteroderides y *Meloidogyne*. *Pratylenchus* también tuvo la mayor densidad media poblacional entre los nematodos endoparásitos con 31,056 nematodos en 100 g de raíz. Nuestros resultados muestran información valiosa para el manejo integrado de nematodos. Los hospederos alternativos como las malezas se deberían manejar selectivamente después del ciclo productivo de la papa. El barbecho podría incrementar las densidades de nematodos fitoparásitos, en detrimento de otros cultivos de la región, como cebolla y zanahoria.

Palabras clave: Cuantificación, *Meloidogyne*, *Pratylenchus*, malezas, papas, raíces

INTRODUCTION

Potato (*Solanum tuberosum* L.) is considered the third most important food in the world, only surpassed by wheat (*Triticum aestivum* L.) and rice (*Oryza sativa* L.) (Juyó *et al.*, 2015). In Costa Rica, there are 3,218 ha of potato, cultivated in Alajuela, Cartago, Heredia and San José Provinces. However, the vast majority of potatoes are grown in Alajuela and Cartago with 867 and 2,553 ha, respectively. Zarcero in Alajuela and the north area in Cartago are the main production areas (SEPSA, 2018).

As with many other crops, potatoes are affected by different pests, such as plant-parasitic nematodes, which can cause yield losses of about 12.2% (Ravichandra, 2014). Among the most important nematodes associated with potatoes worldwide are *Globodera*, *Meloidogyne*, and *Pratylenchus* (Jones *et al.*, 2013; Palomares *et al.*, 2014). These genera have been reported in potato fields in Costa Rica (López and Azofeifa, 1981). *Globodera* spp. were described morphologically and molecularly in 2009 (García *et al.*, 2009), while there are two *Meloidogyne* spp., *M. incognita* and *M. hapla*, and two *Pratylenchus* species, *P. crenatus* and *P. penetrans*, reported in the country (Montero *et al.*, 2007; Sandoval 2015, 2018).

The presence of alternative hosts is a factor that makes plant-parasitic nematode control difficult. Weeds can allow plant-parasitic nematodes to survive with or without the presence of a crop, which provides a constant source of nematode inoculum for future crop cycles (Kaur *et al.*, 2007). In the southeastern United States, *Rotylenchulus reniformis* is an important plant-parasitic nematode and its ability to reproduce on

different weeds such as *Sida spinosa*, *Amaranthus retroflexus*, *Ambrosia artemisiifolia*, and *Ipomoea hederacea* has been demonstrated (Lawrence *et al.*, 2008). In Florida, *Meloidogyne* spp. can reproduce on weeds such as *Eleusine indica*, *Portulaca oleracea*, *Solanum americanum*, *Amaranthus* spp., and *Lepidium virginicum* (Myers *et al.*, 2004) and in the Netherlands on *Senecio vulgaris* and *Capsella bursa-pastoris* (Kutywayo and Been, 2006).

Studies carried out in countries such as Bulgaria and the USA have reported the ability of weeds associated with potato production to host genera such as *Pratylenchus* and *Meloidogyne* (Samaliev and Kalinova, 2013; Smiley *et al.*, 2014). In Québec, Canada, weeds frequently associated with potato crops include *A. retroflexus*, *A. artemisiifolia*, *Cyperus rotundus*, *Chenopodium album*, *Polygonum persicaria*, and *Spergula arvensis*, which are hosts of *Pratylenchus penetrans* (Bélair *et al.*, 2007). Weeds in the genus *Solanum* have been reported as hosts of *G. pallida* in Idaho, USA (Boydston *et al.*, 2010). In Costa Rica, vegetables like carrots, onions, and potatoes are commonly rotated and, in many cases, they are hosts for the same plant-parasitic nematodes. However, studies on the host status of weeds for plant-parasitic nematodes associated with vegetable crops are lacking. López and Salazar (1978) reported *M. hapla* associated with *Sonchus oleraceus*, *Galinsoga parviflora*, *Hipochaeris radicata*, *Iresine celosia*, *Conyza coronapifolia*, *Cirsium mexicanum*, *Rumex acetocella*, *S. americanum*, and *Gnaphalium americanum*. In a nematode survey in the largest vegetable production areas of Costa Rica, *M. hapla* was associated with *Bidens pilosa* and *Cirsium*

subcoriaceum (López and Azofeifa, 1981). The objective of this study was to identify the plant-parasitic nematodes associated with weeds in potato fields in Cartago, Costa Rica.

