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Fifty new genera of Hesperiidae (Lepidoptera)

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Fifty new genera of Hesperiidae (Lepidoptera)

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Abstract. Genomic sequencing and analysis of worldwide skipper butterfly (Lepidoptera: Hesperiidae) fauna points to imperfections in their current classification. Some tribes, subtribes and genera as they are circumscribed today are not monophyletic. Rationalizing genomic results from the perspective of phenotypic characters suggests two new tribes, two new subtribes and 50 new genera that are named here: Ceratrichiini Grishin, trib. n., Gretnini Grishin, trib. n., Falgina Grishin, subtr. n., Apaustina Grishin, subtr. n., Flattoides Grishin, gen. n., Aurivittia Grishin, gen. n., Viuria Grishin, gen. n., Clytius Grishin, gen. n., Incisus Grishin, gen. n., Perus Grishin, gen. n., Livida Grishin, gen. n., Festivia Grishin, gen. n., Hoodus Grishin, gen. n., Anaxas Grishin, gen. n., Chiothion Grishin, gen. n., Crenda Grishin, gen. n., Santa Grishin, gen. n., Canesia Grishin, gen. n., Bralus Grishin, gen. n., Ladda Grishin, gen. n., Willema Grishin, gen. n., Argemma Grishin, gen. n., Nervia Grishin, gen. n., Dotta Grishin, gen. n., Lissia Grishin, gen. n., Xanthonymus Grishin, gen. n., Cerba Grishin, gen. n., Avestia Grishin, gen. n., Zetka Grishin, gen. n., Turmosa Grishin, gen. n., Mielkeus Grishin, gen. n., Coolus Grishin, gen. n., Daron Grishin, gen. n., Barrolla Grishin, gen. n., Brownus Grishin, gen. n., Tava Grishin, gen. n., Rigga Grishin, gen. n., Haza Grishin, gen. n., Dubia Grishin, gen. n., Pares Grishin, gen. n., Chitta Grishin, gen. n., Artonia Grishin, gen. n., Lurida Grishin, gen. n., Corra Grishin, gen. n., Fidius Grishin, gen. n., Veadda Grishin, gen. n., Tricrista Grishin, gen. n., Viridina Grishin, gen. n., Alychna Grishin, gen. n., Ralis Grishin, gen. n., Testia Grishin, gen. n., Buzella Grishin, gen. n., Vernia Grishin, gen. n., and Lon Grishin, gen. n. In addition, the following taxonomic changes are suggested. Prada Evans is transferred from Hesperiinae to Trapezitinae. Echelatus Godman and Salvin, Systaspes Weeks, and Oenides Mabille are removed from synonymy and are treated as valid genera. The following genera are new junior subjective synonyms: Tosta Evans of Eantis Boisduval; Turmada Evans of Neoxeniades Hayward, Arita Evans of Tigasis Godman, and Alera Mabille of Perichares Scudder. Eantis pallida (R. Felder) (not Achlyodes Hübner), Gindanes kelso (Evans) (not Onenses Godman and Salvin), Isoteinon abjecta (Snellen) (not Astictopterus C. and R. Felder), Neoxeniades ethoda (Hewitson) (not Xeniades Godman), Moeris anna (Mabille) (not Vidius Evans), and Molo pelta Evans (not Lychnuchus Hübner) are new genus-species combinations. The following are species-level taxa: Livida assecla (Mabille) (not a subspecies of Livida grandis (Mabille), formerly Pythonides Hübner) and Alychna zenus (E. Bell) (not a junior subjective synonym of Alychna exclamationis (Mabille), formerly Psoralis Mabille); and Barrolla molla E. Bell (formerly Vacerra Godman) is a junior subjective synonym of Barrolla barroni Evans (formerly

^{*}these two authors contributed equally to this work

Paratrytone Godman). All these changes to taxonomic status of names are propagated to all names currently treated as subspecies (for species), subgenera (for genera) and synonyms of these taxa. Finally, taxa not mentioned in this work are considered to remain at the ranks and in taxonomic groups they have been previously assigned to.

Key words. Genomics, higher classification, taxonomy, biodiversity, phylogeny.

Introduction

Hesperiidae, commonly known as skippers, are a charismatic group of butterflies that frequently look moth-like due to their stout bodies (Watson 1893), mostly brown and gray colors and fast wing beats. Some are crepuscular and even come to light (Austin 2008). However, recent DNA-based studies argue that they may have originated deep within butterfly radiation, after the swallowtails (Papilionidae) have split from the common ancestor (Wahlberg et al. 2005; Kawahara and Breinholt 2014; Espeland et al. 2018).

Having worldwide distribution, Hesperiidae are highly speciose with more than 3500 species described. However, they have received less attention than other butterfly families. The major milestone works remain those of Evans, who offered a comprehensive taxonomic treatment of the group in six volumes (Evans 1937, 1949, 1951, 1952, 1953, 1955). Refinement of this classification was catalyzed by new methodologies. The groundbreaking work of Warren et al. (2008, 2009) re-shaped the higher classification based on combination of DNA sequences of several genes with morphological characters. Several follow up studies employed a larger set of genes, up to several hundred (Sahoo et al. 2016, 2017; Toussaint et al. 2018). Then, a genome-scale revisionary work has followed (Li et al. 2019; Zhang et al. 2019b, c).

Genomic analyses revealed many nuances not readily apparent from a morphological standpoint. However, retrospective comparison of phenotypic characters is consistent with the picture emerging from the comparison of phylogenetic trees based on nuclear and mitochondrial genomes (Li et al. 2019; Zhang et al. 2019c). We carried out genomic sequencing of representative Hesperiidae species from all known genera. As a result, we found inconsistencies in assignment of species to genera, and many genera and some tribes and subtribes were not monophyletic as currently defined. While the details of the phylogenetic analysis and their implications for the higher classification of Hesperiidae will be presented elsewhere, some of these inconsistencies are corrected here by proposing new names for two tribes, two subtribes and 50 genera detected in phylogenetic trees. Here, we show only a subset of data necessary to justify our conclusions.

Materials and Methods

This study is based on whole genomic shotgun DNA sequences that were mostly obtained from pinned and dried specimens in collections. Many of these specimens were collected more than a century ago and a number of them were primary type specimens. See Table 1 for a brief data summary of the specimens used in this work, and Table S1 in the Supplemental file deposited at https://osf.io/5cfht/?view_only=21 eb53b6f8f344afaee3de2be90bf5d2 for details. We use either an abdomen, pieces of muscle tissue taken from the thorax through the abdomen attachment site (for previously dissected specimens), or a leg for DNA extraction. The abdomen is used when genitalic dissection is needed. Muscle tissue is a viable alternative to the leg when leg material is not sufficient or using a leg is not possible. Legs were the most convenient choice because they were easier to sample and often yielded better-quality DNA. Therefore, most specimens were sequenced from legs. The details of protocols for DNA extraction, genomic library preparation, sequencing and analysis are given in our previous publications (Cong et al. 2015a, b, 2016a, b, 2017a, b, 2018; Shen et al. 2015, 2016a, b, 2017; Zhang et al. 2017a, b; Li et al. 2019; Zhang et al. 2019a, b). In the methods employed, this study is identical to that of Li et al. (2019), and only difference is that a larger number of species and specimens were used.

Due to this very large number of specimens we have sequenced, the phylogenetic trees were built for smaller phylogenetic groups (subfamilies, tribes and subtribes) and analyzed together with the "backbone" tree for the entire family constructed with selected reference species and given in Li et al. (2019). Type species (or their close relatives when the type is not available) of available genus-level names (including the names treated as synonyms) were marked on the trees to ensure that any available name for each

clade is used. Manual analysis of the trees involved searching for non-monophyletic genera and checking clades that are prominently separated from others. Both nuclear genome and mitogenome trees were checked side-by-side to evaluate the consistency between them. Statistical support values (bootstrap) were taken into consideration to judge the validity of each observed clade. When a clade without an available name was found, identification of specimens in the clade was confirmed by the analysis of their wing patterns and genitalia. These new clades and the genus of former placement of each species proposed as a type species of a new genus name were rationalized in terms of genitalic morphology and wing patterns to search for diagnostic characters in phenotype. Finally, genitalia and wing patterns were used to decide the generic placement of species for which DNA data were not available.

Diagnostic DNA characters were identified in nuclear genomic sequences using our recently published procedure (see SI Appendix to Li et al. 2019). The positions in exons were found that are most likely synapomorphic to the clade defined as a genus. For the clades where we had several species sequenced, positions that are invariant in all species from this clade and have a base pair different from a (mostly invariant) base pair in the outgroups were found and those with the smallest number of species with missing data were selected. If a genus has only one species sequenced, it is difficult to distinguish between characters of the genus and characters of the species. Therefore, we frequently resorted to a different method of defining DNA characters of a genus that would increase the robustness of these characters. First, we looked for characters to define the sister clade of the genus. Sister clades usually included more than one species, and thus characters for the sister clade were better defined. We find the characters for the sister clade and take their states that differ from those in the sister clade as diagnostic for the genus in question. Second, we found synapomorphic characters for the clade that leads to the common ancestor of the genus and its sister clade. We used the combination of these latter characters with the diagnostic ones to define the genus. Such a treatment increases the chances that the character found is not a random non-conserved change or a sequencing error. Number of sequence reads covering this position was taken into account in choosing the characters, and those positions with better coverage were given priority. The character states are given in diagnoses below as abbreviations. For example, aly728.44.1:G672C means position 672 in exon 1 of gene 44 from scaffold 728 of the Cecropterus [formerly Achalarus] lyciades (aly) reference genome (Shen et al. 2017) is C, changed from G in the ancestor. When characters were found for the sister clade of the diagnosed taxon, the following statement was used: aly5294.20.2:A548A (not C), which means that position 547 in exon 2 of gene 20 on scaffold 5294 is occupied by the ancestral base pair A, which was changed to C in the sister clade (so it is not C in the diagnosed taxon). The sequences of exons from the reference genome with the positions used as character states highlighted in green are given in the supplemental file deposited at https://osf.io/5cfht/?view_only=21eb53b6f8f344afaee3de2be90 bf5d2. Distribution of these sequences together with this publication ensures that the numbers given in the diagnoses can be easily associated with actual sequences. This publication has been registered with ZooBank as http://zoobank.org/BA35690A-FC73-4E5A-A805-FE9550275FEC and individual ZooBank registration numbers for each new name are given below.

The specimens were examined and sampled for sequencing in the following collections (abbreviations in parenthesis, used in Table 1 and Table S1 in the Supplemental file deposited at https://osf.io/5cfht/?view onl y=21eb53b6f8f344afaee3de2be90bf5d2): American Museum of Natural History, New York, NY, USA (AMNH), Natural History Museum, London, UK (BMNH), Burke Museum of Natural History and Culture, Seattle, WA, USA (BMUW), Carnegie Museum of Natural History, Pittsburgh, PA, USA (CMNH), Colorado State University Collection, Fort Collins, CO, USA (CSUC), The Field Museum of Natural History, Chicago, FL, USA (FMNH), Los Angeles County Museum of Natural History, Los Angeles, CA, USA (LACM), Museum of Comparative Zoology, Harvard University, Cambridge, MA, USA (MCZ), Mississippi Entomological Museum, Starkville, MS, USA (MEM), Muséum National d'Histoire Naturelle, Paris, France (MNHP), Natural History Museum, Frankfurt, Germany (SMF), Texas A&M University Insect Collection, College Station, TX, USA (TAMU), National Museum of Natural History, Smithsonian Institution, Washington, DC, USA (USNM), University of Texas Southwestern, freezers of the Grishin lab, Dallas, TX, USA (UTSW), Zentrum fur Biodokumentation des Saarlandes, Schiffweiler, Germany (ZfBS), Museum für Naturkunde, Berlin, Germany (ZMHB), Zoologische Staatssammlung München, Germany (ZSMC), and research collections of Jim P. Brock (JPBrock), Ernst Brockmann (EBrockmann), Robert Gallardo (RGallardo), Bernard Hermier (BHermier), Kiyoshi Maruyama (KMaruyama) James A. Scott (JAScott), Texas Lepidoptera Survey, Houston, TX, USA (TLS, since then donated to McGuire Center for Lepidoptera and Biodiversity, Gainesville, FL, USA, MGCL), and Mark Walker (MWalker).

Results and Discussion

Analysis of the trees revealed rampant inconsistencies in the current classification of Hesperiidae into genera. These inconsistencies were not evenly distributed among phylogenetic lineages of Hesperiidae. Fewer problems were observed among the Old World taxa due to better knowledge about them, and maybe due to their smaller number. Most problems involved non-monophyletic genera with one or more clades of the former genus not having an available name. In several instances, a genus remained monophyletic as currently circumscribed but was prominently divided into two clades. Genetic diversification within each clade was comparable to that in other genera. For instance, the COI barcode difference taken as a proxy for diversification was typically below 10%, and the distances in nuclear and mitochondrial genomic DNA trees were about the same as in other species-rich genera. However, the distance between the clades was prominently larger than within each clade, typically more than 10% identity in the barcode.

These clades are named here and the following are the standardized descriptions of new taxa found during this analysis. The names were chosen to be simple and short, and mostly either reflect names or properties of their type species to facilitate memorization, or are fusions of genus names, euphonized and shortened. For each genus, a ZooBank registration number is given. The type species name is listed in its original genus combination and spelling, followed by the author and year the name was made available (not a bibliographic reference, but part of the name). Definition section indicates closest genera, mostly sisters or a group of genera if there are no clear sister genera, states the generic placements prior to this study (type species are given where appropriate to help assign a clade to a genus), gives reference to diagnostic characters as they are given in previous publications mostly in the Evans volumes (Evans 1937, 1949, 1951, 1952, 1953, 1955). It was almost always possible to trace the genus observed in the phylogenetic trees to the morphological characters given in the Evans' identification keys. We think that referencing the keys rather than comprehensively listing all the characters would facilitate identification. Nevertheless, a brief morphological diagnosis for each genus is given, summarizing the most prominent phenotypic traits. DNA characters found by our recently developed method (Zhang et al. 2019c) are given at the end of each definition. This method should increase the robustness of the generic-level characters allowing for addition of species to a genus once they are sequenced. Then the gender of the name and an explanation about its origin is provided, species placed in the genus are listed (in their original genus-species name combinations with authors and dates), a parent taxon (mostly a subtribe, tribe or subfamily per Li et al. (2019)) is given, sometimes followed by comments about species involved and proposed additional taxonomic changes.

All of the changes to taxonomic status of names are propagated to all names currently treated as subspecies (for species), subgenera (for genera) and synonyms of these taxa. For instance, we do not list subspecies and synonyms unless changes are made to their status, and we assume that these names go with the species they were previously placed with. Finally, taxa not mentioned in this work are considered to remain at the ranks and in taxonomic groups they have been previously assigned to (Evans 1937, 1949, 1951, 1952, 1953, 1955; Mielke 2005; Li et al. 2019; Zhang et al. 2019b).

Furthermore, several tribes and subtribes would no longer be monophyletic if the higher-level classification remained unchanged. To solve this problem, we establish two new tribes and two new subtribes, which are described here. The following sections follow the standardized format and are either new taxa descriptions or taxonomic changes to existing taxa, as stated in the titles of these sections. Sections dealing with family-group names are given first, so that the new names can be used in the following sections dealing with genus-group names. Then, the sections are arranged in phylogenetic order approximating how these taxa appear in the figured trees.

Ceratrichiini Grishin, new tribe

http://zoobank.org/FC72EB8A-495D-4439-AA53-C310CD0735DE

Type genus. Ceratrichia Butler, [1870].

Definition. Placed near the mostly African tribe Astictopterini Swinhoe, 1912, this tribe is a sister to all Hesperiinae except Aeromachini Tutt, 1906. Backed by the maximal statistical support in all trees (Fig. 1), this clade of closely related genera keys to VI.A.(b) or VII.B.(a)(a¹) in Evans (1937). Antennae long, longer than half of costa, 2nd segment of palpi directed up, forewing vein CuA_2 originates in the middle of discal cell, end of discal cell straight, hindwing vein M_2 prominent, originates closer to vein M_1 than M_3 , vein CuA_2 originates before or opposite to RS, vein 3A long. Male genitalia simple in most species: uncus undivided (except Herila), terminally narrows to a point, narrow in lateral view, gnathos either short or lacking, valva 2-3 times longer than wide, without prominent elaborations, may have small teeth on harpe. In DNA, a combination of the following base pairs is diagnostic: aly1121.3.2:A429G, aly669.27.2:A50G, aly374.13.3:A242T, aly216.78.1:A568C, aly1155.14.6:T406G.

Genera included. *Ceratrichia* Butler, [1870], *Meza* Hemming, 1939, *Herila* Larsen and Collins, 2012, *Pardaleodes* Butler, 1870, *Ankola* Evans, 1937, and a new genus described below.

Parent taxon. Subfamily Hesperiinae Latreille, 1809.

Comments. Judging from the genomic trees, this tribe consists of closely related species, but their wing patterns and colors are quite different from each other and may resemble genera outside the tribe. We were not able to find an obvious morphological synapomorphy for the tribe and, as it frequently is the case, the tribe is best diagnosed by DNA characters.

Gretnini Grishin, new tribe

http://zoobank.org/E7F984DE-BC49-4617-8624-9630FCEE18F3

Type genus. Gretna Evans, 1937.

Definition. Placed as a sister to all other Hesperiinae but Aeromachini Tutt, 1906, Ceratrichiini trib. n. and Astictopterini Swinhoe, 1912 in all genomic trees, this stand-alone lineage has no close relatives (Fig. 1). Keys to VIII.B.(b)(b¹)(a²)(a³)(a⁴) in Evans (1937), and the description of *Gretna* given by Evans (1937: 149) applies to the tribe. Most prominently it is distinguished from other Hesperiinae by a combination of large stout palpi with tiny 3rd segment, curved forewing vein 1A+2A, well-defined hindwing vein M_2 , narrow hindwing cells M_1 - M_2 and M_2 - M_3 around it and unusual vein structure at the origin of vein RS in males, which is close to the origin of vein Sc+R₁, and veins M_1 and RS are curved at their divergence point, U-shaped rather than V-shaped. In DNA, a combination of the following base pairs is diagnostic: aly2012.5.1:A98C, aly2195.3.11:A37G, aly349.39.4:C152G, aly276378.25.8:A84T, aly6398.6.4:A46C.

Genera included. Only the type genus.

Parent taxon. Subfamily Hesperiinae Latreille, 1809.

Comments. Uniqueness of *Gretna* as a stand-alone phylogenetic lineage of the tribal rank was not readily apparent from morphology and the genomic approach was critical in revealing its distinctness.

Falgina Grishin, new subtribe

http://zoobank.org/B32D24C6-8946-44DC-BFCF-15E5582B3ABE

Type genus. Falga Mabille, 1898.

Definition. An assemblage of genera previously placed in three subtribes (Carystina Mabille 1878, Anthoptina A. Warren, 2009 and Moncina A. Warren, 2008), the subtribe is characterized by the maximal statistical support values in all genomic trees (Fig. 1) and is in the same clade with Anthoptina and Moncina. Keys to I.1, I.2, J.8, J.13., J.16c, J.31, J.47, J.49, or K.1 in Evans (1955). The lack of spines on mid-tibiae is the unifying morphological character of this morphologically diverse group best diagnosed by the combination of the following synapomorphic DNA characters: aly1186.4.1:A962T, aly48.2.12:C1493A, aly345.16.5:A563T, aly536.1.3:A312G, aly276378.18.1:A1489G.

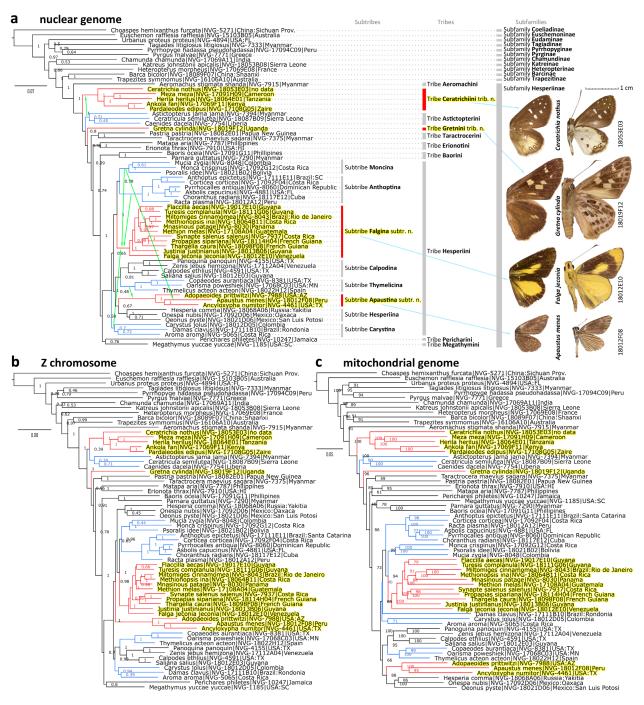


Figure 1. Genomic trees of representative Hesperiidae. The trees are built from protein-coding regions in different genomic partitions: a) Nuclear genome; b) Z chromosome; c) Mitochondrial genome. The trees are rooted with *Pterourus glaucus* (NVG-1670), not shown. See Table 1 and Table S1 in the Supplemental file deposited at https://osf.io/5cfht/?view_only=21eb53b6f8f344afaee3de2be90bf5d2 for additional data about these specimens. Names of species placed in new tribes and subtribes described in this work are highlighted in yellow and clades representing new taxa are colored in red. Clades for tribes and subtribes where species of the new taxa were placed previously are colored in blue and green arrow points from the former taxon to the new taxon (only on nuclear genome tree). Statistical support values are shown by nodes in all but the COI barcode trees. COI barcode NJ dendrogram is given for comparison and is not expected to reflect phylogeny. Subfamilies, tribes and subtribes for species included in the trees are shown to the right of the nuclear tree. Sequenced specimens of the type species of the new tribes and subtribes are illustrated in dorsal (left) and ventral (right) views and indicated by blue arrows.

Genera included. Falga Mabille, 1898, Justinia Evans, 1955, Thargella Godman, 1900, Propapias O. Mielke, 1992, Synapte Mabille, 1904, Turesis Godman, 1901, Flaccilla Godman, 1901, Methion Godman, 1900, Mnasinous Godman, 1900, Miltomiges Mabille, 1903, Methionopsis Godman, 1901 and a new genus described below.

Parent taxon. Tribe Hesperiini Latreille, 1809.

Comments. This subtribe is yet another unexpected assemblage of genera with disparate morphology. However, its monophyly is very strongly supported in all trees (Fig. 1). No phenotypic synapomorphy is apparent to unify these taxa, and the ultimate diagnosis is possible on the basis of synapomorphic DNA characters.

Apaustina Grishin, new subtribe

http://zoobank.org/05EF9D2F-DACA-45DC-B6D7-7960424D3C33

Type genus. Apaustus Hübner, [1819].

