# Morphological and Molecular Characterization of *Gracilacus wuae* n. sp. (Nematoda: Criconematoidea) Associated with Cow Parsnip (*Heracleum maximum*) in Ontario, Canada

Qing Yu,  $^1$  Weimin Ye,  $^2$  and Tom Powers  $^3$ 

Abstract: Gracilacus wuae n. sp. from soil associated with cow parsnip in Ontario, Canada is described and illustrated. Morphologically, females have a long stylet ranging from 80 to 93  $\mu$ m long, the lip region not offset from the body contour, without lateral lips but with large and flat submedian lobes, the mouth opening slit-like elongated laterally and surrounded by lateral flaps, the excretory pore is anterior to the knobs of the stylet; males without stylet and the pharynx degenerated. The fourth-stage juveniles lack a stylet, the pharynx degenerated, and can be differentiated into preadult females and males based on the position of the genital primordia. The third-stage juveniles are similar to females but smaller. Phylogenetic studies using the rDNA small subunit 18S, large subunit 28S D2/D3, and internal transcribed spacer (ITS) sequences collectively provide evidence of a grouping with other *Gracilacus* and some species of *Paratylenchus* with stylet length of females longer than 41  $\mu$ m deposited in GenBank.

Key words: Gracilacus, molecular, morphology, morphometric, new species, Paratylenchus, taxonomy.

The genus *Gracilacus* was established by Raski (1962) to differentiate species of *Paratylenchus* Micoletzky, 1922, based on primarily stylet length and secondarily on body shape and the position of the excretory pore in females. Eight species with a female stylet longer than 48 µm were separated and placed in the genus Gracilacus primarily based on that single character. Shortly after the initial creation of Gracilacus, Siddiqi and Goodey (1964) synonymized Gracilacus with Paratylenchus. Raski (1976) revised Gracilacus, lowered the minimum stylet length to 41 um, and added 13 additional species to the genus. Siddiqi (1986) established the subgenera Gracilacus and Paratylenchus within the genus Paratylenchus based on stylet length. Thorne and Malek (1968) accepted Gracilacus when they described two new Paratylenchus species and evaluated the taxonomic position of G. audriellae (Brown, 1959) Raski, 1962. G. audriellae was later considered as a junior synonym of G. straeleni by Siddiqi (2000). Geraert (1965) divided Paratylenchus sensu lato that includes *Gracilacus* into 10 groups based on the stylet length and the V-position and recently Ghaderi et al. (2014) expanded it by dividing the genus into 11 groups based on stylet length, lateral fields, and advulval flaps. With the advent of molecular biology, phylogenetic studies have been conducted to examine the relationships among paratylenchids including very few species of Gracilacus. Lopez et al. (2013) used the first ITS1 region to examine paratylenchid relationships including G. bilineata Brzeski, 1995 and G. aculenta (Brown,

1959) Raski, 1962 in their analyses. Van Den Berg et al. (2014) used 28S D2/D3 and ITS sequence to examine species groupings within *Paratylenchus* including the above two *Gracilacus* species. Wang et al. (2015) described *P. guangzhouensis*, a species with the stylet averaging 47  $\mu$ m long, and used ITS sequence to show that this new species is clustered with those four previously sequenced *Gracilacus* species. In another study, Wang et al. (2016) also used ITS to demonstrate that their newly described species *P. nanjingensis* with a 66  $\mu$ m long stylet is grouped with *G. bilineata* and *G. aculenta*. All these studies provided evidence of the possible existence of a genetic grouping of long-stylet-bearing paratylenchids.

This study describes a new species of *Gracilacus* from Canada. There are 43 species listed in the subgenus *Gracilacus* by Siddiqi (2000). Four species have been previously recorded from Canada (Brown, 1959; Wu, 1974). These include *G. acicula* (Brown, 1959) Raski, 1962, *G. aculenta*, *G. audriellus*, and *G. robusta* (Wu, 1974) Raski, 1976. The new species described herein, *G. wuae* n. sp., was found in soil associated with wild parsnip (*Heracleum maximum*) in Ontario, Canada.