MATERIALS AND METHODS

Collection of weed samples

Between August 2016 and March 2017, 15 potato fields (3 and 4 months old, no larger than 1 ha) located in the northern area of Cartago, Costa Rica (Table 1) were sampled for weed identification and nematode analysis. The minimum sampling area was determined according to Fuentes (1986), as follows: a square frame of 1 m² PVC frame divided with nylon in 100 squares of 10 cm² was randomly placed in a potato field and

plant species within the frame were recorded. The initial area was doubled (2 m²), and new weed species were recorded. This process was repeated until no new species were recorded. For all potato fields, the minimum sampling area was 4 m². At each field, five points of 4 m² were selected randomly for the weed survey following the quadrant method (Mostacedo and Fredericksen, 2000). The ground cover percentage (total of squares occupied by each weed species) was calculated with the 1 m² frame. Weeds were identified based on visible characteristics such as inflorescence, flower structure, leaf features (shape, presence or absence of hairs, margin, venation, and arrangement on the stem). To evaluate weed species composition, frequency of occurrence of weeds (FO%) was calculated for the 15 fields that were sampled (Ntidi *et al.*, 2012):

Table 1. Frequency of occurrence of weed species (FO%) sampled in the northern area of Cartago, Costa Rica.

Genus and species	FO%
<i>Brassica campestris</i>	46.7 ay ^z
<i>Bromus carinatus</i>	6.7 c
<i>Calceolaria mexicana</i>	13.3 bc
<i>Capsella bursa-pastoris</i>	6.7 c
<i>Cardamine flaccida</i>	6.7 c
<i>Chenopodium album</i>	6.7 c
<i>Commelina difusa</i>	13.3 bc
<i>Cyperus sculentus</i>	6.7 c
<i>Cyperus sesquiflorus</i>	6.7 c
<i>Cyperus</i> sp.	6.7 c
<i>Eleusine indica</i>	6.7 c
<i>Galinsoga parviflora</i>	6.7 c
<i>Galinsoga quadriradiata</i>	6.7 c
<i>Juncus bufonius</i>	20 b
<i>Lepidum virginicum</i>	46.7 a
<i>Lolium perenne</i>	20 b
<i>Melampodium perfoliatum</i>	13.3 bc
<i>Plantago australis</i>	6.7 c
<i>Poa annua</i>	46.7 a
<i>Polygonum aviculare</i>	13.3 bc
<i>Polygonum persicariodes</i>	40 a
<i>Richardia scabra</i>	13.3 bc
<i>Rumex acetocella</i>	6.7 c
<i>Rumex obtusifolius</i>	40 a
<i>Senecio vulgaris</i>	6.7 c
<i>Sonchus oleraceus</i>	6.7 c
<i>Spergula arvensis</i>	33.3 a

^zFrequency of occurrence data were analyzed for effects using χ^2 analysis. Values followed by the same letter are not significantly different from each other ($P < 0.05$).

FO% = (Number of potato fields in which the weed species occurred/number of potato fields sampled) \times 100. Then, the FO% data were then analyzed using the non-parametric χ^2 test for independence for effects of location using R software (R Core Team, 2017).

Extraction and identification of nematodes

The five weed species with the highest ground cover percentage in each potato field were sampled for nematode analysis. Five composite root samples (3 to 5) of each selected weed were collected in a zig-zag pattern (soil surrounding the roots was also collected when immature females of cyst-forming nematodes were observed). Each root system was checked under a stereomicroscope (Nikon SMZ745T; Nikon Instruments Inc., Melville, NY) for galls or cyst-forming nematodes. Nematodes were extracted from roots using the floatation-centrifugation method (Caveness and Jensen, 1955). Plant-parasitic nematodes were identified to the genus level based on morphology. A soil sample of 200 g was processed using the Fenwick method (Shepherd, 1986) in cases where immature females of cyst-forming nematodes were observed attached to the host roots and identified to the genus level based on cyst shape. The frequency of occurrence of plant-parasitic nematodes was calculated according to Barker (1985): (number of samples where a nematode genus was present/total amount of samples) \times 100, (number of samples where a nematode genus was present/amount of samples per district) \times 100, (number of samples where a nematode genus was present/amount of samples per weed species) \times 100 and (number of samples where a nematode genus was present/amount of samples per weed family) \times 100. Maximum and mean population densities were also estimated. The frequency of occurrence was analyzed by the χ^2 test for independence for effects of districts. Kruskal-Wallis and Mann-Whitney tests were used to analyze effects of districts on population nematode densities.

RESULTS

Weed species identification in potato fields

Fifteen fields distributed in six districts in the northern area of Cartago were sampled; Capellades (altitude ranged between 2,500 and 2,900 m.a.s.l.),

Cervantes (1,650 m.a.s.l.), Cot (1,680 and 1,780 m.a.s.l.), Llano Grande (2,100 and 2,190 m.a.s.l.), Pacayas (1,630 and 2,620 m.a.s.l.), and Potrero Cerrado (2,110 and 2,170 m.a.s.l.). Twenty-seven weed species were identified as the most important. *Brassica campestris*, *Lepidum virginicum*, *Poa annua*, *Polygonum persicarioides*, *Rumex obtusifolius*, and *Spergula arvensis* had the highest frequency of occurrence ($P < 0.05$) (Table 1).