Definition. A subtribe without clear phylogenetic affinities within Hesperiini, but not a sister to Thymelicina (Fig. 1). Keys to J.1, M.1, or M4 or in Evans (1955). Characterized by elongated wings, weak flight, short antennae and gracile bodies. Diagnosed by a combination of: flattened antennal club without apiculus, long and thin 3rd segment of palpi extending beyond the 2nd segment (less so in *Adopaeoides*), the lack of spines on mid-tibiae (except *Apaustus*), and the lack of brands or stigmas on wings. In DNA, a combination of the following base pairs is diagnostic: aly3507.5.1:A578C, aly1297.14.4:A4190C, aly123.4.7:A70T, aly123.4.7:G71C, aly315.4.4:A453C.

Genera included. Apaustus Hübner, [1819], Adopaeoides Godman, 1900, and Ancyloxypha C. Felder, 1862.

Parent taxon. Tribe Hesperiini Latreille, 1809.

Comments. Our genomic findings corroborate recent anchored phylogenomic results (Toussaint et al. 2018) in dividing the former Thymelicina into two phylogenetic lineages of different origins (Fig. 1). While the association of *Apaustus*, formerly placed in Moncina A. Warren, 2008, with the other two genera formerly placed in Thymelicina Tutt, 1905, was unexpected at first, it makes morphological sense considering similarities in wing shapes and the gracile bodies of these butterflies.

Composition of the subtribes Calpodina Clark, 1948 and Thymelicina Tutt, 1905

Separated from Carystina Mabille 1878 in Li et al. (2019), Calpodina is a sister and a close relative of Thymelicina (Fig. 1). In addition to the type genus *Calpodes* Hübner, [1819], we keep three genera in this subtribe: *Saliana* Evans, 1955, *Panoquina* Hemming, 1934, and *Zenis* Godman, 1900. All other genera previously placed in Calpodina (Warren et al. 2008, 2009) belong to Carystina or other subtribes as detailed in Li et al. (2019) and this work. Its sister Thymelicina, after removal of the two genera being placed in the new subtribe described above, is composed of only three genera: *Thymelicus* Hübner, [1819], *Oarisma* Scudder, 1872, and *Copaeodes* Speyer, 1877.

Flattoides Grishin, new genus

http://zoobank.org/9D7A5387-08F8-4C25-B584-B67F90EEC43A

Type species. Codatractus amazonensis Bell, 1947.

Definition. A sister genus to *Oileides* Hübner, [1825] (type species *Oileides vulpinus* Hübner, [1825]), where *C. amazonensis* was placed previously (Fig. 2). However large genetic distance between *O. vulpinus* and *O. amazonensis* (COI barcodes differ by 12.3%, the difference is typically smaller than 10% for species within a genus) argues for their distinction. Keys to D.9.4 in Evans (1952). Genitalia illustrated by Bell in his fig. 2 (Bell 1947). Distinguished from its relatives by the following combination of characters: shorter than tegumen and broad uncus with small (not longer than wide) arms; prominent gnathos not

shorter than uncus; valva gradually curved dorsad, ending with two pointed teeth; tuft of apricot-colored modified scales in a groove along the vein near the base of ventral hindwing in males; forewing with a macular white band from mid-costa to tornus. In DNA, a combination of the following base pairs is diagnostic: aly8661.4.1:A916G, aly10226.44.1:C1975G, aly709.1.2:T194C, aly36556.1.1:T2416T (not A), aly274.33.1:A188A (not G), aly82.28.5:T398T (not G).

Etymology. The name is a masculine noun in the nominative singular, for the type species that resembles butterflies from the genus *Celaenorrhinus* known as "Flats" in some English-speaking countries.

Species included. Only the type species.

Parent taxon. Subtribe Oileidina Grishin, 2019.

Aurivittia Grishin, new genus

http://zoobank.org/C2C0299C-67C8-4F0E-ACB8-29A7292E10BD

Type species. Plesioneura aurivittata Moore, 1878.

Definition. Surprisingly, a possible sister genus to Alenia Evans, 1935 (type species: Pyrgus sandaster Trimen, 1868), and in the same clade with Apallaga Strand, 1911 (type species Apallaga separata Strand, 1911), but genetically far removed from these and all other taxa (Fig. 3). Species in this genus were formerly placed in Celaenorrhinus Hübner, [1819] (type species Papilio eligius Stoll, [1781];) and key to B.6.30 in Evans (1949), notably lacking hair pencil on hind tibiae, which other Celaenorrhinus-like taxa

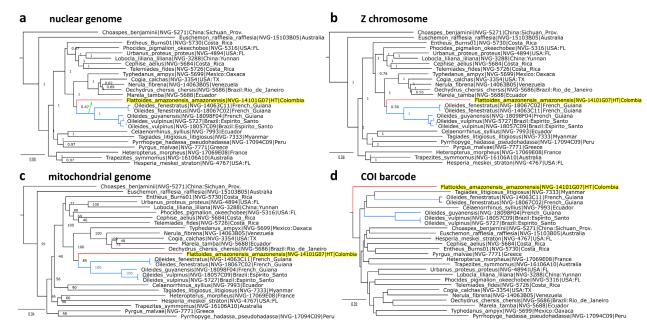


Figure 2. Genomic trees of Eudaminae and representatives of other Hesperiidae subfamilies. The trees are built from protein-coding regions in different genomic partitions: a) Nuclear genome. b) Z chromosome. c) Mitochondrial genome; d) COI barcode. The trees are rooted with *Pterourus glaucus* (NVG-1670), not shown. See Table 1 and Table S1 in the Supplemental file deposited at https://osf.io/5cfht/?view_only=21eb53b6f8f344afaee3de2be90bf 5d2 for additional data about these specimens. Where possible, taxa are ordered similarly to that in the nuclear genome tree. Names of species placed in new genera described in this work are highlighted in yellow and clades of these genera are colored in red. Clades for genera where these species were placed previously are colored in blue and green arrow points from the former genus to the new genus (only on nuclear genome tree). Names of genera resurrected from synonymy are highlighted in cyan. Names of genera that form new genus-species combinations proposed in this work are highlighted in magenta. Statistical support values are shown by nodes in all but the COI barcode trees. COI barcode NJ dendrogram is given for comparison and is not expected to reflect phylogeny. The same notations are used in Fig. 2–13.

possess. Morphologically, distinguished from *Celaenorrhinus* (sensu stricto) by essentially undivided uncus, sometimes with small knobs (not processes) as arms and the lack of hair pencil on hind tibiae; and from *Alenia* (a genus with undivided uncus) by the shape of valva that is not terminally split like a crab claw, and the presence of yellow band across the forewing. In male genitalia most similar to *Apallaga*, a genus with many species characterized by undivided uncus, but differs in longer gnathos (not less than half of the uncus length), thicker penis, valva with more robust and broad harpe that is gradually curved dorsad (not ventrad), does not carry any processes and is not forked, but narrows to a single point, and ampulla with a long process (style) along the harpe. In wing patterns, hindwing is not prominently variegated dorsally and is mostly unmarked, forewing with a compact yellow discal band with regular edges that does not extend into the discal cell and frequently with a triplet of apical yellow spots. In DNA, a combination of the following base pairs is diagnostic: aly2532.2.1:T488A, aly997.12.1:C310A, aly527.19.4:T178A, aly822.15.1:C589A, aly235.8.18:G1178A.

Etymology. The name is a feminine noun in the nominative singular, formed to reflect the golden stripe on the forewing, similar to the name of the type species.

Species included. The type species, *Plesioneura cameroni* Distant, 1882, and *Celaenorrhinus vietnamicus* Devyatkin, 1998.

Parent taxon. Tribe Celaenorrhinini Swinhoe, 1912.

Comments. These south Asian butterflies are not so prominently distinct from the striped species of *Celaenorrhinus*. Therefore, their uniqueness was not apparent prior to our genomic study, although genitalic features revealed *a posteriori* can diagnose the genus morphologically. Phylogenetic placement of this Asian genus in the African clade of Celaenorrhinini Swinhoe, 1912 is interesting.

Viuria Grishin, new genus

http://zoobank.org/1D91E55F-38B7-4E1E-997F-49FBFD10CE02

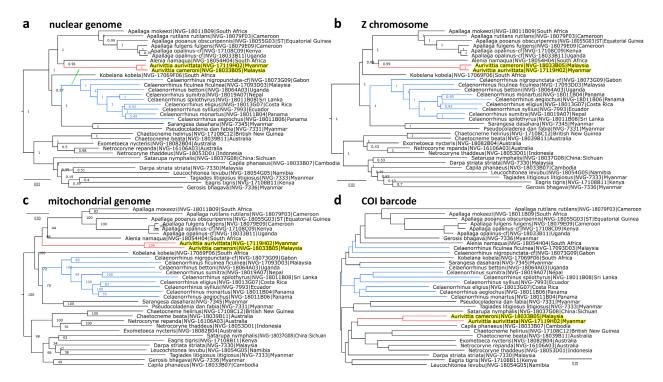


Figure 3. Genomic trees of Tagiadinae. See Fig. 2 legend for notations.

Type species. Pellicia licisca Plötz, 1882.

Definition. A sister genus to Viola Evans, 1953, distant from Pachyneuria Mabille, 1888 (type species Pachyneuria obscura Mabille, 1888), where species of this genus were formerly placed (Fig. 4). Keys to E.20.7 in Evans (1953). Differs from Pachyneuria by the presence of long tufts of scales near hindwing costa above in males and swollen hindwing vein Sc+R₁, similar to Viola and Nisoniades Hübner, [1819], among others. Differ from Viola and other genera in genitalia and diagnosed by asymmetric uncus with asymmetric processes, reduced harpe of the right valva compared to the expanded rounded ampulla. In DNA, a combination of the following base pairs is diagnostic: aly1222.46.1:A28C, aly207.4.6:C1025G, aly1019.7.7:T59A, aly2012.7.7:C98G, aly638.27.5:T694A.

Etymology. The name is a feminine noun in the nominative singular, a fusion of Vi[ola] and [Pachyne] uria. Species included. The type species, $Pachyneuria\ lista$ Evans, 1953, and $Pellicia\ herophile$ Hayward, 1940. Parent taxon. Tribe Carcharodini Verity, 1940.

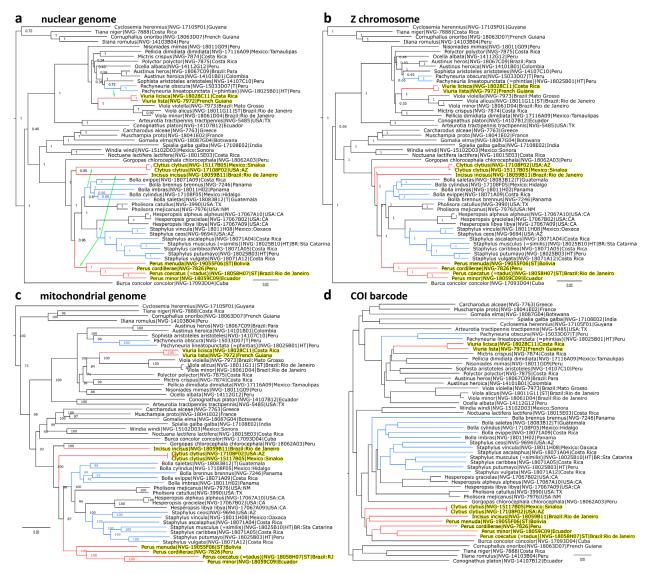


Figure 4. Genomic trees of Carcharodini. See Fig. 2 legend for notations.

Clytius Grishin, new genus

http://zoobank.org/AEAE278A-4A9B-45C3-9903-E886FBCEA5F7

Type species. Pholisora clytius Godman and Salvin, [1897].

Definition. Not closely related to any other genus. Formerly placed in *Bolla* (type species *Bolla pullata* Mabille, 1903, currently a junior subjective synonym of *Staphylus imbras* Godman and Salvin, [1896]), but is not monophyletic with *Bolla* species (Fig. 4). Keys to E.31.22 in Evans (1953). Morphologically, distinguished from *Bolla* species by broad-ended valva without processes, but with concave costa ending with bulky ampulla weakly separated from short serrated harpe, and the lack of gray or ochreous overscaling on wings above. In DNA, a combination of the following base pairs is diagnostic: aly997.12.1:G125C, aly536.106.2:A2067G, aly127.74.3:A1567G, aly320.9.2:A806G, aly1011.11.1:A34C.

Etymology. The name, a masculine noun in the nominative singular, echoes the type species name.

Species included. Only the type species.

Parent taxon. Tribe Carcharodini Verity, 1940.

Incisus Grishin, new genus

http://zoobank.org/2ED74AC8-4CCF-4796-952B-0292620F0855

Type species. Antigonus incisus Mabille, 1878.

Definition. A genus without close relatives. Formerly placed in *Staphylus* Godman and Salvin, [1896], *incisus* is not monophyletic with the *Staphylus* type species *Helias ascalaphus* Staudinger, 1876, nor with *Scantilla opites* Godman and Salvin, [1896] (a junior subjective synonym of *Tagiades vincula* Plötz, 1886) the type species of *Scantilla* Godman and Salvin, [1896], a genus-group name treated as a subjective synonym of *Staphylus* (Fig. 4). Keys to E.32.35b in Evans (1953). Readily distinguished from its relatives by the wing shape: forewing inner margin concave, hindwing outer margin excavate; forewing with hyaline apical spots; uncus long and narrow, pointed at the tip, without side processes, no defined gnathos, valva less than twice longer than wide, penis shorter that in *Staphylus*, with a spined cornutus. In DNA, a combination of the following base pairs is diagnostic: aly23605.8.3:C247T, aly5294.23.1:A538C, aly9588.6.1:T843C, aly1370.7.2:A1760G, aly345.13.8:A62C.

Etymology. The name, a masculine noun in the nominative singular, echoes the type species name.

Species included. The type species, *Staphylus fasciatus* Hayward, 1933 and *Pholisora* (?) *angulata* Bell, 1937.

Parent taxon. Tribe Carcharodini Verity, 1940.

Comments. Steinhauser doubted the placement of some of these species in *Staphylus* on the basis of morphological analysis (Steinhauser 1989), and he was correct.

Perus Grishin, new genus

http://zoobank.org/349FEEDB-0503-4302-AA25-B985D8A8CB45

Type species. Pholisora cordillerae Lindsey, 1925.

Definition. Not closely related to any other genus. Formerly placed in *Staphylus* Godman and Salvin, [1896], *cordillerae* is not monophyletic with the *Staphylus* type species *Helias ascalaphus* Staudinger, 1876, nor with *Scantilla opites* Godman and Salvin, [1896] (a junior subjective synonym of *Tagiades vincula* Plötz, 1886) the type species of *Scantilla* Godman and Salvin, [1896], a genus-group name treated as a subjective synonym of *Staphylus* (Fig. 4). Keys to E.32.16, E.32.24b, E.32.26, or E.32.34 in Evans (1953). Distinguished from *Staphylus* species by the following combination of characters: head and palpi

above mostly brown, more prominent submarginal pale spots on both wings above, harpe shorter than or the same length as moderately expanded ampulla, directed posteriad, end of uncus pointed, tegumen expanded on the sides, bulbous, with a pair of side processes in some species. In DNA, a combination of the following base pairs is diagnostic: aly3014.2.4:A833G, aly9588.14.1:C77A, aly171.12.3:A1360C, aly2532.10.1:A1294C, aly2790.11.3:A787C.

Etymology. The name is a masculine noun in the nominative singular, derived from Peru, the locality of the type species and the county where many other species of this genus occur.

Species included. The type species, *Staphylus mossi* Evans, 1953, *Staphylus minor* Schaus, 1902, *Antigonus coecatus* Mabille, 1891, and *Nisoniades menuda* Weeks, 1902.

Parent taxon. Tribe Carcharodini Verity, 1940.

Tosta Evans, 1953 is a synonym of Eantis Boisduval, 1836

Phylogenetic analysis confidently reveals (Fig. 5) that *Tosta* Evans, 1953 (type species *Tosta tosta* Evans, 1953) originates within *Eantis* Boisduval, [1836] (type species *Urbanus thraso* Hübner, [1807]) and therefore is best considered its junior subjective synonym.

Eantis pallida (R. Felder, 1869), new combination

In all trees, *Helias pallida* R. Felder, 1869 is not monophyletic with *Papilio busirus* Cramer, [1779], the type species of *Achlyodes* Hübner, [1819], but originates within *Eantis* Boisduval, [1836] (type species *Urbanus thraso* Hübner, [1807]) (Fig. 5). Therefore, *H. pallida* is placed in this genus to form a new combination: *Eantis pallida*.

Pythonides assecla Mabille, 1883, reinstated status

We consider *Pythonides assecla* Mabille, 1883 to be a species distinct from *Pythonides grandis* Mabille, 1878 due to difference in size and extent of the blue coloration of the hindwing, both in males and females, and differences in shade of blue, being paler in *P. grandis*.

Livida Grishin, new genus

http://zoobank.org/24179CB6-20EF-4DF2-B545-CFF25554444C

Type species. Pythonides assecla Mabille, 1883.

Definition. A genus clearly near *Gindanes* Godman and Salvin, 1895, *Pythonides* Hübner, [1819], *Quadrus* Lindsey, 1925, *Zera* Evans, 1953 and *Ouleus* Lindsey, 1925, but without closer affinity to any one of them (Fig. 5). The species included here in this genus were formerly placed in *Pythonides*, but are not monophyletic with the type species of this genus, *Papilio jovianus* Stoll, 1782. Keys to E.41.5 in Evans (1953). Morphologically, distinguished from related genera by the combination of the shape of uncus (flattened at the tip with a short and thin central projection) with the shape of valva (broad at the base with concave costa and diamond-shaped harpe longer than the rest of the valva, with serrated dorsoposterior edge), only one upper spur on hind tibiae, the lack of tuft in hind tibiae and thoracic pouch in males, ventral hindwing mostly pale blue, dorsal hindwing pale blue in males at least near tornus. In DNA, a combination of the following base pairs is diagnostic: aly1672.3.1:T128G, aly1672.3.1:C110G, aly2618.5.1:C3626G, aly140.1.2:T1822T (not A), aly1349.7.9:C1010C (not G), aly1877.13.1:A785A (not C), aly23605.1.46:T5641T (not A), aly23605.1.46:C5642C (not G), aly536.102.1:C56C (not A), aly1935.2.1:G242G (not C), aly345.3.1:C328C (not G), aly767.12.13:C118C (not A).

Etymology. The name is a feminine noun in the nominative singular, from the Latin lividus (blue), to indicate the distinctive cornflower blue color of the hindwing in males of the type species.

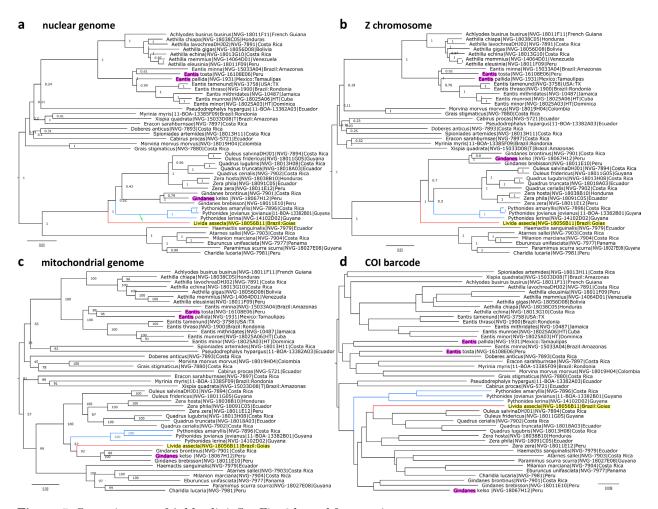


Figure 5. Genomic trees of Achlyodini. See Fig. 2 legend for notations.

Species included. The type species and *Pythonides grandis* Mabille, 1878.

Parent taxon. Subtribe Pythonidina Grishin, 2019.

Comments. Another genus that would be very difficult to detect without genomic data, because by appearance the adults of this species do not look noticeably different from *Pythonides*. However, genomic analysis suggests that the two are distant.

Gindanes kelso (Evans, 1953), new combination

Phylogenetic analysis shows (Fig. 5) that *Onenses kelso* Evans, 1953 falls within *Gindanes* Godman and Salvin, [1895] (type species *Gindanes panaetius* Godman and Salvin, [1895], treated as a subspecies of *Hesperia brebisson* Latreille, [1824]) and in a different tribe (Achlyodini) from *Onenses* Godman and Salvin, [1895] (type species *Leucochitonea hyalophora* R. Felder, 1869, tribe Pyrgini). Therefore, we introduce a new combination: *Gindanes kelso*.

Festivia Grishin, new genus

http://zoobank.org/16737C47-F8DA-40C1-8D05-B670F1E6FDD0

Type species. Syrichthus [sic] festiva Erichson, [1849].

Definition. A sister genus of *Sostrata* Godman and Salvin, 1895 (Type species. *Leucochitonea scintillans* Mabille, 1876, which is treated as a junior subjective synonym of *Sostrata bifasciata* (Ménétriés, 1829)), where these species were placed before (Fig. 6). Keys to E.42.1a in Evans (1953). Distinguished from *Sostrata* by blue ventral hindwing, broader wings and in male genitalia by longer harpe, clearly separated from the rest of valva. In DNA, a combination of the following base pairs is diagnostic: aly276665.9.3:C95G, aly923.23.1:G274A, aly2790.3.6:A367C, aly923.23.1:T688G, aly2101.22.7:A31C.

Etymology. The name is a feminine noun in the nominative singular, given for the type species and for the festive looks that distinguish this genus from *Sostrata*.

Species included. The type species, *Leucochitonea cronion* C and R. Felder, 1867, *Pythonides caerulans* Mabille and Boullet, 1917, *Pythonides adamantinus* Mabille, 1898, *Sostrata grippa* Evans, 1953, and *Sostrata jinna* Evans, 1953.

Parent taxon. Subtribe Erynnina Brues and Carpenter, 1932.

Comments. Although this genus does not disrupt monophyly of the former concept of *Sostrata* that included these species, genetic distance between these sister genera is not less than that between *Potamanaxas* and *Anaxas*, gen. n., or between *Mylon* and *Anastrus*. This comparatively large genetic differentiation is present in both nuclear and mitochondrial genomes. The two genera are also clearly diagnosed by wing patterns and shapes.

Echelatus Godman and Salvin, [1894] is a valid genus

Echelatus Godman and Salvin, [1894] (type species Anastrus varius Mabille, 1883, which is a junior subjective synonym of Achlyodes sempiternus Butler and Druce, 1872) is reinstated as a valid genus from its synonymy with Anastrus Hübner, [1824], because the group of species that includes Anastrus obscurus Hübner, [1824] (the type species of Anastrus) is sister to Mylon Godman and Salvin, [1894] (type species Leucochitonea lassia Hewitson, 1868) and thus is not monophyletic with the type species of Echelatus (Fig. 6), as suggested by Austin (1998).

Hoodus Grishin, new genus

http://zoobank.org/CC847380-9804-439B-9370-DD2CE72BB0FE

Type species. Hesperia pelopidas Fabricius, 1793.

