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New species, a new combination, and a new country record in American Clytini (Coleoptera: Cerambycidae: Cerambycinae)

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New species, a new combination, and a new country record in American Clytini (Coleoptera: Cerambycidae: Cerambycinae)

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Abstract. Two new species of Mexican Clytini (Coleoptera: Cerambycidae: Cerambycinae) are described: Trichoxys giesberti Botero, Santos-Silva and Wappes (also added to a recent key) and Megacyllene giesberti Botero, Santos-Silva and Wappes. The geographical distribution of Megacyllene melanaspis (Chevrolat, 1862) is expanded to include Bolivia, new country record, and compared to the similar and sympatric Megacyllene proxima (Laporte and Gory, 1841); Megacyllene asteca (Chevrolat, 1860) is proposed as a new combination for the previous Plagionotus asteca, and Amyipunga armaticollis (Zajciw, 1964) is redescribed to correct previous errors regarding it in the literature. Additionally, characters to help separate it from the similar species Amyipunga moritzii (Thomson, 1861) are provided.

Key words. Amyipunga, Mexico, taxonomy, Trichoxys.

Introduction

Two undescribed clytine (Coleoptera: Cerambycidae: Cerambycinae) species found in the Edmund Giesbert collection at the Florida State Collection of Arthropods (FSCA) are described herein, Trichoxys giesberti Botero, Santos-Silva and Wappes and Megacyllene giesberti Botero, Santos-Silva and Wappes. The Giesbert-labelled Trichoxys Chevrolat, 1860 specimen had been collected by him in Mexico and he recognized it as new as evidenced by his handwritten, paratype, manuscript name label (T. incomptus Giesbert) on its pin. Recently, the Giesbert-proposed holotype of the species (also with a handwritten label by him) was located at the EMEC and is included in the type series but not as the holotype. A recent key to species of *Trichoxys* by Lingafelter and Wappes (2012) is modified to add the new species. Another Megacyllene, M. melanaspis (Chevrolat, 1862) is newly recorded for Bolivia, new country record, and is found to be sympatric with the widely distributed and very similar Megacyllene proxima (Laporte and Gory, 1841). Diagnostics are given to separate these two species, and a known distribution map within Bolivia provided. Another Bolivian Clytine, Amyipunga armaticollis (Zajciw, 1964), typically found at high altitudes in or near Amboro National Park, is redescribed to correct errors in its original and subsequent descriptions, record intraspecific variation in its dorsal appearance and define characters useful for separating it from the similar Venezuelan species, Amyipunga moritzii (Thomson, 1861). Lastly, *Plagionotus asteca* Chevrolat, 1860, based on the shape of its mesoventral process, is transferred and becomes *Megacyllene asteca* **new combination**.

Materials and Methods

Photographs were taken in Museu de Zoologia, Universidade de São Paulo with a Canon EOS Rebel T3i DSLR camera, Canon MP-E 65mm f/2.8 1-5X macro lens, controlled by Zerene Stacker AutoMontage software. Measurements were taken in "mm" using a measuring ocular Hensoldt/Wetzlar - Mess 10 in the Leica MZ6 stereomicroscope, also used in the study of the specimens. All specimens examined were adults; immature stages of the new species have not yet been collected.

The acronyms used in the text are as follows:

ACMT American Coleoptera Museum (James Wappes), San Antonio, TX, USA

CBFL Colección Boliviana de Fauna. LaPaz, Bolivia

EMEC Essig Museum of Entomology, Berkeley, CA, USA

FSCA Florida State Collection of Arthropods, Gainesville, FL, USA

FWSC Fred W. Skillman collection, Pearce, AZ, USA

MNRJ Museu Nacional, Universidade Federal do Rio de Janeiro, Rio de Janeiro, Rio de Janeiro, Brazil

MZSP Museu de Zoologia, Universidade de São Paulo, São Paulo, Brazil

RFMC Roy F. Morris Collection, Lakeland, FL, USA

SLPC Steven Lingafelter personal collection, Hereford, AZ, USA

TAMU Texas A and M University Insect Collection, College Station, TX, USA

Taxonomy

Trichoxys Chevrolat, 1860

Trichoxys Chevrolat, 1860: 454; Thomson, 1861: 219; Lacordaire 1869: 63; Lingafelter and Wappes 2012: 155.

Trichoxys was described by Chevrolat (1860) as a subgenus of Clytus Laicharting, 1784. He included 14 species in the group but did not designate a type species. Later, Thomson (1861) elevated Trichoxys to genus level and designated Clytus (Trichoxys) bilineatus Chevrolat, 1860 as type species. Trichoxys is currently composed of 15 species, all of them recorded for Mexico with five of the species extending their known distribution into Central America (Tavakilian and Chevillotte 2018; Bezark 2019). Recently, Lingafelter and Wappes (2012) provided a diagnosis for the genus as follows: "Trichoxys Chevrolat is characterized from other Clytini by 1) the abruptly recessed parasutural region of the elytron that starts at the anterior one-third and diverges more widely to the apex where it is approximately one-third of the discal width, and the strongly elevated anterior portion of the mesosternum (appearing abruptly recessed from ventral view) and 2) the evenly rounded pronotum that is distinctly narrower than the elytral base and without calli or lateral asymmetry. All the known species have bold, pubescent patterns on the elytra which are easily diagnostic for each species." They also placed Trichoxys ochraetheoides Linsley, 1935 in synonymy with T. hirtellus (Chevrolat, 1860), described T. penrosei as a new species, and provided a key to the species based on their illustrations of elytral patterns.

$\it Trichoxys~giesberti~Botero, Santos-Silva~and~Wappes, new species~(Fig.~1–4)$

Description of female. Integument mostly dark brown, almost black in some areas, especially head; palpi dark reddish-brown; labrum mostly dark reddish-brown; antennae and legs reddish-brown (slightly darker in some areas of femora); ventrites dark reddish-brown, except basal 2/3 of ventrite I dark brown.

Head. Frons finely, sparsely punctate; with dense yellowish-brown pubescence laterally (more yellowish depending on light intensity), gradually yellowish-white, slightly less dense toward central area, except median groove glabrous, as is triangular central area close to postclypeus; with a few long, erect yellowish setae laterally. Area between antennal tubercles with yellowish-white pubescence not obscuring integument toward frons, yellowish-brown (more yellowish depending on light intensity), dense toward