Weed species were grouped into 12 botanical families and analyzed based on frequency of occurrence (Table 2). The Brassicaceae were found more frequently in the districts of Capellades and Pacayas ($P < 0.05$). The Polygonaceae were the most frequently encountered family of weeds in the district of Pacayas ($P < 0.05$). Meanwhile, families such as Plantaginaceae and Scrophulariaceae were found only in the district of Capellades ($P < 0.05$) (Table 2). The frequency of occurrence of weed species were analyzed for each district; however, there was no statistical differences between weed species in each district ($P > 0.05$; data not shown).

Frequency and population density of plant-parasitic nematodes

Twenty-seven weed species were sampled for nematode analyses resulting in a total of 338 composite root samples. A total of 18 plant-parasitic nematodes were identified associated with weeds. Although there was no clear differentiation of frequency of occurrence and mean population densities between all plant-parasitic nematodes found in this study, there were significant differences among the two most damaging endoparasites, *Pratylenchus* and *Meloidogyne*, in vegetables in the region. *Pratylenchus* had the highest frequency of occurrence and mean population densities per 100 g of roots ($P < 0.05$) (Table 3).

Heteroderids were found in five of the six sampled districts and the highest mean population density was found in the district of Llano Grande ($P < 0.05$; Table 4). Immature females of Heteroderinae were observed only on roots of *R. obtusifolius* and cysts recovered from soil surrounding the roots were lemon-shaped and had prominent vulval cone characteristic of the genus *Heterodera* (Fig. 1). *Meloidogyne* spp. were found in all six districts and were most frequently found in the districts of Cervantes and Cot. Meanwhile,

Table 2. Frequency of occurrence (FO%) of weed families in six districts of the Northern area of Cartago, Costa Rica.

Weed family ^y	District					
	Capellades	Cervantes	Cot	Llano Grande	Pacayas	Potrero Cerrado
	FO%					
Asteraceae	14 b ^z	0 c	34.5 a	0 c	34.5 a	17 b
Brassicaceae	29.8 a	11.9 b	5.9 b	11.9 b	29.8 a	10.7 b
Cariophylliaceae	60 a	0 c	0 c	0 c	0 c	40 b
Chenopodiaceae	0 b	0 b	0 b	100 a	0 b	0 b
Commelinaceae	0 b	0 b	50 a	0 b	50 a	0 b
Cyperaceae	0 c	0 c	66.7 a	0 c	33.3 b	0 c
Juncaceae	66.7 a	0 c	0 c	0 c	33.3 b	0 c
Plantaginaceae	100 a	0 b	0 b	0 b	0 b	0 b
Poaceae	27.3 a	9.0 b	0 c	27.3 a	27.3 a	9.0 b
Polygonaceae	13.3 bc	6.7 c	13.3 bc	20 b	40 a	6.7 c
Rubiaceae	0 b	50 a	50 a	0 b	0 b	0 b
Scrophulariaceae	100 a	0 b	0 b	0 b	0 b	0 b

^yNumber of observations per weed family = 29 (Asteraceae), 84 (Brassicaceae), 25 (Cariophylliaceae), 5 (Chenopodiaceae), 10 (Comelinaceae), 15 (Cyperaceae), 15 (Juncaceae), 5 (Plantaginaceae), 55 (Poaceae), 75 (Polygonaceae), 10 (Rubiaceae), and 10 (Scrophulariaceae).

^zFrequency of occurrence data were analyzed for effects using χ^2 analysis. Values followed by the same letter in the same row are not significantly different from each other ($P < 0.05$).

Table 3. Frequency of occurrence (FO%), maximum (max) population density, and mean population density (nematodes/100 g of roots) when present in a sample of plant-parasitic nematodes in the northern area of Cartago, Costa Rica^x.

Genus/family	Parameter		
	FO %	Max	Mean
<i>Aphelenchoides</i>	11.8 defg ^y	160	29 d ^z
<i>Aphelenchus</i>	16.6 cde	880	98 cd
Criconematidae	4.1 efg	80	29 cd
<i>Ditylenchus</i>	8 defg	120	27 d
<i>Gracilacus</i>	0.9 fg	18	13 abcd
<i>Helicotylenchus</i>	46.7 ab	8,520	526 b
<i>Hemicycliophora</i>	15.1 def	105	27 d
Heteroderidae	26 bcd	7,350	293 bc
<i>Hoplolaimus</i>	0.9 fg	20	17 abcd
<i>Longidorus</i>	0.9 fg	10	10 abcd
<i>Meloidogyne</i>	43.2 b	31,120	1,076 b
<i>Paralongidorus</i>	0.3 g	10	10 abcd
<i>Paratylenchus</i>	0.6 fg	10	10 abcd
<i>Pratylenchus</i>	69.5 a	104,720	9,520 a
Trichodoridae	4.4 efg	29	14 d
<i>Tylenchulus</i>	0.9 fg	160	107 abcd
<i>Tylenchus</i>	39.6 bc	240	49 cd
<i>Xiphinema</i>	0.3 g	42	42 abcd

^xNumber of samples included in the analysis was 338.