#### MATERIALS AND METHODS

Nematode samples: Soil samples associated with wild parsnip were collected in Kanata, west of Ottawa, Ontario, Canada, with latitude 45°18′43.33″N and longitude 75°55′54.08″W. The nematodes were extracted with the Baermann funnel method. Freshly extracted nematodes were processed for both morphological and molecular studies.

Morphological study: The method used was as described previously (Yu et al., 2014). The nematodes were all fixed in TAF and mounted in dehydrated glycerin on slides for morphological studies. Specimens were examined using a Leica DM5500 B compound microscope (Wetzlar, Germany) using differential interference contrast and pictures were taken with a Leica

Received for publication May 5, 2016.

<sup>&</sup>lt;sup>1</sup>Ottawa Research and Development Center, Agriculture and Agri-Food Canada, Ottawa, ON K1A 0C6, Canada.

<sup>&</sup>lt;sup>2</sup>Nematode Assay Section, Agronomic Division, North Carolina Department of Agriculture & Consumer Services, Raleigh, NC 27607-6465.

<sup>&</sup>lt;sup>3</sup>Department of Plant Pathology, University of Nebraska–Lincoln, Lincoln, NE 68583-0722.

This article was registered to the Official Register of Zoological Nomenclature (ZooBank) as 16B8EBD7-4107-438E-A2D2-04AC63B3968B.

The authors would like to thank Dr. Z.A. Handoo, curator for the USDA nematode collection for lending the type materials of *G. macrodorus*, and Denise Chabot at the Ottawa Research and Development Center for helping the SEM works. Tom Powers would like to acknowledge funding from NSF DEB-1145440.

E-mail: qing.yu@agr.gc.ca. This paper was edited by Sergei A. Subbotin.

DFC 420 digital camera. Measurements were made using a Leica micro application system on the images. Scanning electron microscope (SEM) pictures were taken with a FEI Quanta 600 SEM.

## Molecular study

DNA extraction: DNA extraction solution consisted of  $1 \times PCR$  buffer containing Proteinase K at a concentration of 0.25 mg/ml. Five nematodes were crushed in  $50~\mu l$  of the DNA extraction solution and incubated at  $65^{\circ}C$  for 2.5 hr, followed by incubation at  $95^{\circ}C$  for 15 min and on ice for 5 min. The DNA extracts were stored for later use at  $-20^{\circ}C$ .

PCR and sequencing: Internal transcribed spacer was amplified with the primers ITS-F (5'-TTGATTACGTC CCTGCCCTTT-3') and ITS-R (5'-TTTCACTCGCCGT TACTAAGG-3') (Vrain et al., 1992) using the following thermocycling conditions: an initial denaturation at 95°C for 3 min, followed by 40 cycles of denaturation (30 sec at 95°C), annealing (30 sec at 56°C), and extension (1 min at 68°C), and a final extension step (3 min at 68°C). 18S rRNA gene (18S) was amplified with the primers 18S-F (5'-CGATCAGATACCGCCCTAG-3') and 18S-R (5'-TACAAAGGGCAGGACGTAAT-3') (Powers et al., 2010) using the following thermocycling conditions: an initial denaturation at 95°C for 3 min, followed