upper eye lobes; remaining surface of vertex, and area behind upper eye lobes with dense yellowishbrown pubescence (more yellowish depending on light intensity) obscuring integument; with sparse, erect, yellowish-brown setae interspersed behind upper eye lobes; surface finely, moderately abundantly punctate. Area behind lower eye lobes moderately finely, sparsely punctate close to eye, smooth in remaining surface; with dense yellowish-brown pubescence close to eye (more yellowish depending on light intensity), gradually slightly lighter toward ventral surface, not obscuring punctures, with long, erect yellowish setae interspersed, glabrous on remaining surface. Genae finely, moderately abundantly punctate except smooth distal area; with light yellowish-brown pubescence, gradually sparser toward glabrous apex, with long, erect yellowish setae interspersed. Antennal tubercles glabrous, basally finely, sparsely punctate, smooth, on distal half. Median groove distinct from clypeus to area between upper eye lobes. Postclypeus with abundant yellowish-brown pubescence (more yellowish depending on light intensity) in wide central area, glabrous laterally. Labrum coplanar with anteclypeus at posterior half, inclined, nearly vertical, anterior half concave; with short, sparse yellowish setae with long, erect yellowish-brown setae interspersed in posterior half, longer laterally, and moderately short, erect yellowish setae in anterior half. Gulamentum smooth, glabrous in posterior half; anterior half gradually sloped toward distinctly elevated anterior margin, finely punctate (punctures gradually more abundant toward anterior margin), with moderately short, decumbent yellowish setae with long, erect brownish setae interspersed in anterior third. Distance between upper eye lobes 0.75 times length of scape; in frontal view, distance between lower eye lobes 0.88 times length of scape. Antennae 1.17 times elytral length, slightly surpassing base of posterior quarter of elytra; segments with yellowish-white pubescence not obscuring integument, gradually shorter toward distal segment; pedicel and antennomeres III-VI with long, erect reddish-brown setae ventrally, gradually sparser toward VI; antennomeres VII-IX with a few long, erect reddish-brown setae at ventral apex, gradually shorter toward IX; antennomeres III-X with long, erect reddish-brown setae dorsally, gradually shorter toward X. Antennal formula (ratio) based on length of antennomere III: scape = 1.43; pedicel = 0.43; IV = 1.17; V = 1.40; VI = 1.40; VII = 1.38; VIII = 1.33; IX = 1.19; X = 1.07; XI = 1.33.

Thorax. Prothorax slightly wider than long; sides uniformly rounded from anterolateral angles to basal constriction. Pronotum moderately finely, densely punctate; with transverse sulcus centrally in posterior constriction; with light yellowish-brown pubescence (yellower depending on light intensity) partially obscuring integument, slightly denser laterally, with moderately long, suberect brownish setae interspersed, and a few long, erect, pale yellow setae in sides of posterior area. Sides of prothorax with sculpturing and pubescence as in sides of pronotum, pubescence slightly sparser toward prosternum. Prosternum moderately finely, abundantly punctate, with dense yellowish pubescence, except nearly smooth, glabrous longitudinal band laterally, and smooth, nearly entirely glabrous area close to procoxal cavities; with sparse, long, erect yellowish setae. Mesoventrite with sparse yellowish-gray pubescence gradually denser toward sides; mesoventral process, mesanepisternum, and mesepimeron with dense yellowish pubescence. Metanepisternum and most of metaventrite with dense yellowish pubescence, sparser toward glabrous central area of metaventrite, and glabrous in narrow anterolateral area of metaventrite; sides of metaventrite with sparse, long, erect yellowish setae. Scutellum with dense light yellowish-brown pubescence.

Elytra. Very finely, abundantly punctate except smooth carina; dorsal carina well-marked from slightly before middle to near apex; apex wide, obliquely truncate, with minute projection at outer angle; with dense yellowish-brown pubescence obscuring integument. Legs. Femora with dense yellowish-white pubescence (slightly yellower in some areas depending on light intensity), except inner surface of meta-femora partially glabrous. Tibiae with bristly yellowish pubescence not obscuring integument (more whitish depending on light intensity), denser, brownish in distal half of ventral surface of protibiae, moderately denser, yellowish-brown in distal area of ventral surface of meso- and metatibiae. Metatar-somere I 1.65 times as long as II–III together.

Abdomen. Ventrites I–IV with dense yellowish pubescence laterally (except sparse pubescence in basal area of III–IV), distinctly sparser centrally (more yellowish white toward glabrous posterior area) ventrite V with sparse yellowish-white pubescence except nearly glabrous basal area, with long, erect brownish setae interspersed; apex of ventrite V truncate, slightly emarginate centrally.

Dimensions (mm). Total length, 15.35; prothoracic length, 3.10; anterior prothoracic width, 3.25; posterior prothoracic width, 2.75; maximum prothoracic width, 3.80; humeral width, 4.30; elytral length, 10.05.

Type material. Holotype female from MEXICO, *Sinaloa*: 30 km W El Palmito, 2-9.X.1976, E. Giesbert col. (FSCA). Paratype female from MEXICO, *Sinaloa*: 20 miles NE of Concordia, 2.IX.1957, J. C. Schaffner (TAMU).

Remarks. Trichoxys giesberti sp. nov. is similar to T. abbreviatus Bates, 1880, but differs as follows: antennae reddish, distinctly surpassing middle of the elytra; antennomeres elongate, cylindrical; elytral pubescence not forming bands. In T. abbreviatus, the antennae are dark and do not reach the middle of the elytra, distal antennomeres widened, and elytral pubescence forming distinct bands. The new species differs from all the other species in the genus by the uniform elytral pubescence, completely lacking any bands or spots (present in one form or another in all other species of the genus). Trichoxys giesberti can be included in the Lingafelter and Wappes (2012) key to species (modified) as follows:

Etymology. Named to honor Edmund Giesbert (now deceased) skilled collector of long-horned beetles (including collector of the holotype) who collected throughout the American tropics and was a prolific contributor to our knowledge of New World Cerambycidae through his taxonomic writings and art illustrations.

Megacyllene Casey, 1912

Newman (1840) described the new genus *Cyllene* for *Cyllene spinifera* Newman, 1840 (now *Megacyllene nebulosa* Laporte and Gory, 1836) (Monné 2018). Subsequently, Casey (1912) described *Megacyllene* for *Clytus antennatus* White, 1855. Hopping (1932) disagreed with the Casey concept and placed *Megacyllene* into synonymy with *Cyllene*. Fisher (1945) found *Cyllene* Newman, 1840 to be preoccupied by *Cyllene* Gray, 1834 (Mollusca) and designated *Megacyllene* as a replacement. The genus is restricted to America (New World) (most species are neotropical) and is currently composed of 54 species and two subspecies (Bezark 2019; Tavakilian and Chevillotte 2018). Recently, the South American species were revised by Martins and Galileo (2011). A new species from west-central Mexico (Nayarit) is described below.

Megacyllene giesberti Botero, Santos-Silva and Wappes, new species (Fig. 5–8)

Description of female. Integument mostly black; mouthparts, antennae, femora, and part of coxae reddish-brown; pro- and mesotibiae orangish-brown; metatibiae reddish-brown basally, gradually orangish-brown toward apex; tarsi orangish-brown. Yellowish-white pubescence whiter or yellower depending on light intensity; long, erect setae brownish or brownish-yellow depending on light intensity.

Head. Frons moderately finely, abundantly punctate; with dense yellowish-white pubescence obscuring integument, connecting with similar band surrounding eyes, with long, erect setae interspersed laterally. Area between antennal tubercles finely, abundantly punctate; glabrous in area closer to frons, except for sparse, short, erect brownish setae; remaining surface of vertex moderately finely, abundantly punctate; area between middle of antennal tubercles and anterior margin of upper eye lobes with transverse, wide yellowish-white pubescent band obscuring integument, connected with band surrounding eyes, with long, erect setae interspersed; area between upper eye lobes with very short, inconspicuous brownish pubescence with long, erect setae interspersed; area closer to prothoracic margin with dense yellowish-white pubescent band obscuring integument, gradually wider toward center, nearly connected to similar band surrounding lower eye lobes. Area behind upper eye lobes moderately finely, abundantly punctate;