^yFrequency of occurrence data were analyzed for effects using χ^2 analysis. Values followed by the same letter are not significantly different from each other ($P < 0.05$)

^zMean values followed by the same letter are not significantly different ($P < 0.05$) according to Kruskal-Wallis and Mann-Whitney nonparametric statistical tests.

Table 4. Frequency of occurrence (FO%), maximum (max) population density and mean population density (nematodes/100 g of roots) when present in a sample of the endoparasitic nematodes Heteroderidae, *Meloidogyne* and *Pratylenchus* in the northern area of Cartago, Costa Rica^x.

District	Heteroderidae			<i>Meloidogyne</i>			<i>Pratylenchus</i>		
	FO%	Max	Mean	FO%	Max	Mean	FO%	Max	Mean
Capellades	35.1 a ^y	680	110 a ^z	31.9 a	1,360	142 a	97.9 a	79,280	9,802 a
Cervantes	0 b	0	0	96 b	3,079	646 b	0 b	0	0
Cot	28.9 ac	470	58 c	75.6 b	1,580	170 c	20 c	65	31 b
Llano Grande	13.3 bc	7,350	3,077 d	33.3 a	650	141 acd	80 d	11,420	3,569 c
Pacayas	24.8 ac	570	98 a	39 a	31,120	3,158 bc	76.2 d	104,720	15,009 a
Potrero Cerrado	41.7 a	230	41 c	8.3 c	10	10 d	75 d	1,220	328 d

^xNumber of samples included in the analysis = 94 (Capellades), 25 (Cervantes), 45 (Cot), 45 (Llano Grande), 105 (Pacayas), and 24 (Potrero Cerrado).

^yFrequency of occurrence data were analyzed for effects using χ^2 analysis. Values followed by the same letter in the same column are not significantly different from each other ($P < 0.05$)

^zMean values followed by the same letter in the same column are not significantly different ($P < 0.05$) according to Kruskal-Wallis and Mann-Whitney nonparametric statistical tests.

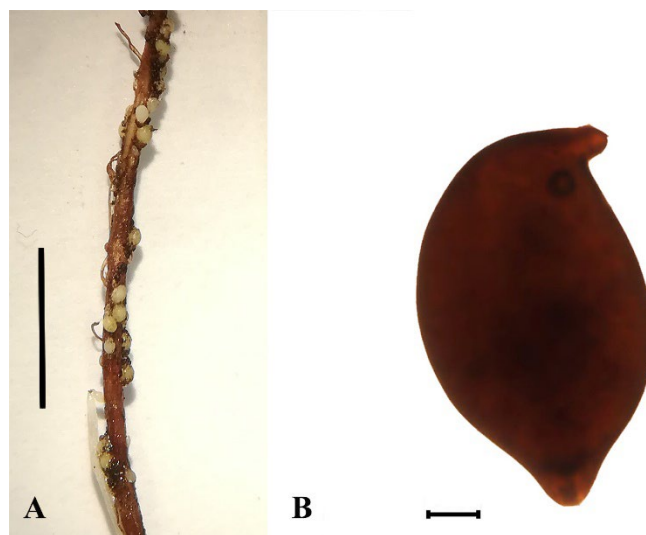


Figure 1. Immature females (A), and cyst (B) of *Heterodera trifolii* in roots of *Rumex obtusifolius*. (Scale bars: A= 5 mm, and B= 100 μ m).

Potrero Cerrado had the lowest frequency of occurrence ($P < 0.05$; Table 4). *Pratylenchus* spp. were found in five of the six sampled districts. Capellades and Cot had the highest and lowest frequency of occurrence, respectively ($P < 0.05$; Table 4). From the five districts where *Pratylenchus* spp. were found, Pacayas and Capellades had the highest mean population densities per 100 g of roots ($P < 0.05$; Table 4).

The average population density of *Pratylenchus* spp. was more than 15,000 nematodes/100 g of roots on weeds species such as

S. vulgaris, *P. annua*, and *Calceolaria mexicana*. The average *Pratylenchus* spp. population density was less than 1,000 nematodes/100 g of roots on *C. bursa-pastoris*, *Galinsoga quadriradiata*, *Melampodium perfoliatum*, and *Richardia scabra*. *Pratylenchus* spp. were not detected in seven weeds (*Cardamine flaccida*, *Commelina diffusa*, *Cyperus esculentus*, *C. sesquiflorus*, *Cyperus* sp., *Eleusine indica*, and *Galinsoga parviflora*). Meanwhile, *G. parviflora* had the highest average population density of *Meloidogyne* spp. with more than 20,000 nematodes/100 g roots (Table 5). The

frequencies of occurrence and mean population densities of *Meloidogyne* and *Pratylenchus* per weed family and species found per district were not significantly different ($P > 0.05$; data not shown).