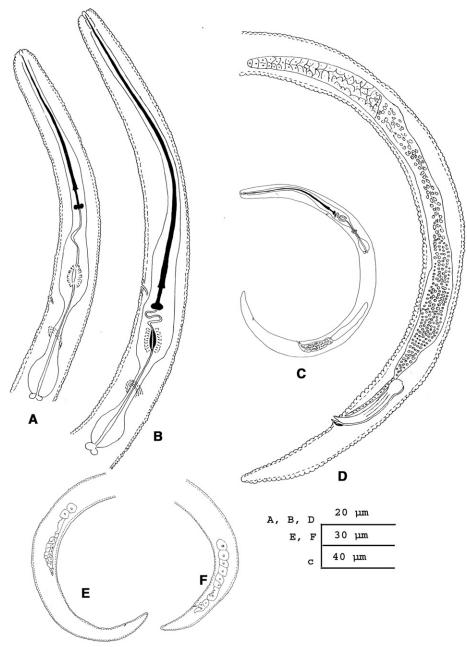


Fig. 1. Line drawing of *Gracilacus wuae* n. sp. associated with cow parsnip in Ontario, Canada, A. Head and pharynx of the third-stage juvenile. B. Head and pharynx of female. C. Female body. D. Male genital system. E. Posterior half of preadult female. F. Posterior half of preadult female.

by 30 cycles of denaturation (30 sec at 95°C), annealing (30 sec at 62°C), and extension (1.5 min at 68°C), and a final extension step (3 min at 68°C). D2/D3 region of the 28S rRNA gene (28S D2/D3) was amplified with the primers D2A (5'-GACCCGTCTTGAAACACGGA-3') (Nunn, 1992) and D3B (5'-TCGGAAGGAACCAGCTACTA-3') (De Ley et al., 1999), using the following thermocycling conditions: an initial denaturation at 95°C for 3 min, followed by 40 cycles of denaturation (30 sec at 95°C), annealing (30 sec at 50°C), and extension (1 min at 68°C), and a final extension step (3 min at 68°C).

All PCR reactions were prepared using the Titanium Taq PCR kit (Clontech Laboratories Inc., Mountain View, CA) and contained 5 or 10 ul of DNA, 2.5 μl of 10× PCR buffer, 1.0 µl of 2.0 mM dNTPs, 0.4 µl 10 µM primers, and 0.2 µl of Titanium Taq DNA polymerase. One microliter of PCR product was used per sequencing reaction. Sequencing was performed as described previously (Marchand et al., 2014) and samples were run on an ABI 3130xl Avant sequencer (Applied Biosystems). Consensus sequences of forward and reverse sequence reactions were obtained and used for a homology search of the National Centre for Biotechnology Information's GenBank database using Basic Local Alignment Search Tool (BLAST).

Phylogenetic study: Phylogenetic analysis was performed as described previously (Zeng et al., 2013). The sequences were deposited into the GenBank database. DNA sequences were aligned by Clustal W using default settings. The DNA sequences of G. wuae n. sp. were compared with those of the other nematode species available at the sequence database using the BLAST homology search program. The model of base substitution was evaluated using MODELTEST, and the Akaike information criterion (AIC) supported model, the base frequencies, the proportion of invariable sites, and the gamma distribution shape parameters and substitution rates were used in phylogenetic analyses. Bayesian analysis using GTR + I + G was performed to confirm the tree topology for each gene separately using MrBayes 3.1.0 (Ronquist and Huelsenbeck, 2003) running the chain for  $1 \times 10^6$  generations and setting the "burnin" at 1,000. We used the Markov Chain Monte Carlo method within a Bayesian framework to estimate the posterior probabilities of the phylogenetic trees using 50% majority rule.

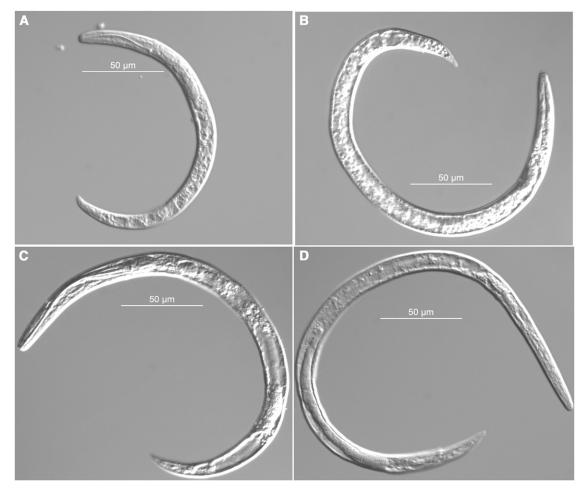


Fig. 2. Light micrographs of Gracilacus wuae n. sp. associated with cow parsnip in Ontario, Canada. A. Body of the third-stage juvenile. B. Body of the fourth-stage juvenile. C. Body of female. D. Body of male.