with very short, slightly conspicuous brownish pubescence with long, erect setae interspersed close to eye, with dense pubescent band close to prothoracic margin; area behind lower eye lobes with dense yellowish-white pubescent band close to eye, glabrous close to prothoracic margin; with sparse, long, erect setae in pubescent area. Genae, except glabrous apex, with dense yellowish-white pubescence obscuring integument. Antennal tubercles with sculpturing and setae as in area between them, with glabrous apex smooth, and posterior area with a few short, decumbent brownish setae. Median groove distinct from clypeus to prothoracic margin. Postclypeus with dense yellowish-white pubescence in wide central area close to frons, sparser in wide central area close to anteclypeus (bristly, yellower centrally), glabrous laterally; with long, erect setae on sides of wide central area. Labrum coplanar with anteclypeus at posterior 2/3, inclined at anterior third; with long, moderately abundant golden setae directed forward in coplanar area; anterior margin with fringe of short golden setae. Gulamentum smooth, glabrous in posterior half; slightly depressed, finely, abundantly punctate in anterior half, with moderately bristly, short setae not obscuring integument, with long, erect setae interspersed. Distance between upper eye lobes 0.76 times length of scape; in frontal view, distance between lower eye lobes 0.93 times length of scape. Antennae 0.9 times elytral length, slightly surpassing midlength of elytra. Scape finely, abundantly punctate except apex smooth; with short, decumbent, moderately abundant yellowish setae except in glabrous smooth area. Pedicel and antennomeres with short setae as on scape, gradually shorter, less conspicuous toward antennomere XI; pedicel and antennomeres III-VIII with long, erect setae ventrally and on apex, distinctly sparser toward VII; antennomeres I-X with sparse, short, erect brownish setae on apex. Antennal formula (ratio) based on length of antennomere III: scape = 1.27; pedicel = 0.37; IV = 0.97; V = 1.23; VI = 1.20; VII = 1.17; VIII = 0.90; IX = 0.80; X = 0.70; XI = 0.98.

Thorax. Prothorax distinctly narrower than humeral width of elytra. Pronotum finely, densely punctate; with three wide yellowish-white transverse bands, one along anterior and posterior margins, and one (the widest) centrally; area between bands with inconspicuous yellowish-brown pubescence, not obscuring integument; with sparse, long, erect setae laterally, and sparse, thick, nearly decumbent black setae centrally. Sides of prothorax with sculpturing as on pronotum; with three yellowish-white pubescent bands in continuation of those on pronotum, apical and distal ones connected with wide yellowish-white transverse pubescent band close to prosternum (central one just short of connecting with it). Prosternum slightly rugose-punctate; with moderately dense yellowish-white pubescence in posterior half, nearly obscuring integument, sparser in anterior half (sparser in area close to posterior half); with long, erect, sparse setae. Prosternal process with yellowish-white pubescence gradually sparser toward apex, with long, erect setae interspersed (longer and more abundant toward apex). Mesoventrite with moderately abundant yellowish-white pubescence not obscuring integument, finer, slightly denser laterally, except sides of anterior half nearly glabrous. Mesoventral process with anterior margin abruptly elevated; with moderately abundant yellowish-white pubescence laterally, sparser, slightly yellower centrally. Mesanepisternum with yellowish-brown pubescence not obscuring integument in area close to prothorax, yellowish-white, dense, obscuring integument in area close to mesepimeron; mesepimeron with yellowishbrown pubescence not obscuring integument. Metanepisternum with two large, dense yellowish-white pubescent maculae obscuring integument, one covering nearly all of basal half, another in posterior half, not reaching apex and margin close to metaventrite. Metaventrite with two wide vellowish-white transverse bands on each side, posterior ones denser; area along central discrimen glabrous; area between transverse bands with moderately sparse yellowish-white and yellowish-brown pubescence, near glabrous area; remaining surface with yellowish-brown pubescence distinctly not obscuring integument; with sparse, long, erect setae. Scutellum with dense yellowish-white pubescence obscuring integument.

Elytra. Very finely, densely punctate; apex obliquely truncate, with outer and sutural angles slightly, somewhat roundly projected. Pubescence as follows: dense yellowish-white pubescent band basally, reaching scutellum and nearly reaching epipleural margin, widened, projected backward in lateral inclined area; dense yellowish-white pubescent band in anterior seventh, arched, not reaching suture, nearly attaining middle of lateral inclined area; dense, yellowish-white semicircular pubescent spot along suture in anterior 2/7; dense yellowish-white pubescent band in about apex of anterior 2/7, slightly arched, not reaching suture, barely reaching lateral inclined area; dense yellowish-white pubescent spot in anterior 2/7 of inclined area; dense yellowish-white sinuous pubescent band near midlength, not reaching

epipleural margin, projected forward and backward along suture; dense yellowish-white oblique band just after midlength, reaching about middle of inclined and dorsal areas; short, dense arched yellowishwhite pubescent band close to suture just after the former band, almost reaching middle of dorsal area; dense distinctly curved backward yellowish-white transverse pubescent band in about apex of posterior 6/7, not reaching epipleural margin or suture; dense yellowish-white oblique pubescent macula near apex; remaining surface with slightly conspicuous yellowish-brown pubescence (appearing to be darker due to the integumental color). Surface with both, short and long, erect brownish setae distally. Legs. Femora and tibiae with short, moderately abundant yellowish-brown setae (more whitish depending on light intensity), distinctly not obscuring integument, bristlier on tibiae; metatarsomere I almost twice length of II–III together.

Abdomen. Last tergite densely covered in yellowish-white pubescence. Ventrites very finely, densely punctate laterally, finely, sparsely punctate centrally, except posterocentral area of ventrites I-IV; ventrites I-IV with large, dense yellowish-white pubescent macula on each side of posterior half, gradually smaller toward IV; ventrite I centrally with inverted V-shape of moderately abundant, short, decumbent, thick yellowish-white setae, sparser distally, except surrounding glabrous smooth areas with a few long, erect yellowish-brown setae interspersed; ventrite II with smaller central area of sparse, short, decumbent, thick yellowish-white setae somewhat bordered by glabrous smooth areas, and sparse, long, erect vellowish-brown setae throughout, longer and more abundant laterally; ventrite III and IV with very sparse, short, decumbent, thick yellowish-white setae centrally, except in glabrous smooth area, and long, erect, sparse yellowish-brown setae throughout, also lacking in glabrous smooth area, longer and more abundant laterally; sides of ventrites I-IV, between anterior margin and dense pubescent macula, with very short yellowish-brown setae not obscuring integument, and remaining surface of II-IV with sparse, decumbent yellowish-brown setae; sides of ventrite V with very short yellowish-brown setae not obscuring integument, and remaining surface with sparse, short, decumbent yellowish-brown setae, with long, erect yellowish-brown setae interspersed, centrobasal area glabrous; ventrite V with apex rounded.

Dimensions (mm). Total length, 21.10; prothoracic length, 3.70; anterior prothoracic width, 3.00; posterior prothoracic width, 3.50; maximum prothoracic width, 4.20; humeral width, 5.85; elytral length, 14.75.

Type material. Holotype female from MEXICO, Nayarit: 28 km NE Las Varas, 8.X.1976, E. Giesbert col. (FSCA).

Remarks. Megacyllene giesberti new species is unusual among its congeners in its slim appearance and evenly rounded pronotum (without basal constrictions) which is visibly much narrower than the elytra. Combined with the west-central distribution in Mexico (Nayarit) it should not be confused with any other species.

Megacyllene giesberti new species can be included in the alternative of couplet "30" from Martins and Galileo (2011). The alternative of couplet "30" encompasses a mistake, since M. neblinosa Di Iorio, 1995 was considered as having the second pubescent band transverse, but it is actually V-shaped. Accordingly, only M. multiguttata (Burmeister, 1865) and M. giesberti can be included in the alternative of couplet "30".