DISCUSSION

The Province of Cartago is recognized as the main vegetable-producing area of Costa Rica. In the northern area of this province, onions, carrots, and potatoes are commonly produced in rotation. Few studies are available on the plant-parasitic nematodes found in these crops or associated with weeds in Costa Rica (García *et al.*, 2009; Sandoval *et al.*, 2020). Two of the most frequent weed species (*B. campestris* and *P. annua*) associated with potato fields in this study have been reported in horticultural crops, such as carrots and onions in the province of Cartago, Costa Rica (Villalobos, 1999; Araya, 2000; Rodríguez *et al.*, 2015). In Guatemala, *B. campestris* is frequently associated with potato fields (Marroquin, 1993), whereas in Colombia, *P. annua*, *S. arvensis*, *S. vulgaris*, *S. oleraceus*, *C. bursa-pastoris*, *Brassica* spp., *Polygonum aviculare*, and *P. segetum* are commonly found in cold weather crops, such as potato (Cárdenas *et al.*, 1970; Jamaica and Plaza, 2014).

Weeds can compete with crops for resources such as water, nutrients, and light. Also, weeds have been reported as pest and pathogen reservoirs, mainly for plant-parasitic nematodes (Quénéhervé *et al.*, 2011). Potato growers in Cartago commonly leave fields fallow between each production cycle, which allows the development of weeds. In this study, 15 genera of nematodes were found associated with weed roots, among them *Aphelenchoides*, *Aphelenchus*, and *Tylenchus* that might include root feeders and/or fungivores (Hofman and S'Jacob, 1989; Yeates *et al.*, 1993).

In some samples, *Ditylenchus* spp. and Trichodorids were found in low frequencies; there are no records of the potential damage to potatoes by these nematodes in Costa Rica. In other countries, these nematodes cause significant crop losses, either by direct impact on the crop or by indirect effects as virus vectors in temperate climates. For example, *D. destructor* and *D. dipsaci* can significantly affect potato production, while species in the genera *Trichodorus* and *Paratrichodorus* are vectors of tobacco rattle virus,

the causal agent of corky ringspot on potatoes (Pérez *et al.*, 2000; Plowright *et al.*, 2002). Further research on the identification of these nematode species should be implemented, which is fundamental to reduce the negative impact of these nematodes on crop production.

Two of the most important plant-parasitic nematodes genera worldwide are *Meloidogyne* and *Pratylenchus* (Jones *et al.*, 2013), both of which were found frequently in this study. These genera have been reported to parasitize a large number of weeds (Myers *et al.*, 2004; Kaur *et al.*, 2007; Brito *et al.*, 2008). *Pratylenchus* is a major nematode problem on potato, where species such as *P. penetrans* can cause up to 50% of yield loss (Holgado *et al.*, 2009; Esteves *et al.*, 2015). *Pratylenchus penetrans* was found at high population densities on *P. annua*, *S. oleraceus* and *S. vulgaris* (Bendixen, 1988; Kuttywayo and Been, 2006; Bélair and Simard, 2008).

Meloidogyne can parasitize more than 2,000 plant species (Agrios 2005; Jones *et al.*, 2013). *Meloidogyne* spp. were found at high population densities on *G. parviflora*, which coincides with several studies in United States, Argentina, Costa Rica and Hungary (Lopez and Salazar, 1978; Doucet, 1993; Kuttywayo and Been, 2006). Also, two species, *M. chitwoodi* and *M. hapla*, have been associated with *G. parviflora* (Lopez and Salazar, 1978; Kuttywayo and Been, 2006), both of which can affect potato tuber quality. There were some weed species with low *Meloidogyne* spp. population densities that can potentially be used for management of this genus. For example, in Costa Rica, *L. virginicum* has been reported as a non-host for *M. incognita* (López and Quesada, 1997), a nematode that causes a reduction in the marketability of the potato tuber due to galling (Montero *et al.*, 2007).

Nematodes of the subfamily Heteroderinae were found in *R. obtusifolius* and identified to genus level as *Heterodera* using morphological characters like shape of the cyst and the presence of the vulval cone (Mulvey, 1974), thus discarding the presence of *Globodera* females or cysts (spherical shape and without vulval cone) (Skantar *et al.*, 2011). Recently, this species was identified using both morphological and molecular approaches as *H. trifolii* (Núñez *et al.*, 2021). *Globodera* was not found in weeds associated with potato in the northern area of Cartago.

Table 5. Population density of plant-parasitic nematodes associated with the predominant weeds in potato fields in Cartago, Costa Rica. Data are expressed as number of nematodes per 100 g of roots and in the form of mean (minimum-maximum).