#### RESULTS AND DESCRIPTION

#### Systematics

Gracilacus wuae n. sp.\* (Figs. 1–4)

*Measurements*: Morphometrics of the holotype, paratype of females, males, and the third- and fourth-stage juveniles of *G. wuae* n. sp. are given in Table 1.

### Description

Female: Body not obese, C-shaped when relaxed; annuli delicate but distinct, lateral fields, each occupy one quarter of the body width, with four straight lines with equal space extending to the full length of the body. Lip region with pronounced large and flat submedian lobes, but without lateral lips (Fig. 4A,B), the mouth aperture is slit like, elongated laterally and surrounded by lateral flaps, submedian lobes rounded, lip region not offset from the body contour. Cephalic frame well developed. Stylet is elongated, 80 to 93  $\mu m$  long, slightly curved ventrally, cone is about 20 times length of shaft and knobs combined, and stylet knobs

large, and robust. Metacorpus is large, well developed, and occupying about three-fourth of body width. Valvular apparatus large and muscular. Isthmus is narrow and surrounded by nerve ring. Basal bulb pyriform. Cardia consists of two rounded cells. Hemizonid is about 3  $\mu m$  anterior to the excretory pore, which is positioned anterior to stylet knobs, and in the range of the stylet shaft. Gonad is prodelphic, short, and outstretched; spermatheca round, offset, filled with globular sperms. Advulval flaps absent. Post uterine sac absent. Anus rather prominent. Tail terminus finely rounded with distinct annuli.

*Male:* Body slender than female body. Lip region differs to that of females without distinct lips or mouth opening (Fig. 4C). Cephalic framework less developed. Pharynx rudimentary, only vague contour of procorpus and metacorpus can be discerned. Stylet lacking. Positions of hemizonid and excretory pore equivalent to those of females. Body annuli more visible than those of females, especially in tail region. Lateral fields with four lines. Gonad about 110  $\mu$ m long. Cloacal opening elevated, forming a short penial sheath. Spicules well

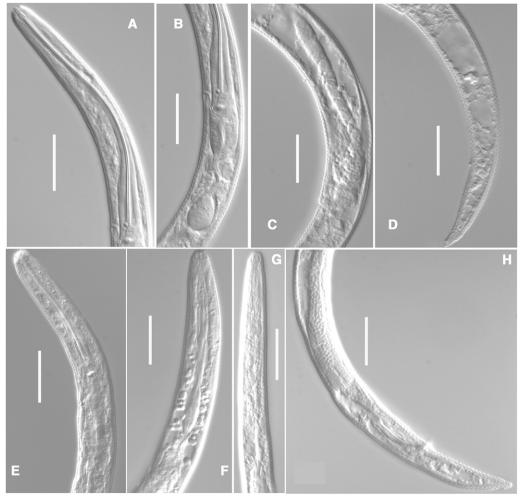


Fig. 3. Light micrographs of *Gracilacus wuae* n. sp. associated with cow parsnip in Ontario, Canada. A. Head and pharynx of female. B. Metacorpus and basal bulb of pharynx of female. C. Female genital. D. Female tail. E. Head of the third-stage juvenile. F. Head of the fourth-stage juvenile. G. Head of male. H. Male genital and tail region.

developed, slightly curved ventrally, protruding from body. Gubernaculum about 4 µm long. Bursa absent. Tail terminus similar to that of female.

The second-stage juvenile: Not found.