30(29). First elytral pubescent band does not extend laterally across base (not reaching anterior curvature); area close to apex not covered with dense yellowish-white pubescence. Bolivia, First elytral pubescent band does extend laterally across base (reaching anterior curvature); area close to apex covered with dense yellowish-white pubescence. Mexico (Nayarit).....

Etymology. Named to honor Edmund Giesbert (collector of the holotype) for his many published accomplishments in the taxonomy of American Cerambycidae.

Megacyllene melanaspis (Chevrolat, 1862)

(Fig. 22–26)

Cyllene menalaspis Chevrolat, 1862: 378; Aurivillius 1912: 385 (cat.); Blackwelder 1946: 581 (checklist).

Cyllene melanaspis; Lucas 1862: XCI; Lacordaire 1869: 62.

Clytus melanaspis; Gemminger 1872: 2932 (cat.).

Megacyllene melanaspis; Monné 1993: 8 (cat.); Martínez 2000: 86 (distr.); Monné et al. 2012: 10 (distr.); Monné and Chaboo 2015: 47 (distr.); Monné 2018: 131 (cat.).

Megacyllene (Megacyllene) melanaspis; Monné and Giesbert 1994: 112 (checklist); Monné 2005: 91 (cat.); Monné and Hovore 2006: 43 (checklist).

Megacyllene menalaspis; Martins and Galileo 2011: 72.

Chevrolat (1862) described *Cyllene menalaspis* in his work named "Description de Clytides de l'ancienne Colombie [Description of Clytides from ancient Colombia]." According to him (translated): "I continue this work [Chevrolat 1860], occupying myself for the moment only those who come from the old Colombia, forming today, as we know, three distinct states: New Granada, Venezuela and Ecuador." *Cyllene menalaspis* was described from New Granada, a country that has been reported to be Colombia (e.g. Monné 2018). However, at that time New Granada encompassed part of Panama as well as Colombia (see discussion detailing this in Santos-Silva and Botero (2018: 5)) accordingly, the correct type locality for *Cyllene menalaspis* is New Granada.

Lacordaire (1869) listed: "Cyll. melanaspis, elongata, crinicornis, caracasensis, Chevrol. Ann. d. l. Soc. entom. 1861, p. 378; Colombie." He changed Cyllene menalaspis to C. melanaspis without explanation. Furthermore, C. menalaspis was not described from Colombia, C. elongata was described from Venezuela on page 379, C. crinicornis was listed on page 380 as a new record for Venezuela, and C. caracasensis was described from Venezuela on page 380. This name change was repeated by Gemminger (1872) who also listed the species as C. melanaspis without explanation. Subsequently, the species was also referred to as C. menalaspis by Aurivillius (1912) and Blackwelder (1946). In Monné (1993) and henceforth the species has been listed as C. melanaspis, except for Martins and Galileo (2011), who listed the species as C. menalaspis. We believe that Chevrolat (1862) really intended to name the species "melanaspis", meaning dark shield, and C. menalaspis is simply a printer's error. On page XCVII of the same volume of the "Annales de la Société Entomologique de France" the name appears as "melanaspis". Thus, this can be considered the first correction of the spelling of a name (ICZN 1999: 32.5.1), and not in Lacordaire (1869). Accordingly, the author of the justified emendation is H. Lucas (1862) (see "Annales de la Société Entomologique de France", 1862, page XCI): "Mr. H. Lucas, assistant secretary, has, as in previous years, since 1860, to be in charge of drawing up this table as well as that of the Authors."

Based on the material available to us, *Megacyllene melanaspis* and *Megacyllene proxima* are sympatric throughout their known distribution in Bolivia. Although, *M. melanaspis* is encountered more commonly in higher elevations than *M. proxima* while the opposite is true in mid to lower elevations. Both have been collected on blossoms of woody plants and taken beating or crawling on fresh cut wood in agricultural cuts. The two species can be readily separated by their yellow pubescent pattern of the elytra. Both species have a similar transverse basal fascia with *M. melanaspis* then having three paired subcircular fascia down the suture (Fig. 22, 25) while *M. proxima* has four (Fig. 27–28).

Currently, *M. melanaspis* (Fig. 22–26) is known from Colombia, Venezuela, Ecuador, and Peru (Monné 2018). To this, Bolivia (Fig. 29) is added as a **new country record**.

Material examined. Megacyllene melanaspis: PERU, Junín: Pampa Hermosa Lodge (22 km N San Ramon, 1220 m, 10°59.3′S / 75°25.5′W), 1 female, 24–27.XI.2007, D. Brzoska col. (ACMT); Chanchamayo, 1 male, 9-12.VIII.1963, Caballero col. (MZSP). Ucayali: Pucallpa, 1 female, VI.1974, no collector indicated (MZSP). COLOMBIA, Cundinamarca: Viotá, 1 female, no date indicated, H. Apol. col. (MZSP). BOLIVIA (Fig. 29) (New country record), Santa Cruz: Road to Amboró above Achira, 3 males, 2 females, 10-11.X.2006, Wappes, Nearns, and Eya col. (ACMT); 1 male, 14–15.X.2006, Wappes, Nearns, and Eya col. (MZSP); above Achira (Rd to Floripondio; 1900 m; 18°09′S / 63°47′W), 1 female, 10.XII.2011, Wappes, Bonaso, and Morris col. (ACMT); 1 male, 1 female, 19.XII.2011, Wappes, Lingafelter and Woodley col. (ACMT); 2 males, 29.XI.2013, Konstantinov col. (SLPC); 1 male, 2 females, 29.XI.2013, Woodley col. (SLPC): 1 female, 19.XII.2011, Lingafelter col. (SLPC); 2 males, 4 females, 19.XII.2011, Woodley col. (SLPC); 4–5 km N Achira (rd to Floripondio; 18°09′S / 63°47′W, 6350′), 1 female, 15.IX.2012, Wappes, Skelley, Bonaso, and Hamel col. (ACMT); 1 female, 10.XII.2011, Morris and Wappes col. (RFMC); 4–6 km N Achira (5400–5800′), 1 female, 20.XI.2003, Wappes, Morris, and Nearns col. (ACMT); Achira (ridge rd to Amboró; 18°07′S / 63°48′W, 2000 m), 1 female, 24–25.I.2007, Wappes and Lingafelter col. (ACMT);

Achira area (N rd to Amboró, on Achira ridge; 18°19′S / 63°48′W), 1 female, 5–6.II.2013, Wappes, Bonaso, Lingafelter and Garzon col. (ACMT); Amboró (rd above Achira Campo, 5-5800′), 1 female, 9–11.X.2004, Morris and Wappes col. (RFMC); 1 male, 27–28.X.2011, Skillman and Wappes Col. (FWSC); Pvc. Florida, Floripondio (west), 18°08′S, 63°45′W, 1880 m, 1 male, 25.XI.2004, on/flying to fresh cut trees, Clarke col. (RCSZ); Florida prov. (16 km NE Mairana; 6600′; 18°05′S / 63°54′W), 1 female, 11.XII.2011, Wappes, Bonaso, and L. Sekerka col. (ACMT); above Achira Campo (road to Amboró, Vicoquin area, 18°07′S / 63°48′W), 1 female, 11.XI.2012, Bonaso and Windsor col. (ACMT); Refugio los Volcanes, 18°06′24″S, 63°35′55″W, 1056 m, 20.XII.2016, R. Perger col. (CBFL); 1 female, 8.XI.2017, Lingafelter col. (SLPC).