Weed species	<i>Ditylenchus</i>	<i>Helicotylenchus</i>	<i>Heteroderides</i>	<i>Meloidogyne</i>	<i>Pratylenchus</i>	<i>Trichodoridae</i>
<i>Brassica campestris</i>	17 (10-30)	193 (10-1,040)	17 (10-30)	158 (10-420)	1,226 (40-3,590)	10
<i>Bromus carinatus</i>	0	0	0	160 (20-380)	4,964 (3,840-5,780)	0
<i>Calceolaria mexicana</i>	0	41 (32-50)	160 (80-240)	295 (73-516)	15,706 (5,360-30,560)	0
<i>Capsella bursa pastoris</i>	0	0	10	0	10	0
<i>Cardamine flaccida</i>	24	1,556 (120-5595)	0	1,918 (620-3,079)	0	0
<i>Chenopodium album</i>	0	0	0	90 (20-160)	1,890 (420-3,640)	0
<i>Commelina diffusa</i>	20	456 (360-930)	0	1403 (30-3,350)	0	0
<i>Cyperus esculentus</i>	60 (20-80)	188 (60-480)	0	10 (10-10)	0	0
<i>Cyperus sesquiflorus</i>	20	710 (40-1,380)	0	284 (10-720)	0	0
<i>Cyperus</i> sp.	90 (60-120)	5100 (1,260-8,520)	0	120 (120-120)	0	0
<i>Eleusine indica</i>	0	692 (300-1,270)	0	295 (20-950)	0	10
<i>Galinsoga parviflora</i>	0	280 (80-480)	0	22,656 (9,440-31,120)	0	0
<i>Galinsoga quadriradiata</i>	10	0	20	10	636 (260-1,220)	10
<i>Juncus bufonius</i>	0	70 (27-160)	0	32 (10-80)	7,949 (10-21,120)	13 (10-20)
<i>Lepidium virginicum</i>	13 (10-18)	59 (10-263)	21 (10-70)	29 (10-80)	5375 (12-18,000)	0
<i>Lolium perenne</i>	0	80	0	0	14,897 (20-26,400)	0

Table 5. Continued.

Weed species	<i>Diitylenchus</i>	<i>Helicotylenchus</i>	<i>Heteroderides</i>	<i>Meloidiogyne</i>	<i>Pratylenchus</i>	<i>Trichodoridae</i>
<i>Melampodium perfoliatum</i>	10 (10-10)	82 (20-210)	23 (10-40)	62 (20-130)	18 (10-30)	10 (10-10)
<i>Poa annua</i>	10 (10-10)	81 (20-240)	91 (10-230)	48 (10-160)	28016 (20-104,720)	10
<i>Plantago australis</i>	0	144 (80-160)	0	512 (80-1360)	13904 (6,560-20,720)	0
<i>Polygonum aviculare</i>	0	10 (10-10)	10 (10-10)	45 (10-80)	1634 (100-5,800)	0
<i>Polygonum segetum</i>	10 (10-10)	1322 (30-6,620)	128 (10-470)	283 (10-1450)	2829 (10-6,450)	15 (10-20)
<i>Richardia scabra</i>	0	226 (40-583)	31 (19-43)	105 (20-324)	50 (19-65)	29 (28-29)
<i>Rumex acetocella</i>	0	17 (10-20)	0	0	3058 (670-6,380)	0
<i>Rumex obtusifolius</i>	0	19 (10-50)	929 (20-7350)	94 (10-240)	9309 (20-32,720)	10 (10-10)
<i>Senecio vulgaris</i>	0	80	0	80	31,056 (20,080-40,160)	0
<i>Sonchus oleraceus</i>	0	0	0	0	7,040 (1,480-19,680)	0
<i>Spargula arvensis</i>	37 (20-53)	13 (10-20)	86 (10-211)	26 (10-39)	2,094 (60-6,382)	0

Further studies, such as the identification of plant-parasitic nematodes on weeds at the species level and pathogenicity assays to assess the ability for these nematodes to reproduce on weeds are valuable for an integrated nematode management. Potential non-host species may be used to reduce the nematode population densities based on our data. Caution must be taken when other hosts like onions and carrots are planted after fallow, which are common crops in the northern area of Cartago.