The third-stage juvenile: Similar to females, but smaller. The body C shaped when relaxed. The stylet is 46 to 52 µm long, almost half of the stylet length of the female, pharynx well developed like in female. Genital primordia visible, located at about 50 µm from the posterior end of the nematode or about 30 µm anterior to the anus.

The fourth-stage juvenile: The fourth-stage juveniles have no stylet, pharynx reduced but clearly defined, no valvar apparatus, anterior portion of lumen somewhat dark, sclerotized. They can be clearly classified into preadult female (Fig. 1E) and preadult male (Fig. 1F) based on the positions of the genital primordia. Preadult male: the posterior end of the genital primordia is near the anus. Preadult female: the posterior end of the genital primordia is 38 to 51 µm anterior to the anus. The ratio of preadult female/preadult male is 2 (total number of preadult female recovered is 24, preadult male 12); the ration is nearly the same of adults (1.9): female (40)/male (21).

Type host and locality: Roadside in Kanata, west of Ottawa, Ontario, Canada, with latitude 45°18'43.33"N and longitude 75°55′54.08″W. It was found in soil associated with cow parsnip (H. maximum), a fairly common weed in the region. Subsequent sampling in other places with cow parsnip did not recover the nematode. The roadside location of the positive collection site had recently been excavated, so it is not known if the disturbed roadside habitat fully represented the original soil and vegetation prior to road construction.

Type specimens: Holotype female, 10 paratype females, 10 males, 10 third-stage juveniles, and 10 preadult

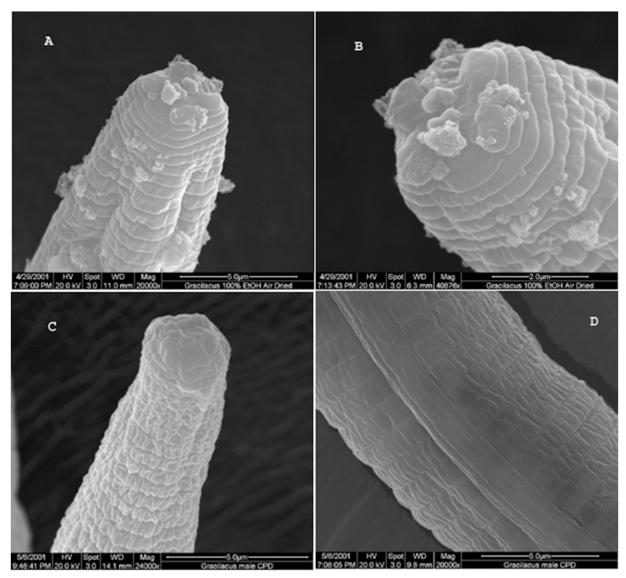


Fig. 4. Scanning electron microscope pictures of Gracilacus wuaen. sp. associated with cow parsnip in Ontario, Canada. A. Head of female. B. Lip region of female. C. Head of male. D. Lateral field of male.

Morphometrics of Gracilacus waren. sp. All measurements are in µm and in the form: mean ± standard deviation (range) Table 1.