Definition. A sister genus to *Echelatus* Godman and Salvin, [1894] (type species *Anastrus varius* Mabille, 1883, which is a junior subjective synonym of *Anastrus sempiternus* (Butler and Druce, 1872)) (Fig. 6). Species of this new genus were placed in *Mylon* (type species *Leucochitonea lassia* Hewitson, 1868) previously, key to E.50.10 in Evans (1953), and were defined as the "pelopidas" group by Austin (2000) who gave a detailed morphological diagnosis on p. 5 and 7, which is not repeated here. Most importantly, the two unique diagnostic characters are: vinculum in male genitalia is expanded dorsad on both sides to cover most of tegumen and uncus arms are very short. In DNA, a combination of the following base pairs is diagnostic: aly1735.8.1:T625C, aly3616.13.2:A50G, aly1036.5.1:A436C, aly1735.8.1:C695T, aly1735.8.1:A772C.

Etymology. The name is a masculine noun in the nominative singular, for the vinculum expanded as a hood over tegumen.

Species included. The pelopidas species group as defined by Austin (2000), which consists of the type species, *Mylon cristata* Austin, 2000, *Leucochitonea jason* Ehrmann, 1907, *Mylon ozema* var. *exstincta* Mabille and Boullet, 1917, *Mylon simplex* Austin, 2000, and *Mylon argonautarum* Austin, 2000.

Parent taxon. Subtribe Erynnina Brues and Carpenter, 1932.

Anaxas Grishin, new genus

http://zoobank.org/FE71A883-25C6-491B-B1BE-3EC4F82C62B1

Type species. Antigonus obliqua Plötz, 1884.



Figure 6. Genomic trees of Erynnini. See Fig. 2 legend for notations.

Definition. A likely sister genus to *Potamanaxas* Lindsey, 1925, and thus not monophyletic with *Anastrus* Hübner, [1824] (type species *Anastrus obscurus* Hübner, [1824]), where these species were formerly placed (Fig. 6). The need for this genus has been suggested previously (Austin 1998). Keys to F.6.3. in Evans (1953). Genitalia illustrated by Grishin in his fig. 30–39 (Grishin 2012). Morphologically, distinguished from *Anastrus* and *Echelatus* by having processes on sacculus of the valva and a second pair of uncus arms, from the ventral side of uncus, similar to some species of its sister *Potamanaxas*, but uncus itself is essentially undivided with arms reduced to small knobs. Primary uncus arms are well-developed in *Potamanaxas*. In facies, characterized by brown wings above with several dark-brown stripes and often with some bluish scaling between the stripes. Below, wings plain brown and some species with slate overscaling on the posterior third of hindwing. In DNA, a combination of the following base pairs is diagnostic: aly3570.7.4:A85T, aly3570.7.4:G86C, aly9673.2.5:T298A, aly1735.8.1:C820C (not T), aly16.28.2:C88C (not A), aly16.28.2:A89A (not T).

Etymology. The name is a masculine noun in the nominative singular, a group of species that were formerly placed in Ana[strus], but are phylogenetically closer to [Potamana]xas.

Species included. The type species, its sister *Anastrus isidro* Grishin, 2012 and *Pellicia petius* Möschler, 1877.

Parent taxon. Subtribe Erynnina Brues and Carpenter, 1932.

Chiothion Grishin, new genus

http://zoobank.org/2A17E48E-AD11-46D1-B987-2213B42EEE24

Type species. Pyrgus georgina Reakirt, 1868.

Definition. A sister genus to *Gorgythion* Godman and Salvin, [1896]. Formerly placed in *Chiomara* Godman and Salvin, 1899 (type species *Achlyodes mithrax* Möschler, 1879), but are not monophyletic with the type species (Fig. 6). Keys to F.13.1a or F.13.4 in Evans (1953). Morphologically, characterized by asymmetry in male and female genitalia and distinguished from all other Erynnini general by a likely synapomorphy: corpus bursae in female genitalia with an additional sack-like compartment stemming from its distal end, separated from the corpus by a narrow neck and folded inside the abdomen to point caudad. In wing patterns, distinguished from *Chiomara* and *Crenda* gen. n. by a discal dark bar in the cell of dorsal forewing, also present in its sister *Gorgythion*. In DNA, a combination of the following base pairs is diagnostic: aly1264.13.9:G91T, aly595.7.8:A121C, aly525.18.1:T1397A, aly536.219.9:C2312G, aly260.4.1:G877C.

Etymology. The name is a masculine noun in the nominative singular, a fusion of *Chio*[mara], where these species were formerly placed, and [Gorgy]*thion*, near where they ended up.

Species included. The type species, *Papilio asychis* Stoll, [1780], *Achlyodes basigutta* Plötz, 1884, and *Chiomara khalili* Riley, 1934.

Parent taxon. Subtribe Erynnina Brues and Carpenter, 1932.

Crenda Grishin, new genus

http://zoobank.org/3E0B892C-FC8F-4FA4-A68F-4181F333333BF

Type species. Chiomara crenda Evans, 1953.

Definition. A genus without close relatives (Fig. 6). Keys to F.13.5 in Evans (1953) who gives a detailed diagnosis. Importantly, genitalia (except saccus and penis) are close to symmetrical, especially the uncus. Both uncus and valvae are highly asymmetrical in *Chiomara* Godman and Salvin, 1899 and other related genera such as *Theagenes* Godman and Salvin, 1896, *Gorgythion* Godman and Salvin, 1896. Symmetrical uncus with a pair of side processes on each side at its base, diagnostic of the genus. In DNA, a combination of the following base pairs is diagnostic: aly318.44.1:C664A, aly114.5.7:A142T, aly114.5.7:G143C, aly1294.17.1:A1681C, aly536.188.2:G103C.

Etymology. The name is a feminine noun in the nominative singular, and is a tautonymous name.

Species included. Only the type species.

Parent taxon. Subtribe Erynnina Brues and Carpenter, 1932.

Santa Grishin, new genus

http://zoobank.org/86DDFA39-B2D5-44DB-843C-CA314846FBF4

Type species. Carrhenes santes Bell, 1940.

Definition. A genus related to *Plumbago* Evans, 1953, *Paches* Godman and Salvin, 1895 and *Carrhenes* Godman and Salvin, 1895 (Fig. 7). Keys to E.43.3 or E.52.4 in Evans (1953), one species previously placed in *Paches* Godman and Salvin, [1895] (type species *Pythonides loxus* Westwood, [1852]) the other one in *Carrhenes* Godman and Salvin, 1895 (type species *Leucochitonea fuscescens* Mabille, 1891). Distinguished from others by brown wings with darker spots and stripes above and a black spot near the hindwing tornus below. Male genitalia with a process (style) from ampulla, uncus divided, arms about as long as wide, harpe longer than wide. In DNA, a combination of the following base

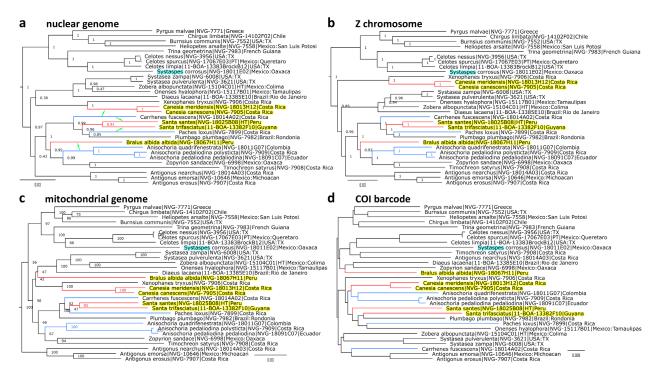


Figure 7. Genomic trees of Pyrgini. See Fig. 2 legend for notations.

pairs is diagnostic: aly5021.3.22:G847C, aly2103.4.2:G472A, aly528.2.2:G1190T, aly528.2.2:G1191T, aly839.26.5:A464T.

Etymology. The name is a feminine noun in the nominative singular, derived from the type species name.

Species included. The type species and Paches trifasciatus Lindsey, 1925.

Parent taxon. Tribe Pyrgini Burmeister, 1878.

Comment. While neither of these two species fit well into the genera they were assigned to previously, due to differences in genitalia and wing patterns between the two species, their sister relationship is not obvious without genomic analysis.

Systaspes Weeks, 1905 is a valid genus

Inspection of Pyrgini trees (Fig. 7) reveals that Antigonus corrosus Mabille, 1878, the type species of Systaspes Weeks, 1905, is a sister to Celotes Godman and Salvin, [1899] (type species Pholisora nessus Edwards, 1877) and is not monophyletic with Antigonus Hübner, [1819] (type species Urbanus erosus Hübner, [1812]). Therefore, we reinstate the genus Systaspes and the combination Systaspes corrosus.

Canesia Grishin, new genus

http://zoobank.org/38584341-521B-4D1E-8115-80FA17A39E30

Type species. Leucochitonea canescens R. Felder, 1869.

Definition. A sister genus to *Xenophanes* Godman and Salvin, 1895 (type species *Papilio tryxus* Stoll, 1780), not so closely related to *Carrhenes* Godman and Salvin, 1895 (type species *Leucochitonea fuscescens* Mabille, 1891), where these species were placed previously (Fig. 7). Keys to E.51.2b in Evans (1953), defined as the canescens group by Austin (2000), but excluding *C. santes*, a species placed here in *Santa*, gen. n. Morphologically, distinguished from *Carrhenes* by the lack of mushroom-shaped process from the

ampulla. The ampulla process in this genus is small, and harpe is small and narrow giving valva the appearance of a crab claw. Forewing without hyaline discal spots near costa. In DNA, a combination of the following base pairs is diagnostic: aly1146.42.8:A2540T, aly27.16.1:A577C, aly275211.5.4:T821C, aly536.106.2:A296T, aly536.106.2:C295A.

Etymology. The name is a feminine noun in the nominative singular, inspired by the name of the type species and a Latin word for crab (cancer), indicating the crab-claw shape of the genitalic valva.

Species included. The "canescens" group of Austin (2000), including the type species, *Achlyodes leada* Butler, 1870, *Carrhenes lilloi* Hayward, 1947, *Carrhenes recurva* Austin, 2000, *Carrhenes callipetes* Godman and Salvin, [1895], *Carrhenes meridensis* Godman and Salvin, [1895].

Parent taxon. Tribe Pyrgini Burmeister, 1878.

Bralus Grishin, new genus

http://zoobank.org/48DF0762-C1E2-4E1D-90C1-EA30A5063308

Type species. Anisochoria albida Mabille, 1888.

Definition. A genus related to *Zopyrion* Godman and Salvin, 1896 (type species *Zopyrion sandace* Godman and Salvin, [1896]), *Timochreon* Godman and Salvin, 1896 (type species *Helias satyrus* C. and R. Felder, [1867]), and *Anisochoria* Mabille, 1876 (type species *Anisochoria polysticta* Mabille, 1876, now considered a subspecies of *Anisochoria pedaliodina* (Butler, 1870)) with its junior subjective synonym *Dicrosema* Bryk, 1953 (type species *Dicrosema quadrifenestrata* Bryk, 1953), but without obvious affinity to either one of these genera (Fig. 7). Keys to E.59.8 in Evans (1953). Morphologically, distinguished from its relatives by almost symmetric valvae with smaller unmodified harpe, ampulla without a process (just with a small knob), and uncus with a pair of short and thin processes at its distal end, between the arms that are widely set apart. Hindwing above with a wide whitish area. In DNA, a combination of the following base pairs is diagnostic: aly8048.1.4:A65G, aly3881.2.1:A178G, aly1097.18.4:G805C, aly1443.3.4:T238T (not G), aly1443.3.4:C239C (not A), aly890.35.8:T164T (not A).

Etymology. The name, a masculine noun in the nominative singular, is a fusion of br[unneis], a[lbum], and lu[teu]s: the three colors of the type species.

Species included. Only the type species.

Parent taxon. Tribe Pyrgini Burmeister, 1878.

Comments. Yet another genus that would be difficult to detect without DNA analysis.

Ladda Grishin, new genus

http://zoobank.org/4BBF25C1-3C31-4B69-9499-516C8251D74E

Type species. Cyclopides eburones Hewitson, 1877.

Definition. A sister genus to *Dalla* Mabille, 1904 (type species *Cyclopides eryonas* Hewitson, 1877), where these species were placed previously (Fig. 8). Consists of the ibhara, part of quadristriga and part of caenides groups of Evans (1955) and keys to H.6.56b, H.6.44b, or H.6.53b. Distinguished by the combination of straight (non-concave) costa of forewing with unmarked, dark brown above hindwing. If hindwing is spotted above, then the spot is single, centered around the cell, ventral hindwing not plain yellow and without a pale ray from base to margin and if a pale spot is present in M_3 -CuA₁ cell, then it is either joined with the spot in CuA_1 -CuA₂ cell, or closer the discal cell spot than to margin and ventral hindwing, and the pale spot in CuA_1 -CuA₂ cell on ventral hindwing is not connected with distal margin by a broadening pale area (but could be connected with the area of the same width as the spot or narrower). In DNA, a combination of the following base pairs is diagnostic: aly103.37.1:A134T, aly638.15.11:T5060C, aly23605.1.35:A566G, aly1139.25.1:G552T, aly23605.1.35:C580G.

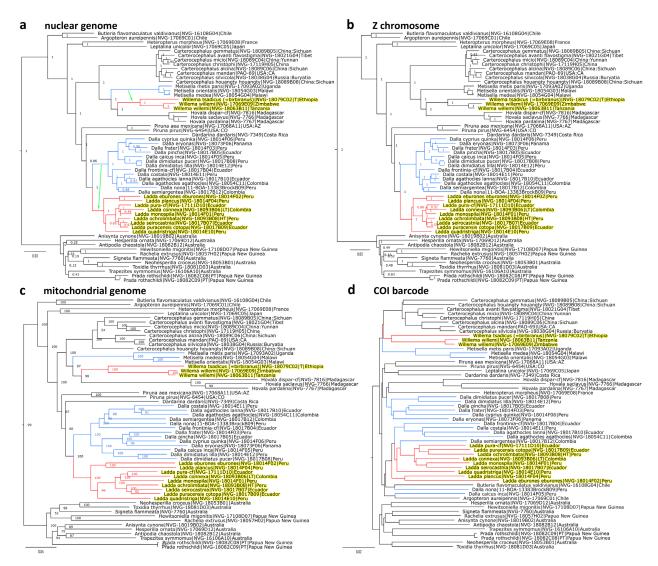


Figure 8. Genomic trees of Heteropterinae and Trapezitinae. See Fig. 2 legend for notations.

Etymology. The name is a feminine noun in the nominative singular, a play of letters, from *Dalla*.

Species included. The type species, Dalla tona Evans, 1955, Dalla decca Evans, 1955, Butleria morva Mabille, 1898, Thanaos ibhara Butler, 1870, Carterocephalus plancus Hopffer, 1874, Cyclopides crithote Hewitson, 1874, Dalla calima Steinhauser, 1991, Dalla rosea Evans, 1955, Butleria granites Mabille, 1898, Dalla parma Evans, 1955, Dalla mars Evans, 1955, Pamphila cuadrada Weeks, 1901, Butleria quadristriga Mabille, 1889, Dalla rubia Evans, 1955, Argopteron xicca Dyar, 1913, Dalla pedro Steinhauser, 2002, Dalla connexa Draudt, 1923, Dalla disconnexa Steinhauser, 2002, Dalla vista Steinhauser, 2002, Butleria ticidas Mabille, 1898, Cyclopides caenides Hewitson, 1868, Dalla bos Steinhauser, 1991, Dalla pura Steinhauser, 1991, Dalla mora Evans, 1955, Dalla carnis Evans, 1955, Butleria monospila Mabille, 1898, Dalla simplicis Steinhauser, 1991, Dalla seirocastnia Draudt, 1923, Dalla celsus Steinhauser, 2002, Dalla pantha Evans, 1955, Dalla puracensis Steinhauser, 1991, Dalla ochrolimbata Draudt, 1923.

Parent taxon. Tribe Heteropterini Aurivillius, 1925.

Comments. Although this genus does not disrupt the monophyly of Dalla when combined with it, Ladda is split from Dalla, due to the large genetic distance between the two genera, each forming a tight cluster of species well separated from the other. Genetic differentiation between these two genera is similar to

that between traditional genera such as *Heteropterus* Duméril, 1806 and *Leptalina* Mabille, 1904, or *Carterocephalus* Lederer, 1852 and *Metisella* Hemming, 1934.

Willema Grishin, new genus

http://zoobank.org/D92BF2E1-F842-44C6-A924-023F495F9589

Type species. Heteropterus willemi Wallengren, 1857.

Definition. A sister genus to *Hovala* Evans, 1937 (type species *Cyclopides pardalina* Butler, 1879), more distant from *Metisella* Hemming, 1934 (type species *Papilio metis* Linnaeus, 1764), where it was placed previously (Fig. 8). Keys to IV.20.B in Evans (1937). Shares many features with *Metisella*, such as hindwing discal cell longer than half of the wing, forewing veins R₁ and R₂ very near each other or touching (not separated). Distinguished from its relatives by the pattern of hindwing underside, which is with dark web-like pattern along veins with some streaks across and orange, ochreous, yellow or white spots. Hindwing below pale-yellow with dark web-like pattern along veins with some streaks across, but without darker markings. Genitalia with narrow and long penis, uncus with tegumen bulbous in dorsal view, uncus narrows to a point (except in *W. willemi* where distal end is mushroom-shaped and base of uncus with side processes), gnathos better developed than in Metisella, ampulla separated from harpe, protruding beyond its caudal end (except in *W. willemi* where harpe upturned embracing ampulla). In DNA, a combination of the following base pairs is diagnostic: aly363.37.1:A929C, aly1450.10.1:A576G, aly283.4.1:T1129C, aly3555.2.2:T116C, aly451.23.2:C1177A.

Etymology. The name is a feminine noun in the nominative singular, derived from the type species name.

Species included. The type species, Cyclopides formosus var. tsadicus Aurivillius, 1905, Cyclopides angolana Karsch, 1896, Metisella kumbona Evans, 1937, Cyclopides carsoni Butler, 1898, Heteropterus formosus Butler, 1894, and Cyclopides perexcellens Butler, 1896.

Parent taxon. Tribe Heteropterini Aurivillius, 1925.

Prada Evans, 1949 belongs to Trapezitinae Waterhouse and Lyell, 1914

Prada (type species *Plastingia rothschildi* Evans, 1928) is transferred to Trapezitinae because it is monophyletic with this subfamily, receiving the strongest statistical support in all trees (Fig. 8), and is not monophyletic with Hesperiinae where it was formerly placed (Warren et al. 2009).

Argemma Grishin, new genus

http://zoobank.org/3A82BAF3-A287-4EBD-8969-327471780BC1

Type species. Apaustus argyrosticta Plötz, 1879.

Definition. A sister genus of *Ceratrichia* Butler, [1870] (type species *Papilio nothus* Fabricius, 1787), where these species were formerly placed, but separated from it by a prominent genetic distance consistent with how other genera are defined (Fig. 9). Keys to VI.31.B in Evans (1937) and differs from its relatives by pale apical spot on ventral hindwing in cell R_5 - M_1 is in line with the spots in cells next to it (not offset strongly basad); uncus bulbous in ventral view, which gives tegumen with uncus a bowling duckpin shape; harpe widening distad, separated from ampulla; ventral hindwing ochreous with many small silvery spots, dorsal forewing brown with large orange-yellow spots. In DNA, a combination of the following base pairs is diagnostic: aly669.9.1:A231G, aly214.13.5:A98G, aly4389.5.1:G869A, aly1329.4.13:T479C, aly214.13.5:A97C.

Etymology. The name is a feminine noun in the nominative singular, a fusion of the Latin words Arg[entum] (silver) and [g]emma (jewel), for the shiny, silvery gem-like spots on the hindwing below.

Species included. The type species, *Ceratrichia aurea* Druce, 1910, *Ceratrichia maesseni* Miller, 1971, *Ceratrichia mabirensis* Riley, 1925, and *Ceratrichia bonga* Evans, 1946.

Parent taxon. Tribe Ceratrichiini, trib. n.

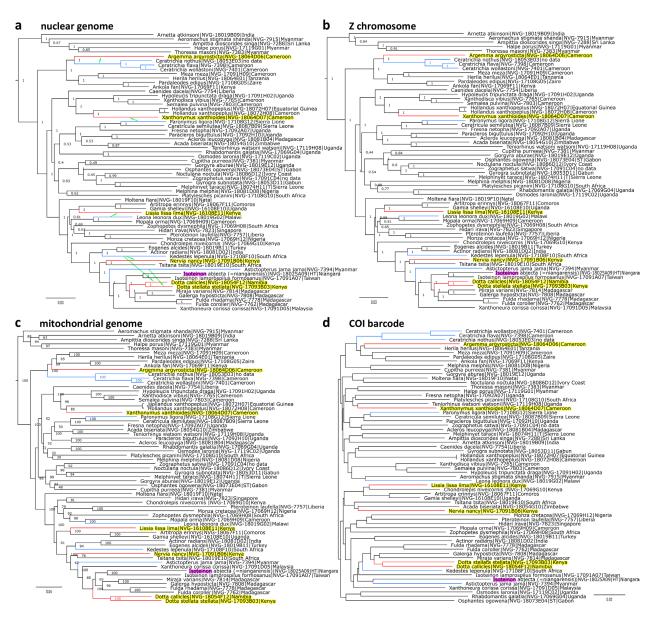


Figure 9. Genomic trees of Astictopterini and relatives. See Fig. 2 legend for notations.

Nervia Grishin, new genus

http://zoobank.org/22F7A77E-E3FF-448C-853D-165AB8D0582A

Type species. Hesperia nerva Fabricius, 1793.

Definition. A sister genus to *Tsitana* Evans, 1937 (type species *Cyclopides tsita* Trimen, 1870) and not monophyletic with *Kedestes* Watson, 1893 (type species *Hesperia lepenula* Wallengren, 1857), where it was previously placed (Fig. 9). Kedestes nerva group (Hancock and Gardiner 1982), keys to V.27.A. in Evans (1937). Distinguished from *Kedestes* by mostly dark below antennae (not pale or checkered), orange submarginal markings on forewing above (at least in females), rusty ventral hindwing margin, narrow uncus, upturned almost rectangular harpe with serrated dorsal edge, not separated from ampulla. In DNA, a combination of the following base pairs is diagnostic: aly6841.65.2:C1166G, aly2578.13.2:C1091A, aly127.64.1:A1342T, aly83.5.1:A418C, aly904.15.14:C3808G.

Etymology. The name is a feminine noun in the nominative singular, formed from the type species name.

Species included. The type species, *Hesperia mohozutza* Wallengren, 1857, *Kedestes protensa* Butler, 1901, *Pyrgus chaca* Trimen, 1873, *Kedestes heathi* Hancock and Gardiner, 1982, *Kedestes ekouyi* Vande weghe and Albert, 2009, *Kedestes michaeli* Gardiner and Hancock, 1982, *Kedestes monostichus* Hancock and Gardiner, 1982, *Kedestes nancy* Collins and Larsen, 1991, and *Kedestes pinheyi* Hancock and Gardiner, 1982.