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LITERATURE CITED

- Agrios, G. 2005. Plant pathology. 5th edition. California, USA: Academic Press.
- Araya, E. 2000. Control químico de *Polygonum aviculare* y otras malezas en el cultivo de cebolla (*Allium cepa* L.). Tesis de Licenciatura, Universidad de Costa Rica, San José, Costa Rica.
- Barker, K. R. 1985. Sampling nematode communities. Pp. 3-17 in K. R. Barker, C. C. Carter, and J. N. Sasser, eds. An Advanced Treatise on *Meloidogyne*. Volume II: Methodology. Raleigh, NC: North Carolina State University Graphics.
- Bélair, G., N. Dauphinais, D. Benoit, and Y. Fournier. 2007. Reproduction of *Pratylenchus penetrans* on 24 common weeds in potato fields in Québec. *Journal of Nematology* 39:321-326.
- Bélair, G., and L. Simard. 2008. Effect of the root-lesion nematode (*Pratylenchus penetrans*) on annual bluegrass (*Poa annua*). *Phytoprotection* 89:37-39.
- Bendixen, L. 1988. Weed hosts of *Heterodera*, the cyst, and *Pratylenchus*, the root-lesion, nematodes. Ohio Agricultural Research and Development Center, Special Circular 117. Wooster.
- Boydston, R., H. Mojtahedi, C. Bates, R. Zemetra, and C. Brown. 2010. Weed hosts of *Globodera pallida* from Idaho. *Disease Notes* 94:918.
- Brito, J., R. Kaur, R. Cetintas, J. Stanley, M. Mendes, E. McAvoy, T. Powers, and D. Dickson. 2008. Identification and characterization of *Meloidogyne* spp. infecting horticultural and agronomic crops, and weeds in Florida. *Nematology* 10:757-766.
- Cárdenas, J., O. Franco, C. Romero, and D. Vargas. 1970. Malezas de clima frío. Colombia: Plant Protection Center Oregon State-Instituto Colombiano Agropecuario.
- Caviness, F., and H. Jensen. 1955. Modification of the centrifugal flotation for the isolation and concentration of nematodes and their eggs from soil plant tissue. *Proceedings of the Helminthological Society of Washington* 22:87-89.
- Doucet, M. 1993. Consideraciones acerca del género *Meloidogyne* Goeldi, 1887 (Nemata: Tylenchida) y su situación en Argentina. *Asociaciones y distribución. AgriScientia* 10:63-80.
- Esteves, I., C. Maleita, and I. Abrantes. 2015. Root-lesion and root-knot nematodes parasitizing potato. *European Journal of Plant Pathology* 141:397-406.
- Fuentes, C. 1986. Metodologías y técnicas para evaluar las poblaciones de malezas y su efecto en los cultivos. *Revista Comalfi* 13:29-50.
- García, D., C. García, Z. Montero, L. Salazar, A. Brenes, and L. Gómez. 2009. Morphological and molecular identification of potato cyst-forming nematode *Globodera pallida* in soil samples from Costa Rica. *Revista Latinoamericana de la Papa* 15:38-45.
- Hofman, T., and J. S'Jacob. 1989. Distribution and dynamics of mycophagous and microbivorous nematodes in potato fields and their relationship to some food sources. *Annals of Applied Biology* 115:219-298.
- Holgado, R., K. Oppen, and C. Magnusson. 2009. Field damage in potato by lesion nematode *Pratylenchus penetrans*, its association with tuber symptoms and its survival in storage. *Nematología Mediterránea* 37: 25-29.
- Jamaica, D., and G. Plaza. 2014. Evaluación de diferentes metodologías convencionales de muestreo de malezas en cultivos de papa y espinaca. *Agronomía Colombiana* 31:36-43.
- Jones, J. T., A. Haegeman, E. G. Danchin, H. S. Gaur, J. Helder, M. G. Jones, T. Kikuchi, R. Manzanilla-Lopez, J. E. Palomares-Rius, W. M. Wesemael, and R. N. Perry. 2013. Top 10 plant parasitic nematodes in molecular plant

- pathology. *Molecular Plant Pathology* 14:946–961.
- Juyó, D., F. Sarmiento, M. Álvarez, H. Brochero, C. Gebhardt, and T. Mosquera. 2015. Research: Genetic diversity and population structure in diploid potatoes of *Solanum tuberosum* Group Phujera. *Crop Science* 55:760–769.
- Kaur, R., J. Brito, and J. Rich. 2007. Host suitability of selected weed species to five *Meloidogyne* species. *Nematropica* 37:107–120
- Kutywayo, V., and T. H. Been. 2006. Host status of six major weeds to *Meloidogyne chitwoodi* and *Pratylenchus penetrans*, including a preliminary field survey concerning other weeds. *Nematology* 8:647–657.
- Lawrence, K., A. Price, G. Lawrence, J. Jones, and J. Akridge. 2008. Weed hosts for *Rotylenchulus reniformis* in cotton fields rotated with corn in the southeast of the United States. *Nematropica* 38:13–22.
- López, R., and J. Azofeifa. 1981. Reconocimiento de nematodos fitoparásitos asociados con hortalizas en Costa Rica. *Agronomía Costarricense* 5:29–35.
- López, R., and M. Quesada. 1997. Reproducción de *Meloidogyne incognita* en varias malezas presentes en Costa Rica. *Agronomía Mesoamericana* 8:112–115.
- López, R., and L. Salazar. 1978. Morfometría y algunos hospedantes de *Meloidogyne hapla* en la Cordillera Volcánica Central de Costa Rica. *Agronomía Costarricense* 2:29–38.
- Marroquin, J. 1993. Determinación del período crítico de interferencia de malezas en el cultivo de papa (*Solanum tuberosum* L.) en la aldea La Toma, Santa María Xalapan, Jalapa. Tesis de Licenciatura, Universidad de San Carlos de Guatemala, Guatemala.
- Montero, Z., C. García, L. Salazar, R. Valverde, and L. Gómez. Detección de *Meloidogyne incognita* en tubérculos de papa en Costa Rica. *Agronomía Costarricense* 31:77–84.
- Mostacedo, B., and T. Fredericksen. 2000. Manual de métodos básicos de muestreo y análisis en ecología vegetal. Santa Cruz, Bolivia. Proyecto de Manejo Forestal Sostenible. Available from: <http://www.bionica.info/biblioteca/mostacedo2000ecologiv egetal.pdf>
- Myers, L., K. Wang, R. Mcsorley, and C. Chase. 2004. Investigations of weeds reservoirs of plant parasitic nematodes in agricultural systems in Northern Florida. Proc. of 26th Annual Southern Conservation Tillage Conference for Sustainable Agriculture, 8–9 June. North Carolina Agricultural Research Service Technical Bulletin TB-321: 256–265.
- Mulvey, R. H. 1974. Cone-top morphology of the white females and cyst of the genus *Heterodera* (subgenus *Heterodera*), a cyst-forming nematode. *Canadian Journal of Zoology* 52:77–81.
- Ntidi, K., H. Fourie, A. McDonald, D. De Waele, and C. Mienie. 2012. Plant parasitic nematodes associated with weeds in subsistence agriculture in South Africa. *Nematology* 14:875–887.
- Núñez, L., L. Flores, and D. Humphreys. 2021. First report of *Heterodera trifolii* on white clover and *Rumex obtusifolius* in Costa Rica. *Plant Disease* 105:230.
- Palomares, J. E., C. M. Oliveira, and V. Blok. 2014. Plant parasitic nematodes of potato. Pp. 148–166 in R. Navarre, and M. Pavet, eds. *The Potato: Botany, Production and Uses*. Boston: CAB International.
- Pérez E., D. Weingartner, and R. McSorley. 2000. Correlation between *Paratrichodorus* minor population levels and corky ringspot symptoms on potato. *Nematropica* 30:247–251.
- Ploeg, A., D. Robinson, and D. Brown. 1993. RNA-2 of Tobacco rattle virus encodes the determinants of transmissibility by trichodorid vector nematodes. *Journal of General Virology* 74:1463–1466.
- Plowright R.A., G. Caubel, K. A. Mizen. 2002. *Ditylenchus* species. Pp. 107–140 in J. L. Starr, R. Cook, and J. Bridge, eds. *Plant Resistance to Parasitic Nematodes*. Wallingford, UK: CAB International.
- Quénehervé, P., M. Godefroid, P. Mège, and S. Marie-Luce. 2011. Diversity of *Meloidogyne* spp. parasitizing plants in Martinique Island, French West Indies. *Nematropica* 41:191–199.
- Ravichandra, N. 2014. *Horticultural nematology*. New Dehli: Springer.
- R Core Team. 2017. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna,