				Paratypes		
	Holotype				J4 (Preadult)	eadult)
Character	Female	Females	Males	J3	Female	Male
n	ı	13	6	6	6	6
Γ	329	$326 \pm 17.9 (300-360)$	$380 \pm 12.2 \ (369-405)$	$233 \pm 4.9 \ (225-239)$	$323 \pm 8.2 \ (320-343)$	$329 \pm 8.4 \ (320-353)$
а	20.5	$20.7 \pm 0.8 \ (20.1 - 22.2)$	$29.1 \pm 1.1 \ (26.4-30.8)$	$17.6 \pm 1.3 \ (15.3 - 19.8)$	$22.7 \pm 1.7 \ (20-26)$	$21.7 \pm 1.8 \ (19-25)$
P	2.5	$2.4 \pm 0.1 \ (2.3-2.6)$	$3.9 \pm 0.2 \ (3.9 - 4.1)$	$2.5 \pm 0.1 \ (2.4-2.6)$	$3.5 \pm 0.1 \ (3.4-3.7)$	$3.4 \pm 0.2 \ (3.1 - 3.5)$
C	11.0	$10.9 \pm 0.3 \ (10.5 - 11.3)$	$12.3 \pm 0.7 \ (11.2 - 13.2)$	$10.9 \pm 0.4 \ (10.4 - 11.4)$	$13.1 \pm 1.3 \ (11.8-15.6)$	$12.9 \pm 1.3 \ (11.2 - 15.1)$
c,	3.6	$3.2 \pm 0.2 \ (3.4 - 3.8)$	$3.7 \pm 0.1 \ (3.6 - 3.9)$	$2.0 \pm 0.1 \ (1.8-2.2)$	$2.5 \pm 0.1 \ (2.2-2.6)$	$2.3 \pm 0.1 \ (2.2-2.6)$
Λ	75.9	$74.7 \pm 3.8 \ (69.9-80.1)$		1		1
G or T	20.1	$20.1 \pm 0.5 \ (18.8 - 20.3)$	$37.4 \pm 1.2 \ (35.7 - 38.9)$	1		ı
Stylet	91.6	$88 \pm 4.7 (80-93)$	1	$48 \pm 2.4 (46-52)$		ı
Body diameter	16.1	$15.7 \pm 0.5 \ (18.8 - 20.4)$	$13.0 \pm 0.6 \ (12-14)$	$13.3 \pm 0.9 (12-15)$	$14.8 \pm 0.9 \ (13-16)$	$12.8 \pm 0.9 \ (11-15)$
Tail	30	$29.8 \pm 1.6 \ (27.1 - 32.1)$	$31.1 \pm 1.1 \ (30-33)$	$21.3 \pm 0.7 (20-22)$	$25.6 \pm 2.6 (22-28)$	$26.6 \pm 2.7 (23-30)$
Spicules	ı	1	$21.8 \pm 0.4 \ (21-22)$	1		ı
Distance of the posterior end	ı	1	1	ı	$60 \pm 6.5 (51-67)$	$25 \pm 2.5 (23-28)$
of the genital primordia						
to the tail tip						
Length of the genital primordia	ı				$42 \pm 3.8 \ (39-47)$	$48 \pm 4.0 \ (44-52)$

females and males each were deposited in the Canadian National Collection of Nematodes under accession number T530. Five paratypes for each stage were in USDA Nematode Collection, Beltsville, MD, with accession numbers T-6778p to T-6797p.

Diagnosis: This new species is characterized by the combination of the stylet 80 to 93 µm long, the lip region continuous without lateral lips, with large and flat submedian lobes, lateral fields with four lines, the anterior positioned excretory pore relative to the knobs of the stylet, and lateral vulvar membranes absent in females.

Relationships: According to the grouping scheme of Paratylenchus sensu lato by Ghaderi et al. (2014), this new species belongs to Group 11 (stylet >40 μm; four lateral lines; and advulval flaps absent). Within this group, the species with stylet longer than 67 µm includes P. laocaiensis Nguyen, Baldwin & Choi, 2004, G. teres Raski, 1976, G. enata Raski, 1976, G. oostenbrinki (Misra and Edward, 1971) Raski, 1976, G. steineri (Golden, 1961) Raski, 1962, and G. macrodorus. Scanning electron microscope en face views of G. teres (Pena and Geraert, 1990) were also used for comparison. This new species differs from P. laocaiensis by a longer stylet (88 [80-93] vs. 73 [67-82] µm), more posterior positioned vulva (V 70–80 vs. 65–67), submedian lobes, and rounded tail terminus vs. pointed terminus. It differs from G. teres by having a longer stylet (88 [80–93] vs. 73 [69–83] µm), the absence of lateral lips, submedian lobes larger and flatter, spermatheca filled with sperms, and conspicuous vulva. It differs from G. enata by having a shorter stylet (88 [80–93] vs. 93 [83–104] μm), a wider female body (a 20–22 vs. 23–28), longer male (309–405 vs. 340–350). It differs from G. oostenbrinki by longer female body  $(300-360 \text{ vs. } 250-280 \text{ } \mu\text{m})$ , longer stylet (80-93 vs.72–88 µm), more anterior positioned excretory pore, and rounded tail terminus vs. acute. It differs from G. teineri by having longer stylet (88 [80–93] vs. 73 [68–75] μm), and rounded tail terminus vs. subacute pointed.