Parent taxon. Tribe Astictopterini Swinhoe, 1912.

Isoteinon abjecta (Snellen, 1872), new combination

Phylogenetic analysis shows (Fig. 9) that *Pamphila abjecta* Snellen, 1872 is not monophyletic with *Astictopterus jama* C. and R. Felder, 1860, the type species of the genus *Astictopterus* C. and R. Felder, 1860, where *Isoteinon abjecta* was previously placed, and instead forms a clade with *Isoteinon lamprospilus* C. and R. Felder, 1862, the type and the only species of *Isoteinon* C. and R. Felder, 1862. Therefore, we reinstate the combination *Isoteinon abjecta*.

Dotta Grishin, new genus

http://zoobank.org/1579BF7B-B1B2-4B01-B428-6593A087A665

Type species. Ceratrichia stellata Mabille, 1891.

Definition. A genus without close relatives, but not monophyletic with *Astictopterus* C. and R. Felder, 1860 (type species *Astictopterus jama* C. and R. Felder, 1860), *Kedestes* Watson, 1893 (type species *Hesperia lepenula* Wallengren, 1857), or *Isoteinon* C. and R. Felder, 1862 (type species *Isoteinon lamprospilus* C. and R. Felder, 1862) (Fig. 9). Keys to V.27.12 or IV.24.4 in Evans (1937). Distinguished from its relatives by long gnathos well-separated from and about the same length as uncus, terminally narrowing uncus, harpe terminally bulbous, finely serrated, separated from ampulla by a gap, black edged pale dots or spots on ventral hindwing, pale spots or dots above. In DNA, a combination of the following base pairs is diagnostic: aly274.20.6:A1605C, aly84.28.1:G1033C, aly274.20.6:T1601A, aly144.50.1:G193A, aly3194.1.2:C65A.

Etymology. The name is a feminine noun in the nominative singular, for the white dots on the wings.

Species included. The type species and *Cyclopides callicles* Hewitson, 1868 (formerly placed in *Kedestes*).

Parent taxon. Tribe Astictopterini Swinhoe, 1912.

Comments. Without DNA analysis, it would be challenging to pull these two species out of the genera they have been assigned to and join them in a single genus. Nevertheless, retrospectively observed genitalic similarity agrees with such placement.

Lissia Grishin, new genus

http://zoobank.org/C9B3D4FD-98CF-4117-B30A-8A3D80CFD7EA

Type species. Leona lissa Evans, 1937.

Definition. A genus more related to *Gamia* Holland, 1896 (type species *Proteides galua* Holland, 1891) and *Artitropa* Holland, 1896 (type species *Pamphila erynnis* Trimen, 1862) than to *Leona* Evans, 1937 (type species *Hesperia leonora* Plötz, 1879) (Fig. 9). Keys to VIII.59.C(b) in Evans (1937) and differs from its relatives, including *Leona*, by a well-defined gnathos reaching about half of uncus length (in lateral view); tegumen plus uncus twice as long as wide, hour-glass shaped in ventral view; uncus divided, arms short and stout, knob-like; valva with harpe close to rectangular; hindwing with a postdiscal band of spots above. In DNA, a combination of the following base pairs is diagnostic: aly127.66.15:T199A, aly1149.1.1:A1727G, aly822.30.12:A417C, aly490.3.1:A60A (not G), aly6339.4.1:G167G (not T), aly1121.3.2:C344C (not T).

Etymology. The name is a feminine noun in the nominative singular, formed from the type species name.

Species included. The type species and *Plastingia luehderi* Plötz, 1879.

Parent taxon. Tribe Astictopterini Swinhoe, 1912.

Xanthonymus Grishin, new genus

http://zoobank.org/964F4410-339B-486C-8993-30E005481233

Type species. Pardaleodes xanthioides Holland, 1892.

Definition. A sister genus to *Hollandus* Larsen and Collins, 2015 (type species *Pardaleodes xanthopeplus* Holland, 1892), formerly placed in *Paronymus* Aurivillius, 1925 (type species *Hesperia ligora* Hewitson, 1876), but is not monophyletic with it (Fig. 9). Keys to VII.45.4 in Evans (1937) and differs from its relatives by narrow, beak-like uncus; tegumen narrowing into uncus gradually, their junctions slightly concave in ventral view; lack of gnathos; harpe gradually upturned, finely serrated at its short but nearly dorsal margin, with a tooth directed dorsad near its base; hindwing outer margin and tornus brown below. In DNA, a combination of the following base pairs is diagnostic: aly822.30.12:C365T, aly1089.8.4:G53A, aly164.3.1:A811G, aly5294.20.2:A547A (not T), aly5294.20.2:A548A (not C), aly838.12.1:T458T (not A).

Etymology. The name is a masculine noun in the nominative singular, fusion of the type species name with the genus name where it was placed previously: *Xanth*[ioides] and [Par]*onymus*.

Species included. The type species and *Xanthodisca astrape*.

Parent taxon. Tribe Astictopterini Swinhoe, 1912.

Cerba Grishin, new genus

http://zoobank.org/199EACE1-E34F-4266-8CAA-46783F667A0A

Type species. Zea martini Distant and Pryer, 1887.

Definition. Previously placed in *Acerbas* de Nicéville, 1895 (type species *Hesperia anthea* Hewitson, 1868), this genus is not monophyletic with it and is sister to *Zela* de Nicéville, 1895 (type species *Zela zeus* de Nicéville, 1895) instead (Fig. 10). Keys to J.22.4 in Evans (1949). Diagnosed by the following combination of characters: mostly brown above wings with small forewing spots, milky areas by hindwing anal margin and broad central band on hindwing below; forewing vein CuA_2 originates in the middle between wing base and the origin of vein CuA_1 ; uncus broad, bilobed caudad, indistinctly separated from tegumen; tegumen with small side processes and a long central process; penis bulky and stout; valva twice as long as broad, with a prominent and rounded ampulla that is separated from a narrow but large harpe gradually upturned and rounded at the dorsal end protruding above ampulla. In DNA, a combination of the following base pairs is diagnostic: aly103.51.12:A1212T, aly2618.5.1:T1489A, aly420.23.4:A277C, aly276561.5.1:G1724G (not C), aly276561.5.1:A1721A (not G), aly1624.1.10:G534G (not A).

Etymology. The name is a feminine noun in the nominative singular, formed by removing the first and last letters from the name of the genus that the type species was previously attributed to.

Species included. Only the type species.

Parent taxon. Tribe Erionotini Distant, 1886.

Avestia Grishin, new genus

http://zoobank.org/F54B9854-755F-4050-8D47-88EDB994DCED

Type species. Hesperia avesta Hewitson, 1868.

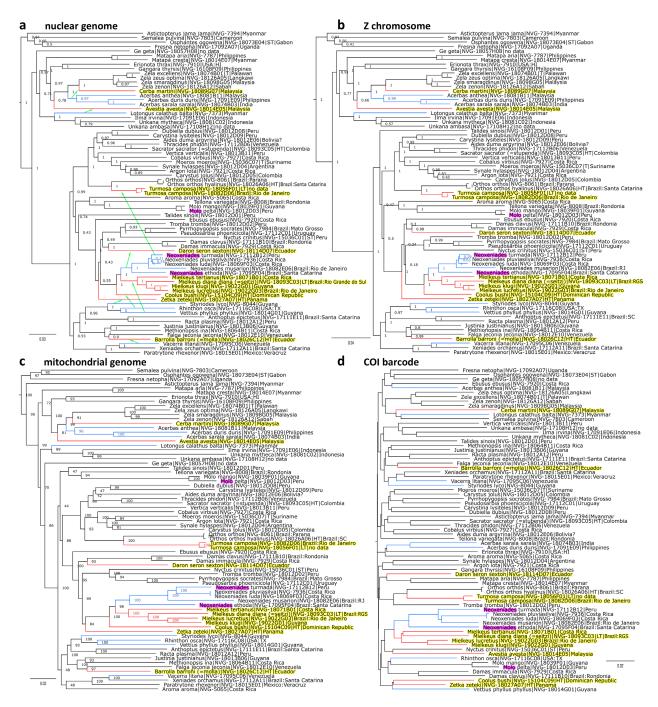


Figure 10. Genomic trees of Erionotini, Carystina and relatives. See Fig. 2 legend for notations.

Definition. A genus sister to *Acerbas* de Nicéville, 1895 (type species *Hesperia anthea* Hewitson, 1868) with *Zela* de Nicéville, 1895 (type species *Zela zeus* de Nicéville, 1895), previously placed in *Lotongus* Distant, 1886 (type species *Eudamus calathus* Hewitson, 1876), but not monophyletic with it (Fig. 10). Keys to J.11.4 in Evans (1949). Morphologically, distinguished from its relatives by hourglass-shaped, terminally bulbous uncus in ventral view; harpe terminally rounded, not prominently separated from ampulla, with a single tooth directed caudad; ventral hindwing yellow band from costa to anal margin, closer to the base, veins brown below (not yellow). In DNA, a combination of the following base pairs is diagnostic: aly1838.39.2:A631G, aly2165.18.2:A22T, aly2096.50.1:A679T, aly1405.13.10:C136C (not G), aly537.26.8:A1279A (not G), aly1264.14.2:A37A (not T).

Etymology. The name is a feminine noun in the nominative singular, formed from the type species name.

Species included. Only the type species.

Parent taxon. Tribe Erionotini Distant, 1886.

Zetka Grishin, new genus

http://zoobank.org/C331EB66-E1BB-4C06-9A8B-22A293353C6D

Type species. Mnasitheus zeteki Bell, 1931.

Definition. Formerly placed in *Styriodes* Schaus, 1913 (type species *Styriodes lyco* Schaus, 1913, in Moncina, Fig. 10–12) this genus does not have close relatives, but belongs in the same clade as *Neoxeniades* Hayward, 1938 (type species *Neoxeniades musarion* Hayward, 1938). Keys to K.2.6. in Evans (1955). Can be told apart from its relatives by the following combination of characters: males with long brands over vein 1A+2A and under vein CuA₂ but not above vein CuA₂, antennae as long as forewing discal cell, palpi broad, 3rd segment short, stout, wings and body brown. Genitalia illustrated by Bell (1931: fig. 1), uncus shorter than tegumen, stout, with small rounded arms, gnathos as long as uncus, saccus vestigial, valva twice as long as wide, harpe angled and squared caudad, upturned dorsocephalad, serrated at the dorsal end that protrudes from valva (Bell 1931). In DNA, a combination of the following base pairs is diagnostic: aly1146.42.8:A1629G, aly1222.14.14:T7181A, aly1405.10.1:C373T, aly251.9.1:C1499T, aly103.32.1:A1000G.

Etymology. The name is a feminine noun in the nominative singular formed from the type species name.

Species included. Only the type species.

Parent taxon. Subtribe Carystina Mabille 1878.

Neoxeniades ethoda (Hewitson, [1866]), new combination

Hesperia ethoda Hewitson, [1866] is transferred to Neoxeniades, where it is positioned in the genomic trees (Fig. 10), not being monophyletic with Xeniades Godman, [1900] (type species Papilio orchamus Cramer, [1777]), where it was placed previously, to form the new combination Neoxeniades ethoda (Hewitson, [1866]). The differences in unci between the two genera agree with this transfer. Uncus arms are well-developed and widely apart in Xeniades, but are vestigial (uncus almost undivided) in Neoxeniades.

Turmada Evans, 1955 is a synonym of Neoxeniades Hayward, 1938

Turmada originates within Neoxeniades (Fig. 10), and genetic distance between the type species of these genera is small (COI barcode difference 5.5%, and differences as much as 10% are frequent between congeners). Therefore, Turmada is designated a new junior subjective synonym of Neoxeniades.

Turmosa Grishin, new genus

http://zoobank.org/E6AD0703-A3E7-4397-84C6-F4D16308504E

Type species. Hesperia camposa Plötz, 1886.

Definition. A sister genus to *Orthos* Evans, 1955 (type species *Eutychide orthos* Godman, 1900) and in a different clade from *Turmada* Evans, 1955 (type species *Dion turmada* Druce, 1912), where it was placed previously (Fig. 10). Keys to K.16.2 in Evans (1955). Differs from other Hesperiidae by unspotted metallic blue-green wings above, hindwing below shiny orange-green with dark veins and narrow discal black band. In DNA, a combination of the following base pairs is diagnostic: aly276634.6.3:C296A, aly320.9.2:A59G, aly276634.6.3:C353T, aly5965.2.3:G988A, aly2582.35.2:A932T.

Etymology. The name is a feminine noun in the nominative singular, a fusion of two words in a former name: Turm[ada] and [camp]osa.

Species included. Only the type species.

Parent taxon. Subtribe Carystina Mabille 1878.

Mielkeus Grishin, new genus

http://zoobank.org/DED1D74F-CC56-48CC-8F38-E4C6AE84D70B

Type species. Cobalus tertianus Herrich-Schäffer, 1869.

Definition. A genus in the same clade with *Neoxeniades* Hayward, 1938 (type species *Neoxeniades musarion* Hayward, 1938) and *Coolus* gen. n., and in a different subtribe (Carystina) from *Vettius* Godman, [1901] (type species *Papilio phyllus* Cramer, [1777]), which is in Moncina (Fig. 10). Keys to J.45.21 and J.45.15b. Distinguished from other Hesperiidae by the following combination of characters: uncus almost undivided, arms medium to short; gnathos small, narrow, about the same as uncus in lateral view; valva plate-like without processes, harpe about half of the valva, with a small notch on dorsal margin; no brands on forewing; mid tibiae smooth; milky-white areas on hindwing below, especially near the base, gradually turning brown towards outer margin; veins not darker. In DNA, a combination of the following base pairs is diagnostic: aly294.13.2:A968C, aly1249.14.7:A1222T, aly822.47.3:A218G, aly1838.8.2:A172C, aly127.74.5:T874C.

Etymology. The name is a masculine noun in the nominative singular, named both for the milky pattern on the hindwing below and to honor skipper taxonomist Olaf H. H. Mielke.

Species included. The type species, Carystus klugi Bell, 1941, Hesperia lucretius Latreille, [1824], and Hesperia diana Plötz, 1886.

Parent taxon. Subtribe Carystina Mabille 1878.

Coolus Grishin, new genus

http://zoobank.org/64340957-82B3-48A9-BF06-16E2726987FE

Type species. Rhinthon bushi Watson, 1937.

Definition. A genus in the same clade with *Neoxeniades* Hayward, 1938 (type species *Neoxeniades musarion* Hayward, 1938) and far removed from *Rhinthon* Godman, [1900] (type species *Proteides chiriquensis* Mabille, 1889 treated as a junior subjective synonym of *Hesperia osca* Plötz, 1882) where it was previously placed, but does not belong (Fig. 10) in agreement with Burns et al. (Burns et al. 2010) and Smith et al. (Smith et al. 1994), who also questioned its position in *Rhinthon* but did not propose a proper genus for it. Keys to L.8.2 in Evans (1955). Distinguished from all other Hesperiidae by unique pattern in the basal half of brown ventral hindwing consisting of five spots: three connected, silvery-white, the one in the distal half of discal cell framed yellow, and two others towards tornus, and two small yellow ones, near the base of the cell and past it; and orange fringes in tornal area of both wings, dark brown elsewhere. In DNA, a combination of the following base pairs is diagnostic: aly2487.23.1:C242G, aly13410.9.1:G2107T, aly2700.17.1:G706A, aly1407.7.6:A2449C, aly1283.9.1:A694G.

Etymology. The name is a masculine noun in the nominative singular meant to suggest the incredibly unique looks of this awesome skipper with icy patches on the hindwing below.

Species included. Only the type species.

Parent taxon. Subtribe Carystina Mabille 1878.

Daron Grishin, new genus

http://zoobank.org/D4A2E257-03F9-4DA7-B668-CDB6BBE9C420

Type species. Thracides seron Godman, [1901].

Definition. Removed from *Neoxeniades* Hayward, 1938 (type species *Neoxeniades musarion* Hayward, 1938) where it was formerly placed and a sister to *Damas* Godman, 1901 (type species *Goniloba clavus* Herrich-Schäffer, 1869), this genus differs from it by about 12% in the COI barcode region (Fig. 10). Keys to 0.16.5 in Evans (1955). Distinguished from its relatives by the following combination of characters: antennae longer than 2/3 of costa, palpi flattened and pointed upwards, 3rd segment short, stout, mid-tibiae without spines, forewings elongated, discal cell without pale spots, as long as inner margin, uncus two times shorter than tegumen, with small knob-like rounded arms, penis undulate, valva simple, twice as long as wide, ampulla leveled with the rest of the valva, harpe about 1/3 of the valva length, slightly upturned, broad, with serrate dorsal margin. In DNA, a combination of the following base pairs is diagnostic: aly767.14.7:A754C, aly1775.4.5:A83T, aly140.7.3:T104A, aly851.3.4:C1900C (not T), aly536.9.2:A221A (not T), aly531.4.3:G193G (not T).

Etymology. The name is a masculine noun in the nominative singular, a fusion of the type species names Da[mas+se]ron.

Species included. Only the type species.

Parent taxon. Subtribe Carystina Mabille 1878.

Vacerra molla E. Bell, 1959 is a synonym of Paratrytone barroni Evans, 1955

This species is known only from one female of $Paratrytone\ barroni\ Evans$, 1955 and one male of $Vacerra\ molla\ E$. Bell, 1959, both from Ecuador, which have similar distinct wing patterns. In particular, like in no other Hesperiidae, the hindwing below has a single cream band from mid-costa to the outer margin at cell CuA_2 -1A+2A, continuing as white fringes, and partly seen on the dorsal surface as a straight streak of spots. Therefore, $V.\ molla$ is designated a junior subjective synonym of $Paratrytone\ barroni\ Evans$, 1955.

Barrolla Grishin, new genus

http://zoobank.org/3BFA1AE8-9838-4E31-AC12-D139E3636083

Type species. Paratrytone barroni Evans, 1955.

Definition. A possible sister to Falga Mabille, 1898 (type species Carystus jeconia Butler, 1870) and in a different subtribe from Vacerra Godman, [1900] (type species Hesperia litana Hewitson, [1866]) (Fig. 10). Keys to M.23.9. in Evans (1955). Diagnosed by large and broad square palpi that are longer than the head, 2nd segment longer than usual and directed forward rather than dorsad, and 3rd segment very small; hindwing with a middle straight cream band on both sides. Genitalia illustrated by Bell (1959, fig. 18): uncus shorter than tegumen, arms rounded, gnathos longer than tegumen, saccus is very short, almost vestigial, valva broadens caudad, harpe short, upturned, joint with ampulla, without teeth or processes (Bell 1959). In DNA, a combination of the following base pairs is diagnostic: aly1603.20.1:G64A, aly84.28.1:C2633G, aly707.2.13:T749A, aly798.5.3:G284G (not A), aly291.21.2:T2561T (not G), aly102.6.2:G1936G (not A).

Etymology. The name is a feminine noun in the nominative singular, formed as a fusion of the species names: senior and junior synonyms referring to the Type species. Bar[roni + m]olla.

Species included. Only the type species.

Parent taxon. Subtribe Falgina trib. n., described above.

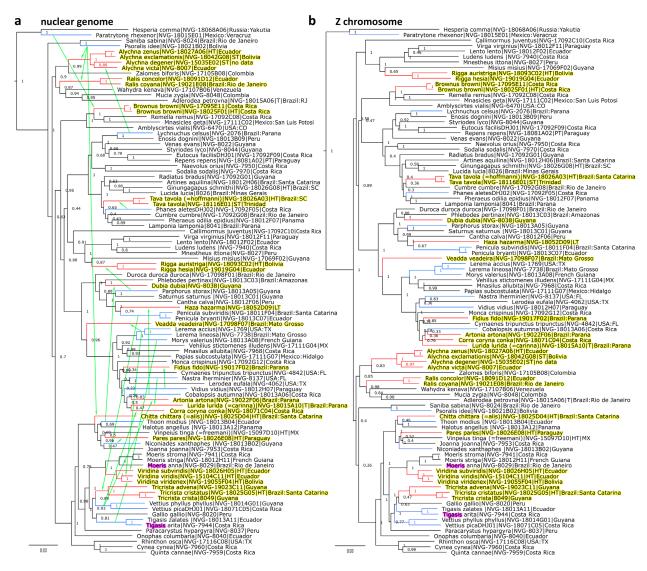


Figure 11. Nuclear genome and Z chromosome trees of Moncina and relatives. See Fig. 2 legend for notations.

Comments. A genus with surprising phylogenetic placement that yet again indicates the value of DNA-based methods at the genomic scale, and instills confidence in the results due to the large datasets used for phylogeny reconstruction and highly confident nodes in phylogenetic trees.

Brownus Grishin, new genus

http://zoobank.org/99C1730A-E965-410D-A15A-D8754C5E3FF0

Type species. Paratrytone browni Bell, 1959.

Definition. Unexpectedly, a relative of *Amblyscirtes* Scudder, 1872 (type species *Hesperia vialis* Edwards, 1862) and is sister to *Remella* Hemming, 1939 (type species *Hesperia remus* Fabricius, 1798) with *Mnasicles* Godman, [1901] (type species *Mnasicles geta* Godman, 1901), which all are in a different subtribe (Moncina) from *Paratrytone* Godman, [1900] (type species *Paratrytone rhexenor* Godman, 1900) (in Hesperiina) where this lineage was placed previously (Fig. 11–12). Distinguished from its relatives by sacculus about as long as vinculum with tegumen in lateral view, valva simple, nearly diamond-shaped

with vary narrow harpe ending in a point directed dorsoposteriad and not separated from ampulla, very long antennae reaching apical spots near costa, all (except the small cell spot) forewing spots forming a continuous line (separated by black veins only) and most spots in a line on ventral hindwing. In DNA, a combination of the following base pairs is diagnostic: aly28779.8.7:T461C, aly577.34.1:C115G, aly1370.21.5:A2861G, aly531.10.5:G517C, aly2124.3.99:A3193G.

Etymology. The name is a masculine noun in the nominative singular, for the type species.

Species included. Only the type species.

Parent taxon. Subtribe Moncina A. Warren, 2008.

Tava Grishin, new genus

http://zoobank.org/A4C0707B-1BB7-410C-9682-7ACC79306988

Type species. Phanis tavola Schaus, 1902.