Megacyllene proxima: BOLIVIA (Fig. 29), Cochabamba, N. of Cristal Mayu, 19.X.2011, Skillman and Wappes col. (FWSC); 2 males, Wappes and Skillman col. (ACMT); Santa Cruz, Florida, 4 km N. Bermejo, Refugio Los Volcanes, 18°06'S, 63°36'W, 1000–1200 m, 7.XII.2013, Skillman and Wappes col. (FWSC); 10.XII.2015, Skillman and Wappes col. (FWSC); 12.XII.2011, Skillman and Wappes col. (FWSC); 1 male, 18-20.IX.2012, Wappes, Skelley, Bonaso and Hamel col. (ACMT); 1 male, 18-22.I.2006, Wappes and Lingafelter col. (ACMT); 1 male, 2 females, 3.XI.2017, Lingafelter col. (SLPC); 5 males, 1 female, 6-10. III.2011, Wappes and Thomas col. (ACMT); 1 female, 1350 m, 9-12.XII.2011, Morris and Wappes col. (RFMC); 1 male, 2 females, 1350 m, 18-24, X.2014, Morris and Wappes col. (RFMC); 1 female, Florida Prv., Vicoquin area above Achira, chaco, 1730 m, 22-25.I.2007, Wappes, Lingafelter and Prena col. (ACMT); 1 male, 1 female, 19.XII.2011, N. Woodley col. (SLPC); 2 males, 2 females, 22-25.I.2007, Lingafelter, Wappes and Prena col. (SLPC); 1 female, 19.XII.2011, Lingafelter col. (SLPC); Pvc. Florida, Floripondio (west), 18°08'S, 63°45'W, 1880 m, 1 male, 1 female, 1.XI.2009, on/flying to flowers of Sagüintillo, Clarke col. (RCSZ); 1 female, 1.XI.2009, flying to fresh cut trees, Clarke col. (RCSZ); 1 male, 25.XI.2004, Clarke col. (RCSZ); 18°08'S, 63°44'W, Floripondio (east), 1 female, on/flying to flowers of Sotillo, 16.XI.2009, Clarke col. (RCSZ); 1 female, 27.XI2009, Clarke col. (RCSZ); 1 female, 18°09'S, 63°47'W, 10.XII.2011, Morris and Wappes col. (RFMC); La Hoyado (above Agua Clara), 2 females, 19.XI.2003, Morris, Nearns and Wappes col. (RFMC). PERU, Madre de Dios, 1 female, 10-13.XI.2007, Amazonas Lodge (N-Atalaya), 480 m, 12°52.2′S, 71°22.6′W, D. Brzoska col. (ACMT).

Megacyllene asteca (Chevrolat, 1860), new combination (Fig. 17–18)

Clytus (Plagionotus) astecus Chevrolat, 1860: 489.

Clytus astecus; Gemminger 1872: 2915 (cat.).

Plagionotus astecus; Bates 1880: 53; 1885: 299 (distr.); Aurivillius 1912: 380 (cat.); Blackwelder 1946: 580 (checklist); Chemsak et al. 1992: 70 (checklist); Monné 1993: 15 (cat.); Chemsak and Noguera 1993: 62 (distr.); Monné and Giesber 1994: 117 (checklist); Noguera and Chemsak 1996: 401 (checklist); Toledo et al. 2002: 528 (distr.); Monné 2005: 112 (cat.); Monné and Hovore 2006: 46 (checklist); Monné 2018: 159 (cat.); Santos-Silva et al. 2018: 190 (distr.).

Mulsant (1839) erected *Platynotus* to include *P. detritus* (Linnaeus, 1758), and *P. arcuatus* (Linnaeus, 1758). He separated *Platynotus* from *Clytus* Laicharting, 1784 (which at that time included species currently placed in *Megacyllene* Casey, 1912) in a key: "Prothorax transverse oval; antennae setaceous, thick, subspinose externally at apex of the antennomeres", leading to *Platynotus*; "Prothorax subglobular or oblong, sometimes almost transverse oval, but then with antennae short and toothless", leading to *Clytus*. Then Mulsant (1842), finding that *Platynotus* was preoccupied, used *Plagionotus* as a replacement. Chevrolat (1860), considered *Plagionotus* as a subgenus of *Clytus*, but did not say why. Subsequently, *Plagionotus* was considered a distinct genus by some authors and subgenus of *Clytus* by others. Finally, following Villiers (1946) *Plagionotus* became widely accepted as valid and thus separate from *Clytus*.

Newman (1840) described *Cyllene* to accommodate *C. spinifera* Newman, 1840. However, comparing the original descriptions of *Platynotus* and *Cyllene*, it is impossible to separate them. Chevrolat (1860) also considered *Cyllene* as a subgenus of *Clytus* without any explanation. Lacordaire (1869) separated *Cyllene* from *Plagionotus* in his key: "Intercoxal abdominal process rounded at apex, especially in females", leading to *Cyllene*; "Intercoxal abdominal process in acute triangle in both sexes", leading to *Plagionotus*. This key is not diagnostic for the genera because the intercoxal process for species in the genera are not exactly as described and in both genera are more variable than stated. Fortunately,

Lacordaire (1869) did correctly report the shape of the mesoventral process for both genera: truncate anteriorly in *Cyllene*; inclined anteriorly in *Plagionotus*, a viable character even today and used to accurately separate the two genera.

The shape of the mesoventral process in 38 species of *Megacyllene* was examined, (including the type species of *Cyllene* and *Megacyllene*), as well as the same feature in *Plagionotus astecus*, and nine species of non-American *Plagionotus* (including the type species of this genus). Accordingly, because the mesoventral process (Fig. 18) is the same as the *Megacyllene* species examined and not the same as in the *Plagionotus* examined, *P. astecus* (Fig. 17) is formally transferred to *Megacyllene*, resulting in *Plagionotus* being excluded as a genus found in America (New World).

Material examined. MEXICO, *Guanajuato*: 2 females, no date indicated, U. R. Martins col. (MZSP). *Morelos*: Cuernavaca, 1 male, no date indicated, U. R. Martins col. (MZSP). *Guerrero*: 10 km N. Iguala, 3,800′-4,300′, 1 male, 2 females, 19–21.IX.1989, J. E. Wappes col. (ACMT). *Jalisco*: 15 km S. Autlan, 1 male, 29.IX.1991, J. E. Wappes col. (ACMT).

Redescription of *Amyipunga armaticollis* (Zajciw, 1964) (Fig. 9–16)

Neoclytus armaticollis Zajciw, 1964: 305; Monné 1993: 43 (cat.); Monné and Giesbert 1994: 113 (checklist); Di Iorio 1995: 335 (distr.); Julio et al. 2000: 19 (holotype); Di Iorio 2005: 46 (distr.); Monné 2005: 96 (cat.); Monné and Hovore 2006: 44 (checklist); Wappes et al. 2006: 19 (distr.).

Amyipunga armaticollis; Martins and Galileo 2011: 156; Monné and Monné 2016: 9 (holotype); Lingafelter et al. 2017: 26; Monné 2018: 108 (cat.).

Redescription. Male (Fig. 9–12). Integument mostly black; mouthparts reddish-brown; antennae dark brown, slightly darker on scape and basal segments; elytra dark brown basally, gradually lighter toward apex; tibiae blackish basally, gradually dark brown toward apex; tarsomeres I–III dark brown (III more reddish on metatarsi); tarsomere V reddish-brown; abdominal ventrites dark brown, slightly darker on abdominal ventrite I.