Etymology: \*The species was named after Dr. Liangyu Wu, a former nematode taxonomist at the Agriculture and Agri-Food Canada, who made significant contribution to the taxonomy of Paratylenchus and Gracilacus.

### Phylogenetic analysis

Three regions of the rDNA repeating region were sequenced and compared with sequences deposited in GenBank. The stylet length greater than 41 µm was mapped to the species in phylogenetic trees (Figs. 5-7).

18S: A 979-bp fragment of the 3' region of 18S rDNA gene was sequenced for G. wuae n. sp. Rooted with Hemicriconemoides wessoni, the 18S tree (Fig. 5) placed G. wuae n. sp. in a clade with G. latescens (AY912039) from Konza Prairie, KS, Paratylenchus colinus (KP966494) from apple in Iran and Gracilacus sp. (KR232931) from pine tree in China with 82% support. These species have

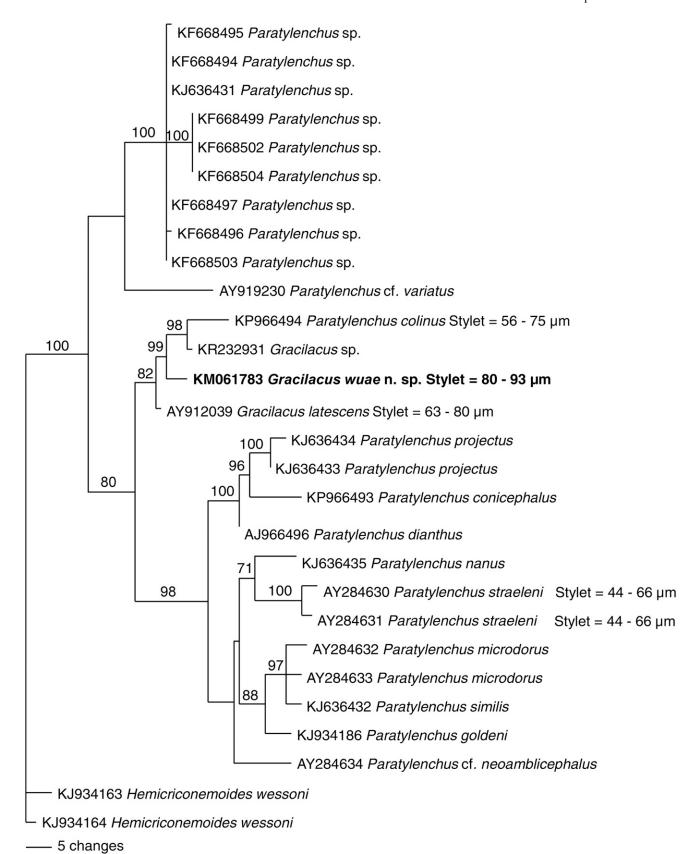
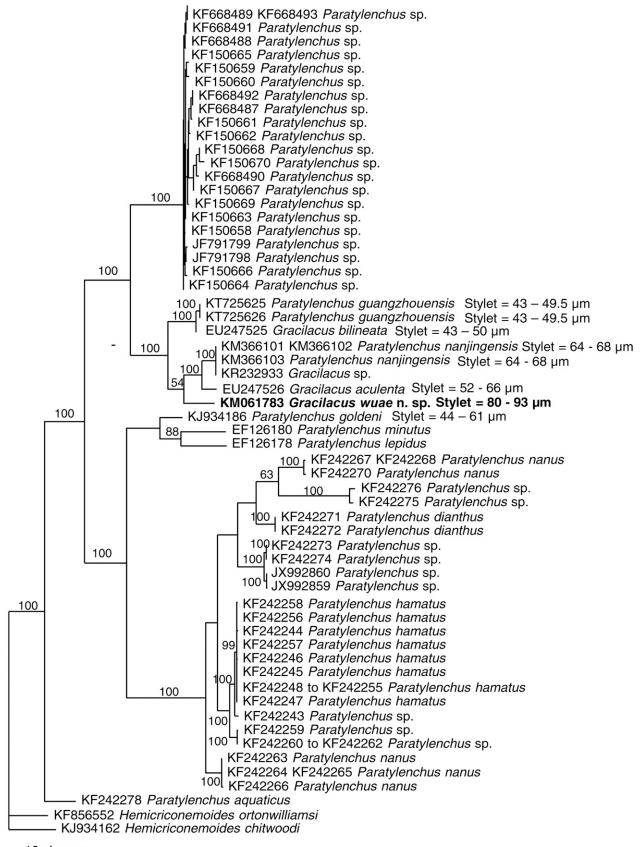