Definition. A genus without apparent close relatives, although in the same clade with *Phanes* Godman, [1901] (type species Thracides aletes Geyer, 1832), where it was placed previously but not monophyletic with it, Cumbre Evans, 1955 (type species Phanis cumbre Schaus, 1902), Lamponia Evans, 1955 (type species Hesperia lamponia Hewitson, 1876), and Pheraeus Godman, [1900] (type species Carystus epidius Mabille, 1891) among others (Fig. 11–12). Keys to J.23.5 in Evans (1955). Similar to many species in this group of relatives in having characteristic nearly square in many specimens, semi-hyaline spots near the bases of cells M₂-CuA₁ and CuA₁-CuA₂ on forewing. Differs from them by the combination of the following characters: antennae about 2/3 of costa length, nudum of 11 segments, palpi longer than wide in ventral view, 3rd segment short and conical, mid-tibiae with spines, forewing discal cell with well-developed recurrent vein, stigma V-shaped, inconspicuous, at the base of CuA,-CuA, cell along veins, wings without apical spots, rufous brown below, hindwings mostly unspotted, in some specimens with a trace of small postdiscal yellowish spots (Bell 1940). Genitalia as figured by Bell (1940: fig. 6), distinctive in the shape of harpe, rather straight, only slightly upturned and narrowing to a point, ampulla with a small tooth, penis wider than harpe, longer than valva, tegumen and uncus shorter than harpe. In DNA, a combination of the following base pairs is diagnostic: aly671.22.3:T167A, aly374.12.1:G1087C, aly2874.22.6:A1535G, aly671.22.3:A155T, aly420.67.1:A1631T.

Etymology. The name is a feminine noun in the nominative singular, derived from the beginning of the type species name.

Species included. Only the type species.

Parent taxon. Subtribe Moncina A. Warren, 2008.

Comments. We sequenced a syntype of *Phanis tavola* and the holotype of *Phanes hoffmanni* Bell, 1940. Analysis of these genomic sequences confirms the synonymy previously suggested for these two taxa (Mielke 1995). For instance, their COI barcodes differ by only one position (0.15%).

Rigga Grishin, new genus

http://zoobank.org/5D98B6E8-8FF9-478D-B37B-D19DACCA9E31

Type species. Vorates auristriga Draudt, 1923.

Definition. A likely sister to *Misius* Evans, 1955 (type and the only species *Pamphila misius* Mabille, 1891), this genus is removed from *Parphorus* Godman, 1900 (type species *Phlebodes storax* Mabille, 1891), where it was placed previously (Fig. 11–12). Keys to J.34.9 or J.34.10a in Evans (1955). Diagnosed by long and prominent triangular hyaline yellow spot filling out forewing cell R₅-M₁ from the base to about a third of its length (except *paramus* E. Bell, 1947, which has typical for the genus genitalia), and no other apical forewing hyaline spots, hindwing below with more or less prominent pale ray along vein

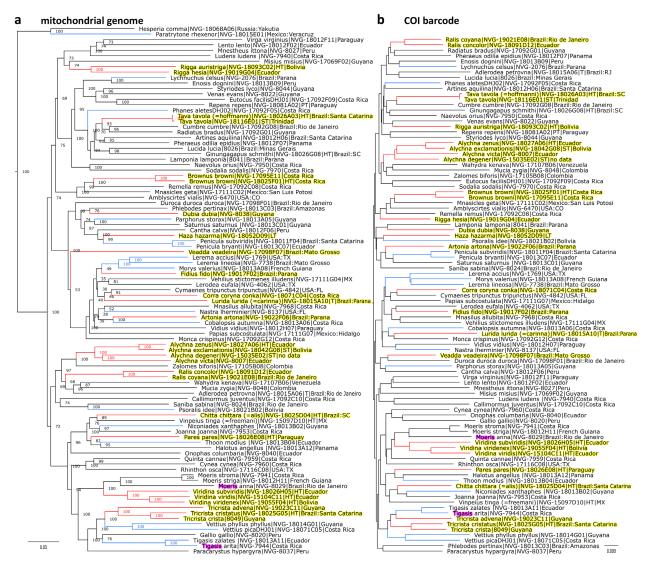


Figure 12. Mitochondrial genome and COI barcode trees of Moncina and relatives. See Fig. 2 legend for notations.

 $\rm M_1$ from wing base to outer margin. Genitalia differ from relatives by a combination of the following characters: uncus twice as long as tegumen, arms parallel and narrow, gnathos vestigial, valva twice as long as wide, harpe not extending much beyond bulky rounded ampulla, upturned and not separated from ampulla. In DNA, a combination of the following base pairs is diagnostic: aly536.116.6:A107G, aly2124.4.17:G268T, aly6377.1.2:A4462G, aly208.50.8:T236A, aly536.108.1:C2126T.

Etymology. The name is a feminine noun in the nominative singular, containing the last four letters of the type species name with an extra 'g' added to avoid a homonym.

Species included. The type species, *Hesperia hesia* Hewitson, 1870, *Apaustus ira* Butler, 1870, *Euroto oeagrus* Godman, [1900], *Vorates paramus* Bell, 1947, and *Vorates sapala* Godman, [1900].

Parent taxon. Subtribe Moncina A. Warren, 2008.

Haza Grishin, new genus

http://zoobank.org/78B60979-8EA8-437D-8B48-24DE9AA92587

Type species. Hesperia hazarma Hewitson, 1877.

Definition. A sister to *Penicula* Evans, 1955 (type species *Pamphila bryanti* Weeks, 1906), this genus is away from *Cobalopsis* Godman, [1900] (type species *Pamphila edda* Mabille, 1891, which is a junior subjective synonym of *Hesperia autumna* Plötz, 1882) (Fig. 11–12). Keys to J.37.9 in Evans (1955), but very variable in hindwing patterns, from virtually brown unspotted to pale cream with brown-yellow veins and several central black spots surrounded by yellow or brown. Diagnosed by male genitalia with uncus not protruding from tegumen, very broad caudad with tiny knob-like arms on the side, uncus together with tegumen almost square in dorsal view, valva with very narrow extended hook-shaped harpe like no other Hesperiini. In DNA, a combination of the following base pairs is diagnostic: aly274.33.1:A430C, aly2202.27.1:C668A, aly2613.3.2:A2043C, aly2613.3.2:C2248A, aly587.20.1:T1522C.

Etymology. The name is a feminine noun in the nominative singular, containing the first four letters of the type species name.

Species included. Only the type species.

Parent taxon. Subtribe Moncina A. Warren, 2008.

Dubia Grishin, new genus

http://zoobank.org/DBA6AFEC-D3DE-4C61-9FA9-7332B4BE5293

Type species. Euroto (?) dubia Bell, 1932.

Definition. A sister genus to *Phlebodes* Hübner, [1819] (type species *Papilio pertinax* Stoll, [1781]), but not monophyletic with and far removed from *Thoon* Godman, [1900] (type species *Proteides modius* Mabille, 1889), where it was previously placed (Fig. 11–12). Keys to J.48.4 in Evans (1955). Genitalia illustrated by Bell in his fig. 3 (Bell 1932). Distinguished from its relatives by elongated uncus with short arms; gnathos widely separated from uncus, about half of its length in lateral view; saccus long, about the same length as valva, valva hourglass shaped in lateral view, harpe rounded, with finely serrated dorsal margin, not separated from ampulla; dorsal forewing with two hyaline spots in discal cell and small brands at the origin and between veins CuA₁ and CuA₂; 2-5 small postdiscal dark spots, some with hyaline centers, placed along smooth curve on brown hindwing below. In DNA, a combination of the following base pairs is diagnostic: aly1281.8.1:A614T, aly390.17.1:T17C, aly1260.9.2:T347G, aly158825.1.3:A37A (not G), aly6286.5.7:A87A (not T), aly5543.13.1:A1601A (not G).

Etymology. The name, a feminine noun in the nominative singular, echoes the type species name.

Species included. Only the type species.

Parent taxon. Subtribe Moncina A. Warren, 2008.

Pares Grishin, new genus

http://zoobank.org/2DD85F52-3615-4008-81A3-52AD86257662

Type species. Phlebodes pares Bell, 1959.

Definition. A genus near *Joanna* Evans, 1955 (type species *Joanna joanna* Evans, 1955) and *Vinpeius* Austin, 1997 (type and the only species *Pompeius tinga* Evans, 1955), and in same clade with *Niconiades* Hübner, [1821] (type species *Niconiades xanthaphes* Hübner, [1821]), where it was placed previously (Fig. 11–12). Indeed resembles some *Niconiades* species due to the presence of large hyaline spots on hindwing and green scaling of the body, but genitalia (illustrated by Bell 1959: fig. 22 and Nicolay 1980: fig. 8) quite distinctive and diagnostic: short uncus and tegumen, together not longer than valva height, uncus rounded and weakly bilobed caudad, saccus long, about half of vinculum length, penis narrowing caudad, valva broad, more than half of its length, ampulla transitions to harpe without a break, harpe projecting

caudad as a broad straight tooth, indented before the tooth along ventral margin. (Bell 1959; Nicolay 1980). In DNA, a combination of the following base pairs is diagnostic: aly890.44.9:C94G, aly9673.13.1:C269G, aly3555.4.2:G923A, aly1041.12.1:T93T (not C), aly537.7.1:T984T (not C), aly1038.8.1:A1367A (not C), aly8857.2.1:G1234G (not A), aly1041.22.3:A377A (not G), aly517.17.2:G336G (not C).

Etymology. The name, a masculine noun in the nominative singular, echoes the type species name.

Species included. The type species and *Thoon maritza* Nicolay, 1980.

Parent taxon. Subtribe Moncina A. Warren, 2008.

Chitta Grishin, new genus

http://zoobank.org/BCD865CA-926C-43DB-9EEB-8E14E7296149

Type species. Phlebodes chittara Schaus, 1902.

Definition. A possible sister to the clade with *Thoon* Godman, [1900] (type species *Proteides modius* Mabille, 1889) and *Halotus* Godman, [1900] (type species *Hesperilla saxula* Mabille, 1891, a subjective junior synonym of *Hesperia angellus* Plötz, 1886) (Fig. 11–12). Not monophyletic with *Psoralis* Mabille, 1904 (type species *Psoralis sabaeus* Mabille, 1904, currently considered a junior subjective synonym of *Pamphila idee* Weeks, 1901) where it was placed previously (Fig. 11–12). Immediately distinguished from *Psoralis* by greenish ventral colors and a pattern of four pale dots on hindwing below, indeed reminiscent of *Thoon*. Genitalia illustrated by Bell (1959: fig. 24, as *Psoralis alis* Bell, 1959, a junior subjective synonym of *Chitta chittara*), quite distinctive and diagnostic: uncus and tegumen very short, look compressed, penis with a long titillator diverging from penis, valva peculiar, crooked with three teeth on harpe, directed dorsad, caudad and ventrad (Bell 1959). In DNA, a combination of the following base pairs is diagnostic: aly1107.9.6:A335C, aly13410.7.2:T37C, aly1838.8.3:G730G (not C), aly127.64.1:G1182G (not A), aly4966.20.2:A1499A (not G).

Etymology. The name is a feminine noun in the nominative singular formed from the type species name.

Species included. Only the type species.

Parent taxon. Subtribe Moncina A. Warren, 2008.

Artonia Grishin, new genus

http://zoobank.org/48FE4172-440B-4E90-90E7-F0CD8BDA61F2

Type species. Hesperia artona Hewitson, 1868.

Definition. A likely sister to *Cobalopsis* Godman, [1900] (type species *Pamphila edda* Mabille, 1891, which is a subjective junior synonym of *Hesperia autumna* Plötz, 1882) and is in a different clade from *Vettius* Godman, [1901]; (type species *Papilio phyllus* Cramer, [1777]) (Fig. 11–12). Keys to J.45.13 in Evans (1955). Superficially indeed similar to some *Vettius* species in the gist of ventral wing patters, but differs from them in lacking brands in males, lacking white streak in cell 1A+2A-3A on dorsal hindwing, but having white spot in cell CuA_2 -1A+2A on dorsal forewing and white veins on ventral hindwing. In male genitalia, uncus short and broad, arms far apart, short, gnathos not expanded in lateral view, penis narrow, curved, harpe close to ampulla, upturned, ends in a finger-like process protruding dorsad from the valva. In DNA, a combination of the following base pairs is diagnostic: aly423.31.1:C427T, aly423.31.1:A428C, aly2548.21.8:T1491C, aly6841.51.2:A710T, aly1405.20.15:G592A.

Etymology. The name is a feminine noun in the nominative singular formed from the name of the type species.

Species included. Only the type species.

Parent taxon. Subtribe Moncina A. Warren, 2008.

Lurida Grishin, new genus

http://zoobank.org/569CDC39-86BD-4E74-ABE2-E52FD84E8EEE

Type species. Cobalus lurida Herrich-Schäffer, 1869.

Definition. A genus near *Cymaenes* Scudder, 1872 (type species *Cobalus tripunctus* Herrich-Schäffer, 1865) and not monophyletic with *Nastra* Evans, 1955 (type species *Hesperia lherminier* Latreille, [1824]), where it was placed previously (Fig. 11–12). Keys to J.26.10 in Evans (1955). Distinguished from its relatives by terminally broad and concave nearly divided uncus; gnathos reaches the end of uncus, weakly separated from it; harpe ventrally excavated, shaped like can opener (with a notch); dark brown cheeks; wings rather rounded, with areas of violet-gray overscaling at outer margins below; a curve of small pale postdiscal spots on ventral hindwing. In DNA, a combination of the following base pairs is diagnostic: aly386.7.5:A205C, aly5582.8.1:A1097T, aly138.11.7:G998C, aly84.96.4:G40A, aly5582.8.1:C1037T.

Etymology. The name, a feminine noun in the nominative singular, echoes the type species name.

Species included. Only the type species.

Parent taxon. Subtribe Moncina A. Warren, 2008.

Corra Grishin, new genus

http://zoobank.org/D013B16C-CC3F-43F5-8B00-6E861AA3B149

Type species. Hesperia coryna Hewitson, [1866].

Definition. A sister genus to *Lurida* gen. n. and is in a different clade from *Vettius* Godman, [1901] (type species *Papilio phyllus* Cramer, [1777]) where it was placed previously (Fig. 11–12). Keys to J.45.20 in Evans (1955). No brands or stigmas in males, forewing above with pale scaling along anal margin, hindwing below with dark costa and pale ray from base to outer margin above the middle of discal cell and a dark ray below it. Uncus and gnathos deeply divided, arms narrow, far apart, gnathos the same length as uncus, saccus long, as vinculum in lateral view, valva broad, not longer than twice the width, harpe narrow, short and upturned, rounded at the tip. In DNA, a combination of the following base pairs is diagnostic: aly686.30.9:A280T, aly345.16.1:A127T, aly527.10.7:A209G, aly451.23.2:A331C, aly529.9.1:C862A.

Etymology. The name is a feminine noun in the nominative singular, formed from the beginning of the type species name.

Species included. Only the type species.

Parent taxon. Subtribe Moncina A. Warren, 2008.

Fidius Grishin, new genus

http://zoobank.org/F4BFE58B-3E0D-4E6F-950D-8A3BD8430313

Type species. Vidius fido Evans, 1955.

Definition. A possible sister to *Cymaenes* Scudder, 1872 (type species *Cobalus tripunctus* Herrich-Schäffer, 1865) and far removed from *Vidius* Evans, 1955 (type species *Narga vidius* Mabille, 1891) where it was placed previously (Fig. 11–12). Keys to J.24.1 in Evans (1955). Genitalia illustrated by Mielke in his fig. 5–8 (Mielke 1980). Distinguished from its relatives by antennae shorter than half of costa length, wings rounded, broad, scales in fringes long, especially on forewing, nearly the same as palpi in length, wings almost entirely unmarked, chocolate brown, in some specimens, wings below with faint, blotchy yellowish spots consisting of a few scales and forming postdiscal bands and a discal cell spot on hindwing. Valva narrow, ~2-3 times longer than wide, harpe distally slightly to moderately

upturned not well-separated from valva, costa of valva straight, uncus narrowly divided, arms short, close together, gnathos the same length as uncus, saccus as long as tegumen with uncus, penis narrow, twice as long as saccus. In DNA, a combination of the following base pairs is diagnostic: aly2096.50.1:A1451C, aly349.40.1:A898C, aly164.63.12:C436A, aly164.63.12:A437T, aly5965.2.3:C1150G.

Etymology. The name is a masculine noun in the nominative singular. It is a fusion of the type species name with its original genus name: Fid[o] + [Vid]ius.

Species included. The type species and *Vidius ochraceus* O. Mielke, 1980.

Parent taxon. Subtribe Moncina A. Warren, 2008.

Veadda Grishin, new genus

http://zoobank.org/49E3A1A5-0F8B-4374-AA86-6EA2C1EEFCB6

Type species. Lerema veadeira Mielke, 1968.

Definition. Not closely related to any other genus, but falls near the clade of many Moncina genera, among which are *Parphorus* Godman, [1900] (type species *Phlebodes storax* Mabille, 1891), *Phlebodes* Hübner, [1819] (type species *Papilio pertinax* Stoll, 1781), *Cantha* Evans, 1955 (type species *Cantha calva* Evans, 1955), *Saturnus* Evans, 1955 (type species *Papilio saturnus* Fabricius, 1787), *Penicula* Evans, 1955 (type species *Pamphila bryanti* Weeks, 1906), and *Duroca* Grishin, 2019 (type species *Hesperia duroca* Plötz, 1882). Not closely related to *Lerema* Scudder, 1872 (type species *Papilio accius* Smith, 1797), where it was originally placed (Fig. 11–12). Distinguished from them by a combination of the following characters: uncus divided, arms long, very close together; gnathos broader than uncus in ventral view, about 2/3 of uncus length; saccus nearly the same length as vinculum in lateral view; harpe short, less than 1/3 of valva, upturned, ending in a tooth directed dorsad, not separated from ampulla and not serrated; dorsal forewing with small ochre spots and tripartite gray stigma, lined with black scales basad; mostly ferruginous (a rather unusual color for Hesperiidae) below, with faint ochreous spots on both wings. In DNA, a combination of the following base pairs is diagnostic: aly318.7.1:A1032G, aly1042.23.2:G113T, aly1042.23.2:A115G, aly1487.4.1:A662G, aly164.9.1:A1676T.

Etymology. The name is a feminine noun in the nominative singular, formed from the first part of the type species name.

Species included. Only the type species.

Parent taxon. Subtribe Moncina A. Warren, 2008.

Tricrista Grishin, new genus

http://zoobank.org/97A09601-6840-4813-A346-31A073A507D4

Type species. Penicula crista Evans, 1955.

Definition. A sister genus to *Vettius* Godman, [1901] (type species *Papilio phyllus* Cramer, 1777) and far removed from *Penicula* Evans, 1955 (type species *Pamphila bryanti* Weeks, 1906) (Fig. 11–12), where these species were placed previously due to the characteristic tuft of hair-like scales in the discal area of hindwing above, apparently a convergent feature different in details between them. Keys to L.10.2a in Evans (1955). Distinguished from its relatives (including *Penicula*) by the structure of its dorsal hindwing area with the tuft of scales: the dense (not thin) tuft from upper part of discal cell at its base, covering gray (not black) area by the thickened bases of veins CuA_2 and CuA_2 (not reaching vein 1A+2A). No hyaline spots in forewing cell $\text{M}_2\text{-M}_3$. In DNA, a combination of the following base pairs is diagnostic: aly2041.22.2:G133A, aly563.7.2:A292G, aly208.38.3:A886T, aly536.2.4:A94A (not T), aly3721.1.24:A167A (not G), aly3721.1.24:A169A (not C).

Etymology. The name is a feminine noun in the nominative singular, and it denotes that three species in the genus have "crist" in their names.

Species included. The type species, *Penicula cristina* Evans, 1955, *Cobalus cristatus* Bell, 1930, *Penicula roppai* Mielke, 1980, and *Rinthon* [sic] *advena* Draudt, 1923.

Parent taxon. Subtribe Moncina A. Warren, 2008.

Viridina Grishin, new genus

http://zoobank.org/0A14CB25-9511-4311-8B76-746AA38011C1

Type species. Lerema (?) viridis Bell, 1942.

Definition. A close sister genus to *Moeris* Godman, [1900] (type species *Talides striga* Geyer, [1832]). Not in the same clade as *Lerema* Scudder, 1872 (type species *Papilio accius* Smith, 1797), *Tigasis* Godman, [1900] (type species *Tigasis zalates* Godman, [1900]) or its subjective synonym *Alerema* Hayward, 1942 (type species *Alerema aeteria* Hayward, 1942 treated as junior subjective synonym of *Phlebodes simplex* Bell, 1930), where species from this genus were placed previously (Fig. 11–12). Keys to J.44.10 or J.41.6. in Evans (1955). Genitalia illustrated by Bell in his fig. 9 (Bell 1942). Distinguished from its relatives by a combination of the following characters: uncus undivided, narrowing to a point; in ventral view: uncus sides slightly concave, gnathos arms widely apart, crescent-shaped, the same length as uncus, forming an φ together with it; saccus short, about the length of uncus, penis stout; harpe shorter than 1/3 of valva, slightly upturned, finely serrated at the distal margin, with a small tooth near ampulla, separated from it by a small notch; antennae longer than half of costa, checkered; wings produced; narrow complete stigma on forewing from base of vein CuA₁ to vein 1A+2A in males; hindwing below greenish-ochreous with a postdiscal irregular row of indistinct pale blotch-like spots. In DNA, a combination of the following base pairs is diagnostic: aly1019.13.1:C3748G, aly208.17.4:G1754C, aly1475.19.1:A667G, aly1041.11.5:G116G (not A), aly1294.15.2:A1078A (not T), aly272.12.6:A338A (not G).

Etymology. The name is a feminine noun in the nominative singular, and denotes that all three species placed in this genus have "virid" in their names.

Species included. The type species, *Pamphila viridenex* Weeks, 1901, which is a very close relative, and *Oeonus subviridis* Hayward, 1940.

Parent taxon. Subtribe Moncina A. Warren, 2008.

Moeris anna (Mabille, 1898), new combination

Pamphila anna Mabille, 1898 was previously placed in Vidius Evans, 1955 (type species Narga vidius Mabille, 1891), but is not monophyletic with it (Fig. 11–12). Instead, it groups closely with Moeris Godman, [1900] (type species Talides striga Geyer, [1832]): compare with its sister, the Viridina, gen. n., cluster in Fig. 11 and 12. Despite obvious differences in wing patterns, the wing shapes and genitalia structures (even the shape of the valva with a somewhat expanded harpe) are quite similar between these species. The COI barcode difference between P. anna and M. striga is 7.4%, suggesting a close relationship. For these reasons, instead of proposing a new genus name for P. anna, we establish a new combination, Moeris anna.

Arita Evans, 1955 is a synonym of Tigasis Godman, [1900]

Inspection of *Tigasis* reveals its close relationship with *Arita* Evans, 1955 (type species *Cobalus arita* Schaus, 1902) (Fig. 11–12). For instance, COI barcodes of the type species in these genera differ by only about 3%. Therefore, we treat *Arita* as a new junior subjective synonym of *Tigasis*.

Alychna Grishin, new genus

http://zoobank.org/D9C177B1-6CE7-4A1E-9B76-BD7D29AE18E6

Type species. Pamphila exclamationis Mabille, 1898.