Head. Frons finely, abundantly punctate close to smooth area surrounding median groove, densely micropunctate in wide area close to eyes; with wide, dense, longitudinal white pubescent band on each side, from base of antennal tubercle to clypeus, with very sparse, short white setae on remaining surface; with long, erect, sparse white setae throughout, slightly more abundant on pubescent bands. Area between antennal tubercles finely, abundantly punctate; nearly glabrous toward frons, with moderately dense grayish-white (whiter depending on light intensity) bristly pubescence toward upper eye lobes, with long, erect grayish-white setae interspersed. Remaining surface of vertex finely, densely punctate, with shallow, coarser punctures interspersed; with yellowish-brown pubescence covering but not obscuring integument. Area behind eyes densely micropunctate with abundant fine punctures interspersed; area close to eye with yellowish-brown pubescence not obscuring integument behind upper eye lobe, gradually denser and white toward lower eye lobe; remaining surface with sparse yellowish-brown pubescence; with a few long, erect yellowish setae toward ventral surface of lower eye lobe. Genae with sculpturing as behind eyes except smooth distal area; with white pubescence not obscuring integument, except glabrous smooth area, and area close to center of inferior surface of lower eye lobe with slightly denser pubescence, and nearly glabrous area close to frons; with sparse, long, erect yellowish setae in pubescent area. Antennal tubercles finely, abundantly punctate basally toward frons, nearly entirely punctate toward upper eye lobes, smooth on remaining surface; glabrous toward frons and apex, with grayish-white pubescence toward upper eye lobes. Median groove distinct from clypeus to prothoracic margin (less so after upper eye lobes). Postclypeus finely, abundantly punctate in wide central area, smooth laterally; with white, dense bristly pubescence on each side of wide central area (continuing longitudinal bands on frons), with sparse, bristly white pubescence centrally, glabrous laterally; with sparse, long, erect white setae in wide central area. Labrum coplanar with anteclypeus at posterior 3/4, inclined at anterior quarter; with abundant, long, erect yellowish setae in coplanar area, abundant, short yellowish-brown setae in inclined area. Gulamentum smooth, glabrous posteriorly; wide anterior area somewhat depressed, except elevated anterior area, coarsely, abundantly rugose-punctate, with bristly white setae not obscuring integument, with long erect yellowish setae interspersed. Distance between upper eye lobes 1.25 times length of scape; in frontal view, distance between lower eye lobes 0.88 times length of scape. Antennae 0.85 times elytral length, reaching midlength of elytra; scape gradually widened toward apex, with maximum width slightly wider than twice basal width; scape, pedicel and antennomeres with yellowish-brown pubescence not obscuring integument, denser, shorter, less conspicuous toward distal segments; pedicel and antennomeres III–IV with long, erect, abundant brownish setae ventrally; antennal formula (ratio) based on length of antennomere III: scape = 1.08; pedicel = 0.30; IV = 0.85; V = 0.81; VI = 0.62; VII = 0.58; VIII = 0.40; IX = 0.38; X = 0.35; XI = 0.46.

Thorax. Prothorax wider than long (including lateral tubercles). Pronotum moderately coarsely, densely punctate; with dense white pubescent band on each side close to anterior and posterior margins, moderately abundant yellowish-brown pubescence posteriorly close to dense white band, moderately abundant white pubescence not obscuring integument between arched lateral carinae, except yellowish-brown pubescence in anterior third, and sparse yellowish-brown setae along central carina, and slightly dense white pubescence laterally. Sides of prothorax with sculpturing and pubescence as on sides of pronotum, with long, erect white setae interspersed. Prosternum moderately finely abundantly punctate at posterior 2/3, finely, transversely striate at anterior third; with moderately dense white pubescence laterally close to sides of prothorax, from posterior margin to near anterior margin, V-shaped, moderately distinct white pubescent band, from central apex of prosternal process to near sides of anterior margin, sparse white pubescence close to sides, procoxal cavities, and remaining surface of anterior third; with long, erect white setae throughout, more abundant posteriorly; posterior sides of prosternal process with sparse white pubescence with long, erect setae of same color interspersed. Mesoventrite finely, densely punctate (becoming slightly asperate); with distinctly sparse, decumbent white setae with long, erect setae of same color interspersed, slightly denser laterally and close to base of mesoventral process; mesoventral process with abundant yellowish-brown pubescence centrally, more white laterally, with long, erect setae (yellowish centrally, whitish laterally) interspersed; mesanepisternum with sculpturing as on mesoventrite, with dense white pubescent band close to mesepimeron, reaching and covering distal side of mesoventrite, and remaining surface with distinctly sparse whitish pubescence; mesepimeron with vellowish-brown pubescence not obscuring integument, with long, erect setae of same color interspersed. Metanepisternum with moderately abundant yellowish-brown pubescence not obscuring integument in anterior 2/3, and dense white pubescence in posterior third. Metaventrite mostly with abundant yellowish-brown pubescence not obscuring integument, except wide, inverted V-shaped white pubescent band centrally in anterior area, and moderately wide, transverse white pubescent band near metacoxal cavities (fused with that on metanepisternum), and glabrous narrow area centrally; with sparse, long erect setae (yellowish in yellowish-brown pubescent area, whitish in white pubescent areas). Scutellum with dense white pubescence.

Elytra. Moderately finely, densely punctate; apex widely obliquely truncate, with outer angle somewhat rounded (Fig. 16); pubescence dense, tawny, except: humeral area with yellowish-white pubescence; with arched white pubescent band in anterior third, almost reaching scutellum, laterally narrowed, arched backward, not reaching epipleural margin; arched white pubescent band starting about midlength, following along suture toward anterior third, laterally narrowed, arched backward, not reaching epipleural margin; another band in posterior third, similar to central band, but laterally less strongly curved backward; narrow pubescent band surrounding distal margin, and distal area of suture. Legs. Meso- and metafemora with club somewhat abruptly widened from peduncle; pro- and mesofemora with white pubescence not obscuring integument, with yellowish-brown pubescence interspersed, especially on ventral surface of peduncle, and long, erect white setae ventrally. Metafemora white pubescent with yellowish-brown pubescence interspersed on peduncle, ventral surface of club, central area of dorsal surface of club, and part of distal area of sides; remaining surface of sides with yellowish-brown pubescence not obscuring integument; entire surface with long, erect whitish setae ventrally; widest area of club about 4.5 times basal width of peduncle. Tibiae with white pubescence dorsally and laterally, not obscuring integument, with yellowish-brown pubescence interspersed; ventral surface with erect yellowish-brown setae, denser toward apex. Tarsi with yellowish-brown pubescence; metatarsomere I about 2.5 times II-III together.

Abdomen. Ventrites finely abundantly punctate (surface somewhat rugose); ventrites I–II with dense white pubescence except narrow anterior area with sparse yellowish-brown pubescence, with long, erect white setae interspersed throughout; ventrites III–IV with dense white pubescence on posterior third, sparse yellowish-brown pubescence on anterior 2/3, with long, erect white setae interspersed throughout; ventrite V with sparse yellowish-brown pubescence, slightly denser laterally and near apex, with long, erect yellowish-brown setae interspersed; abdominal ventrite V slightly narrowed toward apex; apex of ventrite V rounded.

Female (Fig. 13–14). Antennae 0.7 times elytral length, reaching about apex of anterior third of elytra; scape narrower (maximum width slightly less than twice basal width); meso- and metafemoral club gradually widened from peduncle; metafemoral club narrower (widest area of club narrower than 3.0 times basal width of peduncle); abdominal ventrite V distinctly narrowed toward apex.