- Austria. <https://www.R-project.org/>
- Rodríguez, A., N. Chaves, A. Hernández, and F. Herrera, F. 2015. Determinación de la resistencia al cletodim en *Poa annua* en Costa Rica. *Agronomía Mesoamericana* 257-266.
- Sandoval, R. 2015. Determinación molecular de *Pratylenchus* asociadas a cultivos agrícolas de Costa Rica. Tesis de Licenciatura, Universidad de Costa Rica, San José, Costa Rica.
- Sandoval, R. 2018. Frecuencia, distribución, identificación molecular y determinación de la variabilidad intraespecífica de *Globodera* spp. en fincas cultivadas con papa (*Solanum tuberosum* L.) en la provincia de Cartago. Tesis de Maestría, Universidad de Costa Rica, San José, Costa Rica.
- Sandoval, R., L. Flores, and D. Humphreys. 2020. Molecular characterization and distribution of *Globodera pallida* in the main potato production area of Costa Rica. *Nematropica* 50:218-228.
- Samaliev, H., and S. Kalinova. 2013. Host suitability of twelve common weeds to *Pratylenchus penetrans* and *Meloidogyne hapla* in potato fields of Bulgaria. *Bulgarian Journal Agricultural Science* 19:202-208
- Secretaría Técnica de Planificación Sectorial Agropecuaria, Costa Rica (SEPSA). 2018. Superficie y producción. Available from: <http://www.sepsa.go.cr/DOCS/BEA/BEA28.pdf>
- Shepherd, A. 1986. Extraction and estimation of cyst nematodes. Pp. 31-46 in J. Southey, ed. *Laboratory methods for work with plant and soil nematodes*. London, UK: Her Majesty's Stationery Office.
- Skantar, A., Z. Handoo, I. Zasada, R. Inghram, L. Carta, and D. Chitwood. 2011. Morphological and molecular characterization of *Globodera* populations from Oregon and Idaho. *Phytopathology* 101:480-491.
- Smiley, R. W., G. Yan, and J. Gourlie. 2014. Selected Pacific Northwest rangeland and weed plants as host of *Pratylenchus neglectus* and *P. thornei*. *Plant disease* 98:1333-1340.
- Villalobos, A. 1999. Control químico de *Polygonum aviculare* y otras malezas en el cultivo de la zanahoria (*Daucus carota* L.). Tesis de Licenciatura, Universidad de Costa Rica, San José, Costa Rica.
- Yeates, G., T. Bongers, R. De Goede, D. Freckman, and S. Georgieva. 1993. Feeding habits in soil nematode families and genera - an outline for soil ecologists. *Journal of Nematology* 25:315-331.

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