Definition. A sister genus of *Zalomes* Bell, 1947 (type species *Zalomes colobus* Bell, 1947, considered to be a junior subjective synonym of *Hesperia biforis* Weymer, 1890) and in a different clade from both *Psoralis* Mabille, 1904 (type species *Psoralis sabaeus* Mabille, 1904, which is a junior subjective synonym of *Pamphila idee* Weeks, 1901) and from *Lychnuchus* Hübner, [1831] (type species *Lychnuchus olenus* Hübner, [1831] considered to be a junior subjective synonym of *Hesperia celsus* Fabricius, 1793), where species of this genus were placed previously (Fig. 11–12). Keys to J.43.3d, J.43.6, or K.12.1 in Evans (1955). Morphologically, distinguished from its relatives by a combination of the following characters: uncus broad, the same width and length, if divided, arms very short, spike-like; gnathos not shorter than uncus, close to it in lateral view; valva broad, harpe not well-separated from ampulla, varying in shape depending on species, mostly rectangular with irregular margins, in the type species with a hook-like projections narrowly separated from harpe; stigma black, di- or tripartite; pale scales or hyaline spot distad of stigma on forewing; frequently a curved line of small pale spots in discal area of hindwing below. In DNA, a combination of the following base pairs is diagnostic: aly4645.10.1:T700C, aly4645.10.1:G701T, aly1877.13.1:T1118A, aly1877.13.1:G1159A, aly274.43.1:C170A.

Etymology. The name is a feminine noun in the nominative singular, formed from the first five letters of *Lychnuchus* (where one species was formerly placed) with "a" added on both sides to stress that it is not that genus.

Species included. The type species, *Lychnuchus victa* Evans, 1955, *Oeonus zenus* Bell, 1942, *Psoralis mirnae* Siewert, Nakamura and Mielke, 2014, *Psoralis darienensis* Gaviria, Siewert, Mielke and Casagrande, 2018, *Hesperia degener* Plötz, 1882, and *Psoralis venta* Evans, 1955.

Parent taxon. Subtribe Moncina A. Warren, 2008.

Comments. The two species *Lychnuchus celsus* and *Alychna victa* are so close to each other in wing patterns that their placement in two distantly related genera is totally unexpected. This is probably the most surprising discovery in our analysis. Retrospective analysis of genitalia reveals significant differences between these two species that agree with the DNA-guided placement (Fig. 11–12).

Molo pelta (Evans, 1955), new combination

To add to the *Lychnuchus* surprise, we transfer another mimic, *Lychnuchus pelta* Evans, 1955, to *Molo* Godman, [1900] (type species *Hesperia heraea* Hewitson, 1868, considered to be a junior subjective synonym of *Hesperia mango* Guenée, 1865), because it is not in the same clade as *Lychnuchus* (the type species of *Lychnuchus*) but instead is closely related to *Molo mango* (COI barcode difference only 2.7%, indicating congeneric relationship). Therefore, we form the new combination *Molo pelta* (Fig. 10–12).

Alychna zenus (E. Bell, 1942), reinstated species status

We conclude that *Oeonus zenus* E. Bell, 1942 (type locality Ecuador) is a species-level taxon, not a junior subjective synonym of *Pamphila exclamationis* Mabille, 1898 (type locality Bolivia). COI barcodes differ by more than 3% (Fig. 12), the stigma is broader and more curved in *A. zenus* compared to *Alychna exclamationis* and has more extensive black overscaling around it, and the hindwing is more produced at the tornus in *A. zenus*.

Ralis Grishin, new genus

http://zoobank.org/55719473-C71C-4492-B44C-00BB0CFFE700

Type species. Lerema coyana Schaus, 1902.

Definition. A genus without close relatives, but not monophyletic with *Psoralis* Mabille, 1904 (type species *Psoralis sabaeus* Mabille, 1904, which is a junior subjective synonym of *Pamphila idee* Weeks, 1901) (Fig. 11–12). Keys to J.43.9 in Evans (1955). Morphologically, distinguished from its relatives by a very conspicuous, large gray stigma and nearly unmarked dark brown hindwing ventral (sometimes with small pale dots). Male genitalia distinctive, see Nicolay (1980: fig. 7): uncus broad, not narrowing in the middle, tegumen with a pair of long dorsal processes directed caudad, gnathos the same length and width as uncus, prominent in lateral view, penis stout caudad with elongated phallobase, valva nearly oval, twice as long and wide with shirt harpe pointed dorsad, closely approaching ampulla (Nicolay 1980). In DNA, a combination of the following base pairs is diagnostic: aly25.7.1:A134C, aly2532.12.2:A142C, aly594.9.1:G589A, aly423.4.2:C199G, aly25.7.1:A226G.

Etymology. The name is a masculine noun in the nominative singular, denoting the second half of [Pso] *Ralis*, the genus where these species were placed previously.

Species included. The type species and Psoralis concolor Nicolay, 1980.

Parent taxon. Subtribe Moncina A. Warren, 2008.

Testia Grishin, new genus

http://zoobank.org/7063BAAB-1F7C-4EF5-905C-DF50960CCB02

Type species. Atrytone (?) potesta Bell, 1941.

Definition. A sister genus to Oxynthes Godman, [1900] (type species Goniloba corusca Herrich-Schäffer, 1869) combined with Oeonus Godman, [1900] (type species Oeonus pyste Godman, [1900]) and in the same clade with Lindra Evans, 1955 (type species Carystus simulius Druce, 1876) (Fig. 13). Not monophyletic with Orthos Evans, 1955 (type species Eutychide orthos Godman, [1900]), which is in a different subtribe (Carystina Mabille 1878). Keys to L.15.5 in Evans (1955). Distinguished from its relatives by a combination of the following characters (Bell 1941). Antennae about 2/3 of costa in length, male no secondary sexual characters, wings unmarked dark brown above with shiny green hair-like scales in tornal area of hindwing (and on body), with cream spots below: on hindwing near its middle plus diffuse cream area in distal half of CuA₂-1A+2A cell, small spots on hindwing in discal cell and as a postdiscal row in each cell between veins M_2 and 1A+2A. Male genitalia (see Bell 1941: fig. 3 for illustration) with uncus deeply divided, arms long (only slightly shorter than tegumen), parallel, near each other, penis widening terminally (nearly as wide as valva), bulky, with terminal spikes, valva nearly rectangular, sacculus without processes, harpe short (shorter than 1/3 of valva), angular, with a stout tooth pointed dorsad, separated from ampulla by a narrow notch. In DNA, a combination of the following base pairs is diagnostic: aly2874.22.9:G1030A, aly3512.12.2:T234C, aly2811.6.1:T166C, aly1139.93.1:C452C (not G), aly862.12.2:A1755A (not C), aly5021.5.1:G1325G (not C), aly537.7.1:A181A (not G), aly2012.14.2:T317T (not C), aly577.34.1:A485A (not T).

Etymology. The name is a feminine noun in the nominative singular, formed from the type species name.

Species included. Only the type species.

Parent taxon. Subtribe Hesperiina Latreille, 1809.

Buzella Grishin, new genus

http://zoobank.org/3B8255CA-FE24-4197-B04C-0876533EABF7

Type species. Buzyges mellanaformis Austin and A. Warren, 2009.

Definition. Not monophyletic with *Buzyges idothea* Godman, [1900], the type species of *Buzyges* Godman, [1900], this genus is placed in a different clade instead (Fig. 13). Detailed diagnosis of *mellanaformis* given by Austin and Warren (2009:28, fig. 70, 82, 90) can be applied to this genus (Austin

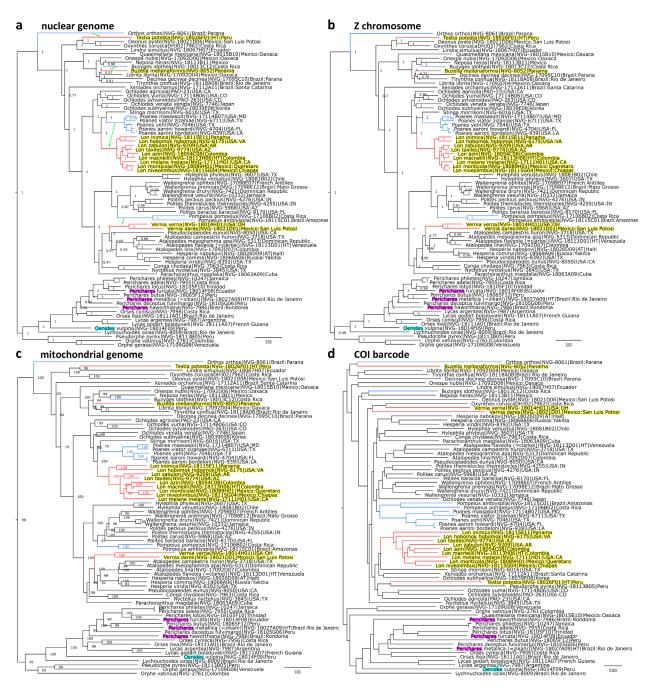


Figure 13. Genomic trees of Hesperiina, Pericharini and relatives. See Fig. 2 legend for notations.

and Warren 2009). In brief, differs from related genera by long antennae, the lack of stigma, short tegumen and uncus and relatively small valva with broad trapezoid harpe (shared with *Buzyges*), but penis narrower and cornuti are smaller than in *Buzyges* (see Fig. 70 and 82 in Austin and Warren (2009)), mid- and hind-tibiae without spines characteristic of *Buzyges* and forewing outer margin more convex. In DNA, a combination of the following base pairs is diagnostic: aly386.7.5:A874T, aly207.4.6:A367T, aly320.7.8:G428A, aly128.6.20:C131C (not G), aly708.6.1:A1197A (not G), aly1838.58.4:T1070T (not C), aly536.174.1:A1825A (not C), aly2582.33.4:A512A (not T), aly3071.1.1:C135C (not T).

Etymology. The name is a feminine noun in the nominative singular, formed from the type species original name Buz[yges m]ella[naformis].

Species included. Only the type species.

Parent taxon. Subtribe Hesperiina Latreille, 1809.

Vernia Grishin, new genus

http://zoobank.org/980834C1-C6C5-4172-B5BB-151246B12F1A

Type species. Pamphila verna Edwards, 1862.

Definition. A genus in the same clade with Hesperia Fabricius, 1793 (type species: $Papilio \ comma$ Linnaeus, 1758), Atalopedes Scudder, 1872 (type species $Hesperia \ huron$ Edwards, 1863), and Pseudocopaeodes Skinner and Williams, 1923 (type species $Copaeodes \ eunus$ Edwards, 1881), but not closely related with any particular one. Pompeius Evans, 1955 (type species $Hesperia \ pompeius$ Latreille, [1824]), where species from this genus were placed previously, is in a different clade from them (Fig. 13). Keys to M.15.2 or M.15.5 in Evans (1955). Distinguished from its relatives by divided uncus with short and stout arms; thin gnathos arms, longer than uncus; valva with harpe about half of its length, unturned, terminating with two broad teeth directed dorsocaudad, no bristles; apiculus of six segments, nudum not black; stigma tripartite on dorsal forewing of males; pale spot distad of stigma segment in dorsal forewing cell CuA_1 - CuA_2 occupies the whole width of the cell. In DNA, a combination of the following base pairs is diagnostic: aly443.32.2:G99C, aly2487.36.2:T119G, aly443.32.2:A97G, aly887.14.12:A1715G, aly2096.38.5:A32C.

Etymology. The name, a feminine noun in the nominative singular, reflects the type species name.

Species included. The type species and *Hesperia dares* Plötz, 1883.

Parent taxon. Subtribe Hesperiina Latreille, 1809.

Lon Grishin, new genus

http://zoobank.org/4CFECC3F-DEDA-4533-9F5C-4418BADA0ABB

Type species. Hesperia zabulon Boisduval and Le Conte, [1837].

Definition. A sister genus to *Stinga* Evans, 1955 (type species *Pamphila morrisoni* Edwards, 1878) combined with *Poanes* Scudder, 1872 (type species *Hesperia massasoit* Scudder, 1863) including its junior subjective synonym *Phycanassa* Scudder, 1872 (type species *Hesperia viator* Edwards, 1865) (Fig. 13). Species of this genus were formerly placed in *Poanes*, but are not monophyletic with it if *Stinga* is considered a valid genus, because *Stinga* is sister to *Poanes* with exclusion of this genus. This genus constitutes the "ordinary terrestrial species" subgroup of Burns (Burns 1992) who gave and illustrated diagnostic genitalic characters for it, in contrast to "marsh dwellers" that are the true *Poanes*. Most obvious diagnostic character is a very long (extending well beyond the body of aedeagus) and armed with spikes titillator in penis (in addition to two smaller ones), so prominent that it is even shown on Evans sketches (Evans 1955). Harpe upturned and touches ampulla, instead of being directed largely distad and leaving a gap between it at ampulla in *Poanes*. In DNA, a combination of the following base pairs is diagnostic: aly525.55.2:G476C, aly378.21.7:A1612G, aly84.28.1:A602G, aly2631.4.13:T68A, aly85.22.2:T1259C.

Etymology. The name is a masculine noun in the nominative singular, and is formed from the last syllable of the type species name.

Species included. The type species, Hesperia hobomok T. Harris, 1862, Pamphila inimica Butler and Druce, 1872, Pamphila taxiles Edwards, 1881, Pamphila azin Godman, 1900, Poanes macneilli Burns, 1992, Hesperia ulphila Plötz, 1883, Atrytone monticola Godman, [1900], Pamphila niveolimbus Mabille, 1889, Hesperia melane Edwards, 1869.

Parent taxon. Subtribe Hesperiina Latreille, 1809.

Alera Mabille, 1891 is a synonym of Perichares Scudder, 1872

We find (Fig. 13) that Alera furcata Mabille, 1891 and Perichares philetes (Gmelin, [1790]), the type species of both genera (technically, Papilio coridon Fabricius, 1775, a homonym, considered to refer to Perichares philetes, is the type species of Perichares) are genetically close (COI barcodes differ by about 6.5%). Moreover, several species of Alera that we sequenced are not monophyletic and are interspersed between species of Perichares. For these reasons, we consider Alera to be a subjective junior synonym of Perichares.

Oenides Mabille, 1904 is a valid genus

Hesperia vulpina C. and R. Felder, 1867 is not monophyletic with Alera furcata, the type species of Alera, where H. vulpina was previously placed, and since it is the type species of the available genus-group name Oenides Mabille, 1904, that was considered a subjective synonym of Alera, we resurrect this genus from synonymy and use it as a monotypic valid genus with the species Oenides vulpina (Fig. 13).

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Table 1. Data for 620 sequenced Hesperiidae specimens. See Table S1 in the Supplemental file deposited at https://osf.io/5cfht/?view_only=21eb53b6f8f344afaee3de2be90bf5d2 for detailed information about these specimens and Materials and Methods section for collection abbreviations. Type status abbreviations are: AT, allotype; HT, holotype; LT, lectotype; PT, paratype, ST, syntype; T, type.

DNA voucher	Taxon name	Type	Brief data	Collection
NVG-18054G10	Acada biseriata		Zimbabwe, 1993	ZMHB
NVG-18081B11	Acerbas anthea		Malaysia, 1917, NHMUK_010430824, 0247275554	BMNH
NVG-17091E09	Acerbas duris duris		Philippines, old (around 1900)	USNM
NVG-18074B03	Acerbas sarala sarala		India, 1890	ZMHB
NVG-18011F11	Achlyodes busirus busirus		French Guiana, 1988	USNM
NVG-18081B04	Acleros leucopyga		Madagascar, 1921, NHMUK_010430828, 0247274717	BMNH
NVG-18081D02	Actinor radians		India, 1885, NHMUK_010430809, 0247275546	BMNH
NVG-18015A06	Adlerodea petrovna	Т	Brazil: Rio de Janeiro, old (around 1900)	USNM
NVG-7988	Adopaeoides prittwitzi		USA: AZ, Santa Cruz Co., 1999	USNM
NVG-7915	Aeromachus stigmata shanda		Myanmar, 2002	USNM
NVG-18038C05	Aethilla chiapa		Honduras	RGallardo
NVG-18013G10	Aethilla echina		Costa Rica, 2005, 05-SRNP-20	USNM
NVG-18011F09	Aethilla eleusinia		Peru, 2012	USNM
NVG-18056D08	Aethilla gigas		Bolivia, old (around 1900)	ZfBS
NVG-7891	Aethilla lavochreaDHJ02		Costa Rica, 2013, 13-SRNP-22231	USNM
NVG-14064D01	Aethilla memmius		Venezuela, 1985	USNM

DNA voucher	Taxon name	Type	Brief data	Collection
NVG-18012E06	Aides duma argyrina		Bolivia?, no date?1970	USNM
NVG-18054H04	Alenia namaqua		South Africa, 2002	ZMHB
NVG-15035E02	Alychna degener	ST	no data	ZMHB
NVG-18042G08	Alychna exclamationis	ST	Bolivia, old (around 1900)	ZMHB
	Alychna victa		Ecuador, 2004	USNM
NVG-18027A06	-	нт	Ecuador, 1939	AMNH
	Amblyscirtes vialis		USA: CO, Grand Co., 2016	UTSW
	Ampittia dioscorides singa		Sri Lanka, 1971	USNM
NVG-18012A02	1 0		Costa Rica, 1977	USNM
NVG-18013F12			Costa Rica, 2008, 08-SRNP-2106	USNM
NVG-18011H11	Anastrus obscurus		Brazil: Rondonia, 1991	USNM
NVG-18067H10			Brazil: Santa Catarina, 2000	EBrockmann
NVG-18056D06	1		Bolivia, old (around 1900)	ZfBS
NVG-4461	1		USA: TX, Dallas Co., 2015	UTSW
	Anisochoria pedaliodina pedalio-			
NVG-18091C07	dina		Ecuador, 2012	EBrockmann
NVG-7909	Anisochoria pedaliodina polysticta		Costa Rica, 2004, 04-SRNP-15751	USNM
NVG-18011G07	Anisochoria quadrifenestrata		Colombia, old (around 1900)	USNM
NVG-18019B02			Australia, 1961	AMNH
NVG-17069F11	Ankola fan		Kenya, 1955	USNM
NVG-17111E11	-		Brazil: Santa Catarina, 1999	LACM
	Antigonus emorsa		Mexico: Michoacan, 1994	TAMU
	Antigonus erosus		Costa Rica, 2013, 13-SRNP-56479	USNM
NVG-18014A03			Costa Rica, 2006, 06-SRNP-32799	USNM
	Antipodia chaostola		Australia, 1934, NHMUK_012824109, 0247278477	BMNH
NVG-18079E09	Apallaga fulgens fulgens		Cameroon, 1987, EL63116	MNHP
NVG-18011B09			South Africa, 1978	USNM
NVG-17108C09			Kenya, 2001	LACM
NVG-18033B11			Uganda, 2008	MWalker
NVG-18055G03	Apallaga pooanus obscuripennis	ST	Equatorial Guinea, 1906	ZMHB
	Apallaga rutilans rutilans		Cameroon, 1991, EL63122	MNHP
	Apaustus menes		Peru, 2011	USNM
	Argemma argyrosticta		Cameroon, old (around 1900)	USNM
	Argon lota		Costa Rica, 2007, 07-SRNP-55877	USNM
	Argopteron aureipennis		Chile, 1982	USNM
NVG-18019B09	Arnetta atkinsoni		India, 1927	AMNH
NVG-5065			Costa Rica, 04-SRNP-1707	USNM
NVG-5485	Arteurotia tractipennis tractipen- nis		USA: TX, Hidalgo Co., 1972	TAMU
NVG-18012H06			Brazil: Santa Catarina, 1999	USNM
NVG-18067F11	Artitropa erinnys		Comoros, 1987	EBrockmann
NVG-19022F06			Brazil: Parana, 1995	USNM
NVG-4881	Asbolis capucinus		USA: FL, Monroe Co., 2015	UTSW
NVG-7394	-		Myanmar, 2001	USNM
NVG-3718			USA: TX, Dallas Co., 2015	UTSW
NVG-18113D01	Atalopedes flaveola (=clarkei)	НТ	Venezuela, 1985	USNM
NVG-17092D07	Atalopedes lina	1	Colombia, 1991	USNM
NVG-5313	_		Dominican Republic, 2015	UTSW
NVG-7903			Costa Rica, 2007, 07-SRNP-59529	USNM
NVG-17119H02			Myanmar, 2002	USNM
1110-111101102	21ai voima aai milata	1	Malaysia, 2003	OBINIVI