Variation (males and females). Integument mostly dark brown; elytra nearly entirely dark reddishbrown; elytra dark reddish-brown with irregular dark brown or black areas interspersed; femora reddish-brown on peduncle, brown or dark reddish-brown on club; tibiae entirely brown; protibiae entirely light reddish-brown, and meso- and metatibiae yellowish-brown in basal third, gradually brown or dark reddish-brown toward apex; pro- and mesotarsi brown or dark reddish-brown, and metatarsi light reddish-brown; abdominal ventrites dark reddish-brown with distal area dark brown; abdominal ventrites entirely dark brown; longitudinal pubescent bands on frons distinctly yellow or light yellowishbrown; erect setae on frons yellowish; area between antennal tubercles with yellow or yellowish-brown pubescence toward upper eye lobes; pubescence behind and close to eye yellow to light yellowish-brown; pubescence on genae light yellowish-brown; pubescence on postclypeus light yellow to yellowish-brown; setae on labrum entirely nearly golden; setae on anterior area of gulamentum yellowish; bristly setae in anterior area of gulamentum distinctly sparse; antennal pubescence yellowish or somewhat golden; transverse pubescent bands on anterior and posterior sides of pronotum yellow; pubescence on pronotum entirely yellowish-brown, almost inconspicuous due to integument color; pubescence on pronotum vellowish-brown, conspicuous, with sparse, whitish setae interspersed; pubescence on prosternum and prosternal process yellowish-white; dense pubescent band on mesanepisternum yellowish-white; dense pubescence on metanepisternum light yellowish-brown or yellow; dense pubescence on metaventrite yellowish-brown, or metaventrite without distinctly dense pubescence contrasting with remaining pubescence; scutellum with dense yellow pubescence; elytral apex in small specimens obliquely truncate, making outer angle somewhat acute (Fig. 15); bands of dense pubescence on elytra yellow; humeral area with yellowish pubescence; central and posterior pubescent bands of elytra not distinctly projected forward along suture; meso- and metafemora in small males female-shaped (club not abruptly widened from peduncle) pubescence and erect setae on femora and tibiae yellowish-brown.

Dimensions (mm), male/female. Total length, 7.55-12.55/8.85-13.40; prothoracic length, 1.75-2.90/1.90-2.35; anterior prothoracic width, 1.40-2.10/1.65-2.00; posterior prothoracic width, 1.30-1.95/1.50-1.90; maximum prothorax width, 2.00-3.25/2.25-2.90; humeral width, 2.00-3.20/2.25-2.90; elytral length, 5.30-8.80/5.80-8.20.

Material examined. BOLIVIA, Santa Cruz: 10 km W Buena Vista-San Mateo road (7,000'), 1 male, 13.X.2006, Wappes, Nearns, and Eya col. (ACMT); Achira (Vicoquin area), 1 female, 8.X.2007, Wappes and Morris col. (MZSP); (5.5 km NE of Achira, Vicoquin area, Subtropical Forest; flying to fresh cut trees; 1800 m), 1 male, 25.XI.2004, no collector indicated (MZSP); 4–5 km N Achira (Road to Amboró), 1 male, 12–13.X.2000, Wappes and Dozier col. (ACMT); Florida prov. (16 km NE Mairana; 6,600'; 18°05'S / 63°54'W), 4 males, 6 females, 11.XII.2011, Wappes, Bonaso, and Sekerka col. (ACMT); Rd to Amboro above Achira, chaco, 18°07.43'S, 63°47.98'W, 2 males, 14–15.X.2006, Wappes, Nearns and Eya col. (ACMT); 1 male, 9–11.X.2004, Wappes and Morris col. (ACMT).

Remarks. Amyipunga armaticollis (Zajciw, 1964) is redescribed here to correct errors in its original description, provide a more complete description including intraspecific variation, and detail the differences between it and the closely related *Megacyllene moritzii* Thomson from Venezuela. It should also be noted that the type was lost in the 2018 fire that destroyed the MNRJ and all its contents.

Zajciw (1964) described Neoclytus armaticollis from Bolivia (Santa Cruz), based on a single specimen.

According to him, the holotype was a male. However, the photograph of the holotype (based on antennal length, and shape of the metafemora), indicates it is a female.

According to Martins and Galileo (2011) (translated): "Zajciw (1964: 305) based the description of A. armaticollis on a male, whose apices of the metafemora, apparently, do not extend beyond the tips of the elytra. We examined a specimen, which we judge to be male, where the apices of the metafemora overlap the elytra with less than half of the club. The length of the metafemora is important to differentiate A. armaticollis from A. moritzii Thomson, in which the metafemora overlap the apex of the elytra at the base of the club, in addition to being remarkably thickened. A. moritzii also differs from A. armaticollis by the white pubescence of the ventral face of the body, which covers small portion of the lateral parts of the metaventrite and occupies much narrower areas in the abdominal segments." Based on multiple specimens of both sexes of Amyipunga armaticollis examined, the amount of ventrite pubescence varies significantly and as such is not a diagnostic character to separate it from A. moritzii. However, the longer length of the metafemora in both sexes in the latter is a consistent character that reliably separates them.