NVG-14101B01 Austinus heroica Colombia, 1946 NVG-18067C09 Austinus heros Brazil: Para, 2009 NVG-18014E05 Avestia avesta Malaysia, old (around 1900) NVG-17091G11 Baoris oceia Phillipines, 1914 NVG-18089F07 Barca bicolor China: Shaanxi, 2009 NVG-18026C12 Barrolla molla HT Ecuador NVG-7246 Bolla brennus brennus Panama, 1981 NVG-17108F05 Bolla cylindus Mexico: Hidalgo, 1982 NVG-18071A09 Bolla evippe Costa Rica, 2009, 09-SRNP-57008 NVG-1801H02 Bolla imbras Panama, 1984 NVG-18083B12 Bolla saletas T Guatemala, BMNH(E)_1669828, 0247277236 NVG-18067H11 Bralus albida albida Peru, 1994 NVG-18025F01 Brownus browni HT Costa Rica, 2006 NVG-17095E11 Brownus browni Costa Rica, 2006	AMNH EBrockmann
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NVG-17108F05 Bolla cylindus Mexico: Hidalgo, 1982 NVG-18071A09 Bolla evippe Costa Rica, 2009, 09-SRNP-57008 NVG-18011H02 Bolla imbras Panama, 1984 NVG-18083B12 Bolla saletas T Guatemala, BMNH(E)_1669828, 0247277236 NVG-18067H11 Bralus albida albida Peru, 1994 NVG-18025F01 Brownus browni HT Costa Rica, 1946	AMNH
NVG-18071A09 Bolla evippe Costa Rica, 2009, 09-SRNP-57008 NVG-18011H02 Bolla imbras Panama, 1984 NVG-18083B12 Bolla saletas T Guatemala, BMNH(E)_1669828, 0247277236 NVG-18067H11 Bralus albida albida Peru, 1994 NVG-18025F01 Brownus browni HT Costa Rica, 1946	USNM
NVG-18011H02 Bolla imbras Panama, 1984 NVG-18083B12 Bolla saletas T Guatemala, BMNH(E)_1669828, 0247277236 NVG-18067H11 Bralus albida albida Peru, 1994 NVG-18025F01 Brownus browni HT Costa Rica, 1946	LACM
NVG-18011H02 Bolla imbras Panama, 1984 NVG-18083B12 Bolla saletas T Guatemala, BMNH(E)_1669828, 0247277236 NVG-18067H11 Bralus albida albida Peru, 1994 NVG-18025F01 Brownus browni HT Costa Rica, 1946	USNM
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NVG-18025F01 Brownus browni HT Costa Rica, 1946	EBrockmann
	AMNH
IN VIG-17033E111 DIOWINGS OFOWNI USE A KICA, ZUUO	USNM
NVG-17093D04 Burca concolor concolor Cuba, 2010	USNM
NVG-7552 Burnsius communis USA: TX, Bexar Co., 1977	TAMU
NVG-16108G04 Butleria flavomaculatus valdivianus Chile, 1979, USNMENT 00894446	USNM
NVG-8052 Buzella mellanaformis Panama, 1982	USNM
NVG-18013C12 Buzyges idothea Costa Rica, 1980	USNM
NVG-5721 Cabirus procas Ecuador, 2004	USNM
NVG-7754 Caenides dacela Liberia, 1988	USNM
NVG-17092C10 Callimormus juventus Costa Rica, 2012, 12-SRNP-20224	USNM
NVG-4591 Calpodes ethlius USA: TX, Cameron Co., 2015	UTSW
NVG-18013G01 Camptopleura theramenes Costa Rica, 2015, 15-SRNP-45798	USNM
NVG-7905 Canesia canescens Costa Rica, 2014, 14-SRNP-1649	USNM
NVG-18013H12 Canesia meridensis Costa Rica, 1997, 97-SRNP-1522	USNM
NVG-18012F06 Cantha calva Peru, 1986	USNM
NVG-18033B07 Capila phanaeus Cambodia, 2006	MWalker
NVG-7763 Carcharodus alceae Greece, 1990	USNM
NVG-18014A02 Carrhenes fuscescens Costa Rica, 1995, 95-SRNP-6819	USNM
NVG-18089C06 Carterocephalus alcina China: Sichuan, 2006	EBrockmann
NVG-18021G04	AMNH
NVG-17119H05 Carterocephalus christophi China, old (around 1900)	USNM
NVG-18089B05 Carterocephalus gemmatus China: Sichuan, 2002	EBrockmann
NVG-18089B08	EBrockmann
PAO-69 Carterocephalus mandan USA: CA, Sierra Co., 2016	UTSW
NVG-18089C04 Carterocephalus micio China: Yunnan, 2006	EBrockmann
NVG-18038G04 Carterocephalus silvicola Russia: Buryatia, 2016, 5195	UTSW
NVG-18012D09 Carystina lysiteles Peru, 1999	USNM
NVG-18012D05 Carystus jolus Colombia, 1971	USNM
NVG-18011B06 Celaenorrhinus aegiochus Panama, 1981	USNM
NVG-18064A03 Celaenorrhinus bettoni Uganda, 1952	USNM
NVG-18013G07 Celaenorrhinus eligius Costa Rica, 2010, 10-SRNP-20588	USNM
NVG-17093D03 Celaenorrhinus ficulnea ficulnea Malaysia, old (around 1900)	USNM
NVG-18011B04 Celaenorrhinus monartus Panama, 1981	USNM
NVG-18073G09 Celaenorrhinus rosetta Gabon, old (around 1900)	ZMHB
NVG-18011B08 Celaenorrhinus spilothyrus Sri Lanka, 1976	USNM
NVG-18019A07 Celaenorrhinus sumitra Nepal, 1937	AMNH
NVG-7993 Celaenorrhinus syllius Ecuador, 2002, USNMENT 01321833	USNM

DNA voucher	Taxon name	Type	Brief data	Collection
11-BOA- 13383BrockB12	Celotes limpia		USA: TX, Brewster Co.	JPBrock
NVG-3956	Celotes nessus		USA: TX, Hidalgo Co., 2015	UTSW
NVG-17067E03	Celotes spurcus	PT	Mexico: Queretaro, 2007, CSU_ENT1024631	CSUC
NVG-5684	Cephise aelius		Costa Rica, 2014, 14-SRNP-70854	USNM
NVG-7398	Ceratrichia flava		Cameroon, 1989	USNM
NVG-18053E03	Ceratrichia nothus		no data	ZMHB
NVG-7401	Ceratrichia wollastoni		Cameroon, 1989	USNM
NVG-18087B09	Ceratricula semilutea		Sierra Leone, 1906, EL62955	MNHP
NVG-18089G07	Cerba martini		Malaysia, 1993	EBrockmann
NVG-18039B11	Chaetocneme beata		Australia, old (around 1900)	FMNH
NVG-17108C12	Chaetocneme helirius		British New Guinea, 1962	LACM
NVG-17069A11	Chamunda chamunda		India, old (around 1900)	USNM
NVG-7981	Charidia lucaria		Peru, 2008	USNM
NVG-18057A01	Chiomara gundlachi		Cuba, 2014	ZSMC
NVG-17109F12	Chiomara mithrax		USA: AZ, Santa Cruz Co., 1992	LACM
NVG-14103A02	Chiothion asychis asychis		Suriname	USNM
NVG-17109F10	v v	1	Brazil: Goias, 1956	LACM
NVG-5100			USA: TX, Starr Co., 2015	UTSW
NVG-18081D11	Chiothion khalili		Brazil: AM, 1929, NHMUK_010430875, 0247277240	BMNH
NVG-14102F02	Chirgus limbata		Chile, 1952	FMNH
NVG-18025D04		нт	Brazil: Santa Catarina, old (around 1900)	AMNH
NVG-5271	` ′		China: Sichuan Prov., 2015	UTSW
NVG-17069G10	Chondrolepis niveicornis		Kenya, 1951	USNM
NVG-18117E12	Choranthus radians		Cuba, 2010	USNM
NVG-14107C04	Clito aberrans-cf		Peru, 1982	USNM
NVG-15117B05			Mexico: Sinaloa, 2003, CSU_ENT1039479	CSUC
NVG-17108F02			USA: AZ, Santa Cruz Co., 1991	LACM
NVG-18013A06	Cobalopsis autumna		Costa Rica, old (around 1900)	USNM
NVG-7927	Cobalus virbius		Costa Rica, 2012, 12-SRNP-22162	USNM
NVG-3354			USA: TX, Hidalgo Co., 2015	UTSW
NVG-7963			Costa Rica, 2009, 09-SRNP-68418	USNM
NVG-14107B12			Ecuador, 1989	USNM
NVG-15104C09		нт	Dominican Republic	AMNH
NVG-8381			USA: TX, Blanco Co., 2017	UTSW
NVG-18063D07			French Guiana, 2004, H21114	BHermier
NVG-18071C04	Corra coryna conka		Costa Rica, 2007, 07-SRNP-35032	USNM
NVG-17092F04	-		Costa Rica, 2012, 12-SRNP-70085	USNM
NVG-14064H09	Crenda crenda		Brazil: Parana, old (around 1900)	USNM
NVG-18081D10	Crenda crenda	нт	Brazil: Parana, NHMUK_010430876, 0247278431	BMNH
NVG-17092G08	Cumbre cumbre		Brazil: Rio de Janeiro, 1994	USNM
NVG-7381	Cupitha purreea		Myanmar, 2001	USNM
NVG-15117B07	Cycloglypha thrasibulus thrasi- bulus		Mexico: Sinaloa, 2003, CSU_ENT1039523	CSUC
NVG-17105F01	Cyclosemia herennius		Guyana, 2000	USNM
NVG-4842	Cymaenes tripunctus tripunctus		USA: FL, Collier Co., 2015	UTSW
NVG-7960	Cynea cynea		Costa Rica, 2010, 10-SRNP-35740	USNM
NVG-18054C11	Dalla agathocles agathocles		Colombia, 1921	ZMHB
NVG-18017B10	Dalla agathocles lanna		Ecuador, 1993	USNM
NVG-18014F05	Dalla caicus inca		Peru, 2011	USNM
NVG-18014E11	Dalla costala		Peru, 2011	USNM

DNA voucher	Taxon name	Type	Brief data	Collection
NVG-18014F06	Dalla cyprius quinka		Peru, 2013	USNM
NVG-18014E12	Dalla dimidiatus lilla		Peru, 2011	USNM
NVG-18017B08	Dalla dimidiatus pucer		Peru, 1992	USNM
NVG-18073F06	Dalla eryonas		Panama, old (around 1900)	ZMHB
NVG-18014F03	Dalla frater		Peru, 2013	USNM
NVG-18017B04	Dalla frontinia-cf		Ecuador, 1986	USNM
11-BOA- 13383BrockB09	Dalla nona		Peru	JPBrock
NVG-18017B05	Dalla pincha		Ecuador, 1982	USNM
NVG-18017B12	Dalla semiargentea		Colombia, 1965	USNM
NVG-17111B10	Damas clavus		Brazil: Rondonia, 1993	LACM
NVG-7929	Damas immacula		Costa Rica, 2010, 10-SRNP-67003	USNM
NVG-7349	Dardarina dardaris		Costa Rica, 2004, 04-SRNP-13073	USNM
NVG-18114D07	Daron seron sexton		Ecuador, 2001	USNM
NVG-7330	Darpa striata striata		Malaysia, old (around 1900)	USNM
NVG-17095C10	Decinea decinea decinea		Brazil: Parana, 1995	USNM
11-BOA-			D	
13385E10	Diaeus lacaena		Brazil: Rio de Janeiro, 1996	USNM
NVG-7893			Costa Rica, 2004, 03-SRNP-23600	USNM
NVG-18054F12	Dotta callicles		Namibia, 1992	ZMHB
NVG-17093B03	Dotta stellata stellata		Kenya, 1957	USNM
NVG-8038	Dubia dubia		Guyana, 1999	USNM
NVG-18012D08	Dubiella dubius		Peru, 1989	USNM
NVG-17098F01	Duroca duroca duroca		Brazil: Rio de Janeiro, 1996, USNMENT 00913432	USNM
NVG-17108B11	Eagris tigris		Kenya, 1985	LACM
NVG-15033A04	Eantis minna		Brazil: Amazonas, 1886	ZMHB
NVG-18025A03	Eantis minor	HT	Dominica, 1934	AMNH
NVG-10487	Eantis mithridates		Jamaica, 2017	UTSW
NVG-18025A06	Eantis munroei	HT	Cuba	AMNH
NVG-1931	Eantis pallida		Mexico: Tamaulipas, 1974	TAMU
NVG-3758	Eantis tamenund		USA: TX, Hidalgo Co., 2015	UTSW
NVG-1900	Eantis thraso		Brazil: Rondonia	TAMU
NVG-16108E06	Eantis tosta		Peru, 1999	USNM
NVG-18012A05	Ebrietas osyris		Mexico: Oaxaca, 1992	USNM
NVG-7977	Eburuncus unifasciata		Panama, 1984	USNM
NVG-7920	Ebusus ebusus		Costa Rica, 2013, 13-SRNP-30669	USNM
NVG-7884	Echelatus sempiternus		Costa Rica, 2007, 07-SRNP-12147	USNM
NVG-18013B09	Enosis dognini		Peru, 1998	USNM
NVG-5730	Entheus Burns01	1	Costa Rica, 2008, 08-SRNP-35619	USNM
NVG-18019B11	Eogenes alcides		Turkey, old (around 1900)	AMNH
NVG-17093E07	Ephyriades arcas		St. Croix, 1996	USNM
NVG-7897	Eracon sarahburnsae		Costa Rica, 2010, 10-SRNP-67957	USNM
NVG-7910	Erionota thrax	1	USA: HI, Molokai, 2005	USNM
NVG-18068D04	Erynnis tages	1	Russia: Siberia, 1999	EBrockmann
NVG-15103B05	Euschemon rafflesia rafflesia		Australia	USNM
NVG-17092F09	Eutocus facilisDHJ01	1	Costa Rica, 2006, 06-SRNP-47351	USNM
NVG-18082B04	Exometoeca nycteris		Australia, NHMUK_010430873, 0247277190	BMNH
NVG-18012E10	Falga jeconia jeconia		Venezuela, 1985	USNM
NVG-18031H12	Festivia adamantinus		Ecuador, 2002	USNM
NVG-2073			Brazil: Parana, 2011	MEM
NVG-18031H06	Festivia festiva		Ecuador, 1990	USNM
NVG-18032A03	Festivia grippa		Ecuador, 2002	USNM
			1 Control of the Cont	1

DNA voucher	Taxon name	Туре	Brief data	Collection
NVG-19017F02	Fidius fido		Brazil: Parana, old (around 1900)	USNM
NVG-19017E10	Flaccilla aecas		Guyana, 2000	USNM
NVG-14101G07	Flattoides amazonensis amazonensis	нт	Colombia, 1946	AMNH
NVG-17092A07	Fresna netopha		Uganda, 1960	USNM
NVG-7762	Fulda coroller		Madagascar, 1991	USNM
NVG-7778	Fulda rhadama		Madagascar, 1990	USNM
NVG-7808	Galerga hyposticta		Madagascar, 1990	USNM
NVG-8020	Gallio gallio		Peru, 1986	USNM
NVG-16108E10	Gamia shelleyi		Uganda, 1953	USNM
NVG-16108F09	Gangara thyrsis		Philippines, 1987	USNM
NVG-18057H08	Ge geta		no data, 1894	ZSMC
NVG-7336			Myanmar, 2003	USNM
NVG-7570			Dominican Republic, 1981	TAMU
NVG-18011E10	Gindanes brebisson		Peru, 2016	USNM
NVG-7901	Gindanes brontinus		Costa Rica, 2008, 08-SRNP-57426	USNM
NVG-18067H12	Gindanes kelso		Peru, 2000	EBrockmann
NVG-18026G08	Ginungagapus schmithi	нт	Brazil: Santa Catarina	AMNH
NVG-18087G04			Botswana, 1997	EBrockmann
NVG-18062A03	Gorgopas chlorocephala chloro- cephala		Peru, 2016	USNM
NVG-18019E12	Gorgyra aburae		Uganda, 1954	AMNH
NVG-17067D06	Gorgythion begga pyralina		Mexico: Sinaloa, 2003, CSU_ENT1039357	CSUC
NVG-18091C03	Gorgythion beggina escalophoides		Ecuador, 2012	EBrockmann
NVG-15101B07	Gorgythion plautia (=beggoides)	ST	Trinidad, Type No. 5981 U.S.N.M.	USNM
NVG-7880	Grais stigmaticus	DI	Costa Rica, 2014, 14-SRNP-30242	USNM
NVG-18019F12	Gretna cylinda		Uganda, 1953	AMNH
NVG-18053D11	Gyrogra subnotata		Gabun, 1892	ZMHB
NVG-7979	Haemactis sanguinalis		Ecuador, 1991	USNM
NVG-18013A12	Halotus angellus		Panama, 1976	USNM
NVG-17119G01	Halpe porus		Myanmar, 2001	USNM
NVG-18052D09	Haza hazarma	LT	no data	ZMHB
NVG-15039D09	Helias phalaenoides phalaenoides		Brazil: Rondonia, 1993	FMNH
NVG-7558	Heliopetes arsalte		Mexico: San Luis Potosi, 1980	TAMU
NVG-18064E01	Herila herilus		Tanzania, 1951	USNM
NVG-18068A06			Russia: Yakutia, 1990	EBrockmann
NVG-4767			USA: FL, Levy Co., 2015	UTSW
NVG-18026D09	Hesperia nabokovi	AT	Haiti, 1922	AMNH
NVG-8392	Hesperia viridis	711	USA: TX, Blanco Co., 2017	UTSW
NVG-17069D12	Hesperilla ornata		Australia, old (around 1900)	USNM
NVG-17067A10	Hesperopsis alpheus alpheus		USA: CA, Kern Co., 2012, CSU_ENT1039256	CSUC
NVG-17067B02	Hesperopsis gracielae		USA: CA, Riverside Co., 1997, CSU_ ENT1039348	CSUC
NVG-17067A09	Hesperopsis libya libya		USA: CA, Inyo Co., 2009, CSU_ENT1039161	CSUC
NVG-17069E08	Heteropterus morpheus		France, 1966, USNMENT 00894386	USNM
NVG-17108D07	Hewitsoniella migonitis		Papua New Guinea, 1996	LACM
NVG-7823	Hidari irava		Singapore, 1989	USNM
NVG-18072H08	Hollandus xanthopeplus		Cameroon, 1895	ZMHB
NVG-18072H07	Hollandus xanthopeplus		Equatorial Guinea, 1906	ZMHB
NVG-7882	Hoodus pelopidas		Costa Rica, 2008, 08-SRNP-55556	USNM
NVG-7816			Madagascar, 1988	USNM
NVG-7767	Hovala pardalina		Madagascar, 1988	USNM
11101	Hovala saclavus		Madagascar, 1988	USNM

DNA voucher	Taxon name	Туре	Brief data	Collection
NVG-3607	Hylephila phyleus		USA: TX, Starr Co., 2015	UTSW
NVG-18081B02			Chile, 1960, NHMUK_010430840, 0247274697	BMNH
NVG-17091H02			Uganda, 1960	USNM
NVG-14103B04	Iliana romulus		Peru	USNM
NVG-17091E06	Ilma irvina		Indonesia, old (around 1900)	USNM
NVG-18059B11	Incisus incisus		Brazil: Rio de Janeiro, 1995	USNM
NVG-18025A09	Isoteinon abjecta (=niangarensis)	нт	Niangara, 1910	AMNH
NVG-17091A07	Isoteinon lamprospilus formosa- nus		Taiwan, 1980	USNM
NVG-14107C02	Jera tricuspidata		Ecuador, 1984	USNM
NVG-7953	-		Costa Rica, 2004, 04-SRNP-14377	USNM
NVG-18013B06	, v		Guyana, 2000	USNM
NVG-18053B08			Sierra Leone, 1887	ZMHB
NVG-17108F10			South Africa, 1943	LACM
NVG-17069F06	Kobelana kobela		South Africa, 1978	USNM
NVG-18093B06		LT	Colombia	SMF
NVG-18033B00		121	Peru, 2008	USNM
NVG-18014F01			Peru, 2010	USNM
NVG-18014F01 NVG-18093B08	Ladda ochrolimbata	HT	Peru, old (around 1900)	SMF
NVG-18093B08		пі		-
	1		Peru, 2013	USNM
NVG-18017B09	1 1		Ecuador, 1993	USNM
NVG-17111D10	1		Ecuador, 1992	LACM
NVG-18014E10	1 0		Peru, 2013	USNM
NVG-18017B07			Ecuador, 1986	USNM
8041	1 1		Brazil: Parana, 1995	USNM
NVG-18012F02			Ecuador, 1998	USNM
NVG-18019G02			Malawi, 1938	AMNH
NVG-17069C05	Leptalina unicolor		Japan, 1951	USNM
NVG-1769			USA: TX, Dallas Co., 2013	USNM
NVG-7738	Lerema lineosa		Brazil: Mato Grosso, 1991	USNM
NVG-4062	Lerodea eufala		USA: TX, Dallas Co., 2015	UTSW
NVG-18054G05	Leucochitonea levubu		Namibia, 2002	ZMHB
NVG-17092D04	Librita librita		Mexico: Oaxaca, 1992	USNM
NVG-18067H07	Lindra simulius		Ecuador, 2012	EBrockmann
NVG-16108E11	Lissia lissa lima		Kenya, 1956	USNM
NVG-18056B11	Livida assecla		Brazil: Goias, 1929	ZfBS
NVG-3288	Lobocla liliana liliana		China: Yunnan, 2009	UTSW
NVG-18054C08	Lon azin		Colombia, 1920	ZMHB
NVG-6175	Lon hobomok hobomok		USA: VA, Augusta Co., 2016	UTSW
NVG-18115E11	Lon inimica		Panama, 2007	USNM
NVG-18113H06	Lon macneilli	нт	Colombia, 1975	USNM
NVG-17111H01	Lon melane melane		USA: CA, San Luis Obispo Co., 1994	LACM
NVG-18089H01	Lon monticola		Mexico: Queretaro, 2004	EBrockmann
NVG-18115G04			Mexico: Chiapas, 1992	USNM
NVG-9774			USA: AZ, Cochise Co., 2017	UTSW
NVG-9209			USA: AR, Montgomery Co., 2017	UTSW
NVG-7373			Myanmar, 2003	USNM
8026	_		Brazil: Minas Gerais, 1994	USNM
NVG-7940			Costa Rica, 2012, 11-SRNP-33493	USNM
	Lurida lurida (=carinna)	Т	Brazil: Parana, old (around 1900)	USNM
	Lycas argentea	1	Argentina, 1998	USNM
	Lycas godart boisduvalii		French Guiana, 1993	USNM
NVG-18111A07			Brazil: Rio de Janeiro, 1995	USNM
TA A CE-0008	пусынисноваев огная		Diami. Ino de ganeiro, 1999	OBMM

DNA voucher	Taxon name	Type	Brief data	Collection
NVG-2076	Lychnuchus celsus		Brazil: Parana, 2011	MEM
NVG-5688	Marela tamba		Ecuador, 1991	USNM
NVG-7787	Matapa aria		Philippines, 1986	USNM
			Myanmar, 2001	USNM
	Megathymus yuccae yuccae		USA: SC, Aiken Co., 2013	UTSW
			Nigeria, 1958, NHMUK_010430880, 0247278457	BMNH
NVG-18074H11	Melphinyet tarace	Т	Sierra Leone, 1889	ZMHB
NVG-17108A04	Methion melas		Guatemala, 1963	BMUW
NVG-18064B11	Methionopsis ina		Costa Rica, 2010, 10-SRNP-43176	USNM
NVG-18054G04	Metisella medea		Malawi, 1996	ZMHB
NVG-17093A02	Metisella metis paris		Uganda, 1958	USNM
			Malawi, 1996	ZMHB
NVG-17091H09	Meza meza		Cameroon, old (around 1900)	USNM
NVG-7874	Mictris crispus		Costa Rica, 2009, 09-SRNP-71399	USNM
	Mielkeus diana diana (=seitzi)	LT	Brazil: Rio Grande do Sul, old (around 1900)	SMF
	, ,	<u> </u>	Guyana, 1999	USNM
			Brazil: Rio de Janeiro, 1995	USNM
			Costa Rica, 2010, 10-SRNP-72549	USNM
			Costa Rica, 2005, 05-SRNP-41228	USNM
	Miltomiges cinnamomea		Brazil: Rio de Janeiro, 1995	USNM
	Miraja varians		Madagascar, 1988	USNM
	<u> </u>		Guyana, 2001	USNM
NVG-17111C02			Mexico: San Luis Potosi, 1981	LACM
	Mnasilus allubita		Costa Rica, 2002, 02-SRNP-13739	USNM
		+	Panama, 1996	USNM
	Mnestheus ittona		Peru, 2012	USNM
	Moeris anna		Brazil: Rio de Janeiro, 1994	USNM
	Moeris striga	+	French Guiana, 1993	USNM
	Moeris striga Moeris stroma	+	Costa Rica, 2013, 13-SRNP-56538	USNM
NVG-15036C07		Т		ZMHB
	Moeros moeros Molo mango	1	Suriname, 1874	FMNH
			Guyana, 2003 Peru, 2012	USNM
NVG-18012D03	<u> </u>			
NVG-18019F10		-	Natal, 1924	AMNH
	Monca crispinus	+	Costa Rica, 2006, 06-SRNP-55847	USNM
NVG-17069H12		+	Nigeria, 1951	USNM
NVG-17069H09		-	Cameroon, old (around 1900)	USNM
i	Morvina morvus morvus		Colombia, 1945	AMNH
	Morys valerius		French Guiana, 1993	USNM
			Colombia, 1992	USNM
	Muschampia proto		France, 2012	EBrockmann
NVG-18015C07	Mylon ander ander		Guyana, 2000	USNM
	Mylon cajus cajus		Peru, 2016	USNM
			Peru, 2016	USNM
NVG-7881	Mylon lassia		Costa Rica, 2009, 09-SRNP-36601	USNM
NVG-18013F07	Mylon maimon		Costa Rica, 2015, 15-SRNP-21203	USNM
NVG-18015C05	Mylon mestor		Colombia, 1992	USNM
	Mylon orsa		Panama, 1975	USNM
	Mylon salvia		Costa Rica, 2006, 06-SRNP-1769	USNM
NVG-18015C01	Mylon zephus albodiscus		Peru, 2008	USNM
11-BOA- 13385F09	Myrinia myris		Brazil: Rondonia, 1989	USNM
NVG-7950	Naevolus orius		Costa Rica, 2010, 10-SRNP-72281	USNM

DNA voucher	Taxon name	Type	Brief data	Collection
NVG-8137	Nastra lherminier		USA: FL, Liberty Co., 2017	UTSW
NVG-18053B01	Neohesperilla croceus		Australia, 1892	ZMHB
NVG-17095F04	Neoxeniades ethoda		Brazil: Santa Catarina, 1991	USNM
NVG-18069F03	Neoxeniades luda		Costa Rica, 2012, 12-SRNP-1402	USNM
NVG-18082E06	Neoxeniades musarion		Brazil: RJ, NHMUK_012824133, 0247279800	BMNH
NVG-7936	Neoxeniades pluviasilva		Costa Rica, 2012, 12-SRNP-30105	USNM
NVG-17112B12	Neoxeniades turmada		Peru, 1986	LACM
NVG-18113B11	Neposa heras		Mexico, old (around 1900)	BMNH
NVG-14063B05	Nerula fibrena		Venezuela, 1985	USNM
NVG-17091B06	Nervia nancy		Kenya, 1960	USNM
NVG-16106A03	Netrocoryne repanda		Australia, 1963	LACM
NVG-18053D01	Netrocoryne thaddeus		Indonesia, 1894	ZMHB
NVG-18013B02	Niconiades xanthaphes		Guyana, 2000	USNM
NVG-18011G09	Nisoniades mimas		Peru, 1982	USNM
NVG-18015E03	Noctuana lactifera lactifera		Costa Rica, 2003, 03-SRNP-22972	USNM
NVG-18086D12	Noctulana noctula		Ivory Coast, EL63203	MNHP
NVG-3845			USA: TX, Cameron Co., 2015	UTSW
NVG-15036C01	Nyctus crinitus	ST	Peru, old (around 1900)	ZMHB
NVG-17068C03			USA: MN, Pipestone Co., 1986, CSU_ ENT1025108	CSUC
NVG-14112G12	Ocella albata		Peru, no date	TLS
PAO-23	Ochlodes agricola		USA: CA, Sierra Co., 2016	UTSW
NVG-18039E08	Ochlodes subhyalina		Korea, 1956	FMNH
PAO-263	Ochlodes sylvanoides		USA: CO, Larimer Co., 2016	UTSW
NVG-7746			Japan, 1933	USNM
NVG-17114B06			USA: CO, Mesa Co., 2001, LepNet 1024498	CSUC
NVG-5686			Brazil: Rio de Janeiro, 1996, 14063A12	USNM
NVG-18014F09			Peru, 2016	USNM
NVG-18021D06	-		Mexico: San Luis Potosi, 1967	AMNH
NVG-14063C11	Oileides fenestratus		French Guiana, 1993, Hermier No 4771	USNM
NVG-18067C02	Oileides fenestratus		French Guiana, 2009	EBrockmann
NVG-18098F04	Oileides guyanensis		French Guiana, 2002, H19994	BHermier
NVG-18057C09			Brazil: Espirito Santo, old (around 1900)	ZSMC
NVG-5727	Oileides vulpinus		Brazil: Espirito Santo, 1969	USNM
NVG-15117B01	_		Mexico: Tamaulipas, 2003, CSU_ENT1039476	CSUC
NVG-17092D06			Mexico: Oaxaca, 1990	USNM
NVG-8040	Onophas columbaria		Ecuador, 2002	USNM
NVG-17109G08	Orphe gerasa		Venezuela, 1993	LACM
NVG-2761	Orphe vatinius		Colombia, 1976	JAScott
NVG-7956			Costa Rica, 2008, 08-SRNP-40358	USNM
NVG-18111A01	-		Brazil: Rio de Janeiro, 1995	USNM
NVG-8061	Orthos orthos		Brazil: Parana, 1991	USNM
NVG-18026A06	Orthos orthos hyalinus	нт	Brazil: Santa Catarina, old (around 1900)	AMNH
NVG-17119C02	Osmodes laronia		Uganda, 1961	USNM
NVG-18073E04	Osphantes ogowena	ST	Gabon, 1888	ZMHB
NVG-18011G05	Ouleus fridericus		Guyana, 2000	USNM
NVG-7894	Ouleus salvinaDHJ01		Costa Rica, 2011, 11-SRNP-57460	USNM
NVG-14113A02	Oxynetra aureopecta	нт	Mexico: Hidalgo, 1987	LACM
7962	Oxynthes coruscaDHJ02		Costa Rica, 2011, 11-SRNP-23410	USNM
NVG-7899	Paches loxus		Costa Rica, 2003, 03-SRNP-30995	USNM
NVG-18025B01	Pachyneuria lineatopunctata (=phintias)	нт	Peru, 1931	AMNH
NVG-15033D07	Pachyneuria obscura	Т	Peru, old (prior to 1888)	ZMHB

DNA voucher	Taxon name	Type	Brief data	Collection
NVG-4155	Panoquina panoquin		USA: TX, Jefferson Co., 2015	UTSW
NVG-17111G07			Mexico: Hidalgo, 1981	LACM
NVG-8037	1		Peru, 2013	USNM
NVG-18063A09	Parachoranthus magdalia		Cuba, 2009	EBrockmann
NVG-17092H10	Paracleros biguttulus		Uganda, 1956	USNM
NVG-18027E08	Paramimus scurra scurra		Guyana, 2001	USNM
NVG-18015E01	Paratrytone rhexenor		Mexico: Veracruz, old (around 1900)	USNM
NVG-17108G05	Pardaleodes edipus		Zaire, 1981	LACM
NVG-18026E08	-	нт	Paraguay	AMNH
NVG-7290	-		Myanmar, 2002	USNM
NVG-17108G12			Sierra Lione, 1974	LACM
NVG-18013A05	Parphorus storax		Guyana, 2000	USNM
NVG-17094H03	-		Costa Rica, 2013, 13-SRNP-6248	USNM
NVG-18082E01			Papua NG, 1982, NHMUK_010430882, 0247277187	BMNH
NVG-17116A09	Pellicia dimidiata dimidiata		Mexico: Tamaulipas, 1974	TAMU
NVG-18013C07			Ecuador, 2001	USNM
NVG-18011F04			Brazil: Santa Catarina, 1990	USNM
NVG-7955			Costa Rica, 2013, 13-SRNP-65013	USNM
NVG-18065F12			Peru, 2015	EBrockmann
	Perichares deceptus fulvimargo		Peru, 2013	USNM
NVG-18014F08	1 , 0		Ecuador, 1976	USNM
NVG-7986	,		Brazil: Rondonia, 1992	USNM
NVG-18105F10			Trinidad, 2000	USNM
NVG-18027A09		нт	Brazil: Rio de Janeiro	AMNH
	Perichares philetes	1	Jamaica, 2017	UTSW
	Perus coecatus (=tadus)	ST	Brazil: Rio de Janeiro	USNM
NVG-7826	` ′		Peru, 1999	USNM
NVG-19055F06		ST	Bolivia, 1899	MCZ
NVG-18059C09			Ecuador, 1988	USNM
NVG-17092F05			Costa Rica, 2015, 15-SRNP-71060	USNM
	Pheraeus odilia epidius		Panama, 1982	USNM
	Phlebodes pertinax		Brazil: Amazonas, 1993	USNM
	Phocides pigmalion okeechobee		USA: FL, Monroe Co., 2015	UTSW
	Pholisora catullus		USA: TX, Starr Co., 2015	UTSW
NVG-7976	Pholisora mejicanus		USA: NM, Colfax Co., 1989	USNM
NVG-17068A11	Piruna aea mexicana		USA: AZ, Santa Cruz Co., 2016, CSU_ ENT1033276	CSUC
NVG-6454	Piruna pirus	1	USA: CO, Grand Co., 2016	UTSW
NVG-17108G10	-	1	South Africa, 1944	LACM
NVG-7982	· ·		Brazil: Rondonia, 1989	USNM
NVG-939		1	USA: LA, 2011	UTSW
NVG-4704			USA: FL, Levy Co., 2015	UTSW
NVG-17114B07	Poanes massasoit		USA: MD, Dorchester Co., 1976, LepNet 1031028	CSUC
NVG-6711	Poanes viator zizaniae		USA: TX, Dallas Co., 2016	UTSW
NVG-7046	Poanes yehl		USA: TX, Hopkins Co., 2016	UTSW
NVG-8170	-	1	USA: FL, Miami-Dade Co., 2017	UTSW
NVG-5968			USA: AZ, Santa Cruz Co., 2016	UTSW
NVG-4276		1	USA: IN, Montgomery Co., 2015	UTSW
NVG-4255		1	USA: IN, Montgomery Co., 2015	UTSW
NVG-7875	Polyctor polyctor		Costa Rica, 2012, 12-SRNP-4870	USNM
NVG-18115C01	Pompeius amblyspila		Brazil: Amazonas, 1993	USNM

DNA voucher	Taxon name	Type	Brief data	Collection
NVG-17106B02	Pompeius pompeius		Costa Rica, 2010, 10-SRNP-103576	USNM
NVG-18038D12	Potamanaxas flavofasciata flavo- fasciata		Ecuador, 2008	EBrockmann
NVG-18082C08	Prada rothschildi	PT	Papua NG, 1899, NHMUK_012824115, 0247281590	BMNH
NVG-18082C09	Prada rothschildi	PT	Papua NG, 1899, NHMUK_010430826, 0247278442	BMNH
NVG-18114H04	Propapias sipariana		French Guiana, 1993	USNM
NVG-7331	Pseudocoladenia dan fabia		Myanmar, 2001	USNM
NVG-8050	Pseudocopaeodes eunus		USA: CA, Inyo Co., 1950	USNM
11-BOA- 13382A03	Pseudodrephalys hypargus		Ecuador	USNM
NVG-18113B05	Pseudorphe pyrex		Peru, 1992	USNM
NVG-17112C01	Pseudosarbia phoenicicola		Uruguay, 1967	LACM
NVG-18021B02	Psoralis idee		Bolivia, old (around 1900)	AMNH
NVG-7757	Pteroteinon laufella		Liberia, 1988	USNM
NVG-7771	Pyrgus malvae		Greece, 1992	USNM
NVG-8060	Pyrrhocalles antiqua		Dominican Republic, 1994	USNM
NVG-17094C09	Pyrrhopyge hadassa pseudoha- dassa		Peru, 2013, USNMENT 00894886	USNM
NVG-7984	Pyrrhopygopsis socrates		Brazil: Mato Grosso, 1991	USNM
NVG-7896			Costa Rica, 2006, 06-SRNP-7674	USNM
11-BOA- 13382B01	Pythonides jovianus jovianus		Guyana, 2000	USNM
NVG-14102D02	Pythonides lerina		Guyana, 2003	FMNH
NVG-7902	Quadrus cerialis		Costa Rica, 2008, 08-SRNP-1186	USNM
NVG-18013H08	Quadrus lugubris		Costa Rica, 2008, 08-SRNP-57856	USNM
NVG-18018A03	Quadrus truncata		Ecuador, 1988	USNM
NVG-18015B10	Quasimellana mexicana		Mexico: Oaxaca, 1989	USNM
NVG-7959	Quinta cannae		Costa Rica, 2012, 12-SRNP-75508	USNM
NVG-18057H02	Rachelia extrusus		Papua New Guinea, old (around 1900)	ZSMC
NVG-18012A12	Racta plasma		Peru, 2011	USNM
NVG-17092G01	Radiatus bradus		Guyana, 1999	USNM
NVG-18091D12	Ralis concolor		Ecuador, 2012	EBrockmann
NVG-19021E08	Ralis coyana		Brazil: Rio de Janeiro, 1996	USNM
NVG-17092C08	Remella remus		Costa Rica, 2006, 06-SRNP-6640	USNM
NVG-18081A02	Repens repens	PT	Paraguay, 1904, NHMUK_010430831, 0247279233	BMNH
NVG-17069G04	Rhabdomantis galatia		Uganda, 1958	USNM
NVG-17116C08	Rhinthon osca		USA: TX, Hidalgo Co., 1974	TAMU
NVG-18093C02	Rigga auristriga	НТ	Bolivia, old (around 1900)	SMF
NVG-19019G04	Rigga hesia		Ecuador, 2002	USNM
NVG-18093C05	Sacrator sacrator (=stupenda)	НТ	Colombia, old (around 1900)	SMF
NVG-18012E03	Saliana salius		Guyana, 2000	USNM
NVG-8024	Saniba sabina		Brazil: Rio de Janeiro, 1996	USNM
NVG-18025B08	Santa santes	НТ	Peru, 1924	AMNH
11-BOA- 13382F10	Santa trifasciatus		Guyana	USNM
NVG-7345	Sarangesa dasahara		Myanmar, 2001	USNM
NVG-18037G08	Satarupa nymphalis		China: Sichuan, 2010	UTSW
NVG-18013C01	Saturnus saturnus		Guyana, 2000	USNM
NVG-7803	Semalea pulvina		Cameroon, 1989	USNM
NVG-7760	Signeta flammeata		Australia, 1985	USNM
NVG-7970	Sodalia sodalis		Costa Rica, 2011, 11-SRNP-75366	USNM

DNA voucher	Taxon name	Type	Brief data	Collection
NVG-14107C10	Sophista aristoteles aristoteles		Peru, 2013	USNM
NVG-18011H10	Sostrata bifasciata		Brazil: Rio de Janeiro, 1995	USNM
NVG-7247	Sostrata nordica		Mexico: Veracruz, 1906	USNM
NVG-18032A04	Sostrata pusilla pulsa		Ecuador, 1976	USNM
NVG-18032A10	Sostrata pusilla pusilla		Peru, 1989	USNM
NVG-17108E02	Spialia galba galba		India, 1961	LACM
NVG-18013H11	Spioniades artemides		Costa Rica, 2010, 10-SRNP-67995	USNM
NVG-18071A04	Staphylus ascalaphus		Costa Rica, 2016, 16-SRNP-56091	USNM
NVG-18071A05	Staphylus caribbea		Costa Rica, 2009, 09-SRNP-42728	USNM
NVG-9694	Staphylus ceos		USA: AZ, Santa Cruz Co., 2017	UTSW
NVG-18025B10	Staphylus musculus (=similis)	нт	Brazil: Sta Catarina, old (around 1900)	AMNH
NVG-18025B03	Staphylus putumayo	HT	Peru, 1931	AMNH
NVG-18011H08	Staphylus vincula		Mexico: Oaxaca, 1988	USNM
NVG-18071A12	Staphylus vulgata		Costa Rica, 2010, 10-SRNP-55088	USNM
NVG-6016	Stinga morrisoni		USA: TX, Jeff Davis Co., 2016	UTSW
NVG-8044			Guyana, 2000	USNM
NVG-18012D04			Argentina, 1998	USNM
NVG-7937	-0		Costa Rica, 2007, 07-SRNP-21744	USNM
NVG-3621	Systasea pulverulenta		USA: TX, Duval Co., 2015	UTSW
NVG-6008	-			
NVG-18011E02	1		USA: TX, El Paso Co., 2016	UTSW
			Mexico: Oaxaca, 1992	USNM
NVG-7333	Tagiades litigiosus litigiosus		Myanmar, 2001	USNM
NVG-18012D01	Talides sinois		Peru, 2015	USNM
NVG-7375	Taractrocera maevius sagara		Myanmar, 2003	USNM
NVG-18116E01	Tava tavola	ST	Trinidad, old (around 1900)	USNM
NVG-18026A03	Tava tavola (=hoffmanni)	HT	Brazil: Santa Catarina, 1913	AMNH
NVG-5726	Telemiades fides		Costa Rica, 2011, 11-SRNP-20768	USNM
NVG-8008	Tellona variegata		Brazil: Rondonia, 1993	USNM
NVG-17119H08	Teniorhinus watsoni watsoni		Uganda, 1961	USNM
NVG-18026F01	Testia potesta	HT	Peru, 1931	AMNH
NVG-18098F08	Thargella caura		French Guiana, 2005, H22106	BHermier
11-BOA- 13386C12	Theagenes aegides		Costa Rica, 1980	USNM
11-BOA- 13386C11	Theagenes albiplaga		Peru, 2008	USNM
NVG-18013B04	Thoon modius		Ecuador, 1992	USNM
NVG-7383	Thoressa masoni		Myanmar, 2001	USNM
NVG-17112B06	Thracides phidon		Venezuela, 1993	LACM
NVG-18022H12	Thymelicus acteon acteon		Spain, 1953	AMNH
NVG-7888	Tiana niger		Costa Rica, 2011, 11-SRNP-35371	USNM
NVG-7944	Tigasis arita		Costa Rica, 2011, 11-SRNP-32281	USNM
NVG-18013A11	Tigasis zalates		Ecuador, 1977	USNM
NVG-7883	Timochares trifasciata		Costa Rica, 2005, 05-SRNP-12097	USNM
NVG-7908	Timochreon satyrus		Costa Rica, 2007, 07-SRNP-58884	USNM
NVG-18118A08	Tirynthia conflua		Brazil: Rio de Janeiro, 1995	USNM
NVG-18081D03	Toxidia thyrrhus		Australia, 1911, NHMUK_010430805, 0247281664	BMNH
NVG-16106A10	Trapezites symmomus		Australia, 1963	LACM
NVG-19023C11	Tricrista advena		Guyana, 2000	USNM
8049	Tricrista crista		Guyana, 2000	USNM
NVG-18025G05	Tricrista cristatus	НТ	Brazil: Santa Catarina, old (around 1900)	AMNH
NVG-7983	Trina geometrina	1	French Guiana, 1993	USNM
NVG-18012D02	Tromba tromba	1	Peru, 2014	USNM
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DNA voucher	Taxon name	Type	Brief data	Collection
NVG-18019E10	Tsitana tsita		South Africa, 1924	AMNH
NVG-18111G06	Turesis complanula		Guyana, 2000	USNM
NVG-18082D06	Turmosa camposa		Brazil: RJ, 1883, NHMUK_012824124, 0247279797	BMNH
NVG-18056F01	Turmosa camposa	LT	no data	ZSMC
NVG-5699	Typhedanus ampyx		Mexico: Oaxaca, 1992, 14104H07	USNM
NVG-17108H12	Unkana ambasa		no data, no date	LACM
NVG-18081C02	Unkana mytheca		Indonesia, 1914, NHMUK_010430823, 0247278996	BMNH
NVG-4894	Urbanus proteus proteus		USA: FL, Miami-Dade Co., 2015	UTSW
NVG-17095C06	Vacerra litana		Venezuela, 1975	USNM
NVG-17098F07	Veadda veadeira		Brazil: Mato Grosso, 1991	USNM
NVG-17111G04	Vehilius stictomenes illudens		Mexico: San Luis Potosi, 1980	LACM
NVG-8022	Venas evans		Guyana, 2000	USNM
NVG-18021D01	Vernia dares		Mexico: San Luis Potosi, 1966	AMNH
NVG-18014H01	Vernia verna		USA: OH, Summit Co., 2012	USNM
NVG-18013B11	Vertica verticalis		Peru, 1983	USNM
NVG-18014G01	Vettius phyllus phyllus		Guyana, 2001	USNM
NVG-18071C05			Costa Rica, 2015, 15-SRNP-71571	USNM
NVG-18012H07	-		Paraguay, old (around 1900)	USNM
NVG-15097D10	Vinpeius tinga (=freemani)	нт	Mexico: Veracruz	CMNH
NVG-18011G11	1 0 ()	ST	Brazil: Rio de Janeiro, old (around 1900)	USNM
NVG-18061D04		1	Brazil: Rio de Janeiro, 1995	USNM
NVG-7973			Brazil: Mato Grosso, 1991	USNM
NVG-18012F11			Paraguay, 1986	USNM
NVG-18026H05	0 0	нт	Ecuador, 1938	AMNH
NVG-19055F04		HT	Bolivia, 1899	MCZ
NVG-15104C11		нт	Ecuador, 1938	AMNH
NVG-18028C11			Costa Rica, 2003, 03-SRNP-27671	USNM
NVG-7972			French Guiana, 1988	USNM
NVG-17107B06	Wahydra kenava		Venezuela, 1978	USNM
NVG-7421			Dominican Republic, 1981	USNM
NVG-17098E07	, ,		French Antilles, 1989	USNM
NVG-17098E12			Brazil: Mato Grosso, 1990	USNM
NVG-10332	0 1		Jamaica, 2017	UTSW
NVG-18079C02	- v	Т	Ethiopia, 1925, EL63085	MNHP
NVG-18063B11			Tanzania, old (around 1900)	ZSMC
NVG-17069E09			Zimbabwe, 1961	USNM
NVG-15102D03			Mexico: Sonora, 1984	USNM
NVG-7765		1	Cameroon, 1987	USNM
NVG-17091D05			Malaysia, old (around 1900)	USNM
NVG-18064D07			Cameroon, old (around 1900)	USNM
NVG-17112A11	V	1	Brazil: Santa Catarina, 1999	LACM
	Xenophanes tryxus	1	Costa Rica, 2010, 10-SRNP-103428	USNM
NVG-15033D08		T	Brazil: Amazonas, prior to 1889	ZMHB
NVG-17105B08		 	Colombia, 1965	USNM
NVG-18074B01		T	Palawan, 1888	ZMHB
NVG-18098G05		 	Malaysia, 2016	UTSW
NVG-18126A12		1	Sabah, 1984	KMaruyama
NVG-18126A05			Langkawi, 2018	UTSW
NVG-17112A04	Zenis jebus hemizona	1	Venezuela, 1993	LACM
NVG-18038B10			Honduras	RGallardo
, G 10000B10				1 Samurao

DNA voucher	Taxon name	Туре	Brief data	Collection
NVG-18091C05	Zera phila		Ecuador, 2012	EBrockmann
NVG-18011E12	Zera zera		Peru, 2014	USNM
NVG-18027A07	Zetka zeteki	HT	Panama, 1928	AMNH
NVG-15104C01	Zobera albopunctata	HT	Mexico: Colima	AMNH
NVG-17091C04	Zographetus satwa		no data	USNM
NVG-17069H08	Zophopetes dysmephila		South Africa, 1953	USNM
NVG-6998	Zopyrion sandace		Mexico: Oaxaca, 1981	TAMU
NVG-1670	Pterourus glaucus glaucus		USA: TX, Denton Co., 2013	USNM