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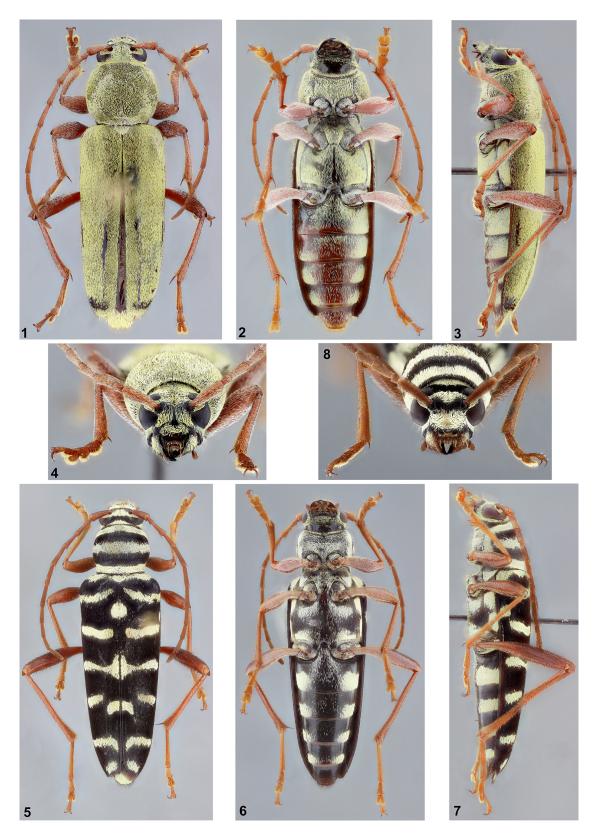
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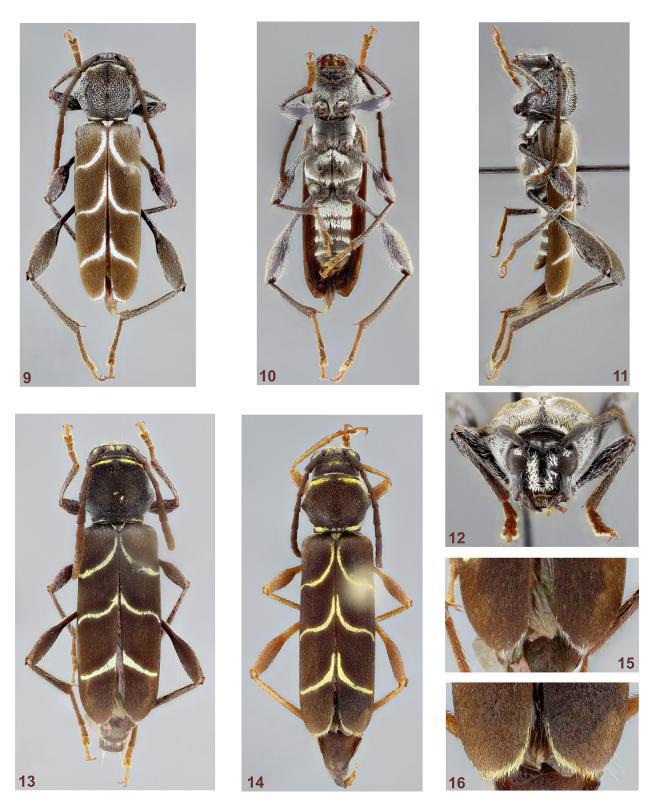
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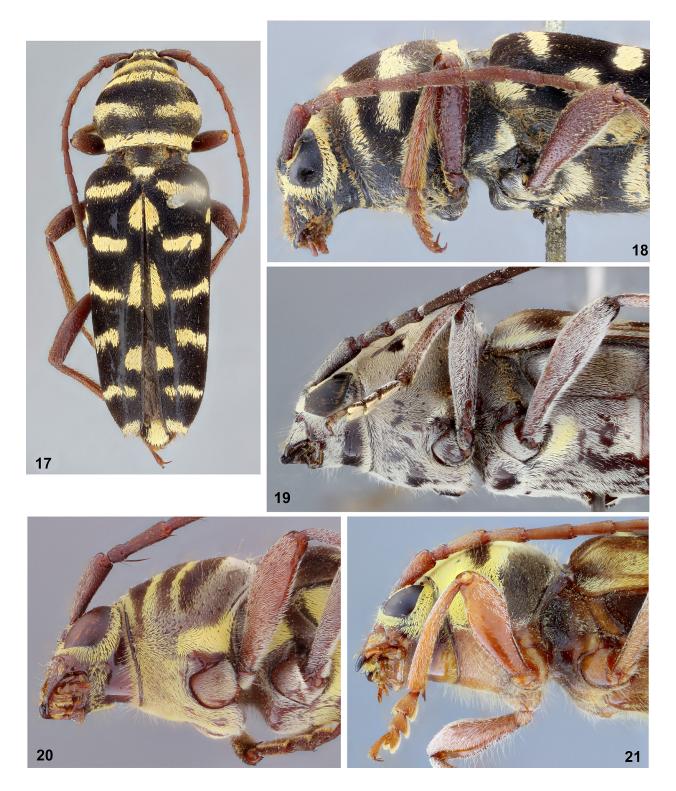
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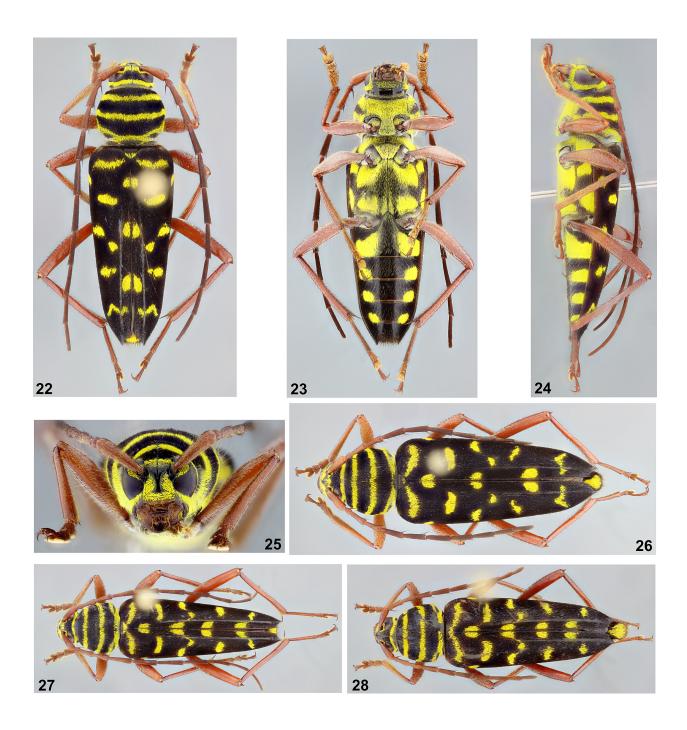
Figures 1–8. 1–4) *Trichoxys giesberti*, holotype female. 1) Dorsal habitus. 2) Ventral habitus. 3) Lateral habitus. 4) Head, frontal view. 5–8) *Megacyllene giesberti*, holotype female. 5) Dorsal habitus. 6) Ventral habitus. 7) Lateral habitus. 8) Head, frontal view.



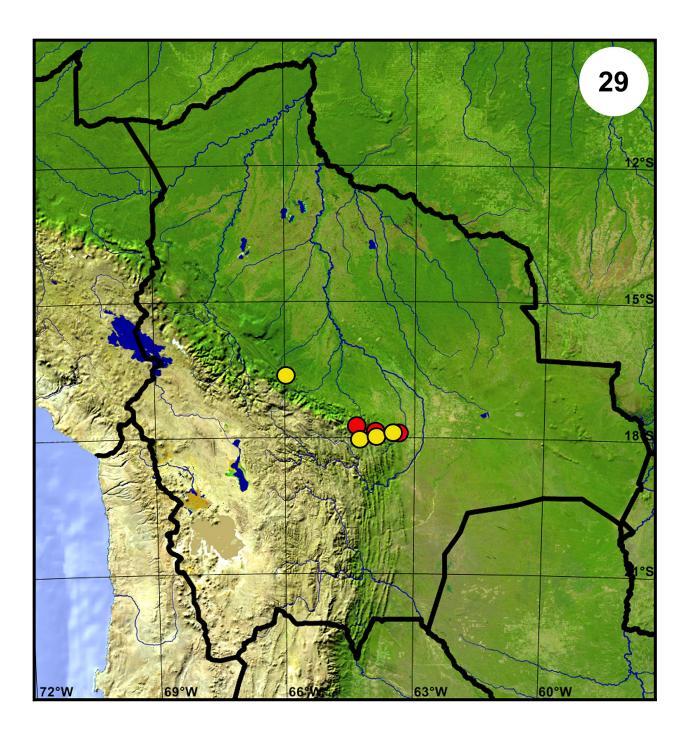
Figures 9–16. Amyipunga armaticollis. 9–12) Male. 9) Dorsal habitus. 10) Ventral habitus. 11) Lateral habitus. 12) Head, frontal view. 13–14) Female, dorsal habitus. 13) Specimen 1. 14) Specimen 2. 15–16) Elytral apex, female. 15) Specimen 1. 16) Specimen 2.



Figures 17–21. 17) Megacyllene asteca, female, dorsal habitus. 18–21) Prosternal process. 18) M. asteca, female. 19) M. nebulosa, female. 20) M. acuta, male. 21) Plagionotus detritus, female.



Figures 22–28. Megacyllene melanaspis. 22–26) Male. 22) Dorsal habitus. 23) Ventral habitus. 24) Lateral habitus. 25) Head, frontal view. 26) Female, dorsal habitus. 27–28) Megacyllene proxima, dorsal habitus. 27) Male. 28) Female.



 $\textbf{Figure 29.} \ \text{Map of known distribution in Bolivia. Red ball}, \textit{Megacyllene melanaspis}; yellow ball, \textit{Megacyllene proxima}.$