Fig. 5. The 10,001st Bayesian tree inferred from 18S under GTR + I + G model (-lnL = 2,216.9043; AIC = 4,453.8086; freqA = 0.2305; freqC = 0.2442; freqG = 0.2862; freqT = 0.2391; R(a) = 0.2285; R(b) = 1.7446; R(c) = 1.0079; R(d) = 0.875; R(e) = 4.7408; R(f) = 1; Pinva = 0.7656; shape = 1.1055). Posterior probability values exceeding 50% are given on appropriate clades.

#### — 10 changes

Fig. 6. The 10,001st Bayesian tree inferred from 28S D2/D3 under GTR + I + G model (-lnL = 5,683.3032; AIC = 11,386.6064; freqA = 0.2094; freqC = 0.2291; freqG = 0.3176; freqT = 0.2438; R(a) = 0.513; R(b) = 2.0052; R(c) = 0.7642; R(d) = 0.4773; R(e) = 4.4825; R(f) = 1; Pinva = 0.2154; shape = 0.5762). Posterior probability values exceeding 50% are given on appropriate clades.



# 10 changes

Fig. 7. The 10,001st Bayesian tree inferred from ITS under GTR + I + G model (-lnL = 12,135.0107; AIC = 24,290.0215; freqA = 0.2228; freqC = 0.2549; freqG = 0.2804; freqT = 0.2419; R(a) = 1.3527; R(b) = 3.0954; R(c) = 2.0549; R(d) = 0.8779; R(e) = 3.8983; R(f) = 1; Pinva = 0.2508; shape = 1.1435). Posterior probability values exceeding 50% are given on appropriate clades.

stylet more than 41  $\mu$ m long. Paratylenchus straeleni (AY284630, AY284631) has a stylet 44 to 66  $\mu$ m long, but grouped with other Paratylenchus species. P. straeleni has been considered as members of the subgenus Gracilacus (Wouts, 2006). This tree revealed long-stylet paratylenchids are not monophyletic.

28S D2/D3: The 873-bp DNA fragment of the 28S D2/D3 was sequenced for *G. wuae* n. sp. Rooted with *Hemicycliophora lutosa*, the tree (Fig. 6) placed *G. wuae* n. sp. in a 100%-supported monophyletic clade with other long-stylet species, namely *G. audriellus* (KU291238), *G.* sp. (KR232932), *P. aculentus* (KR270597), and *P. colinus* (KP966492). Two populations of *P. straeleni* (KF242235, KF242236) from Mendocino and Napa Counties, CA (Van Den Berg et al., 2014) were grouped with other species of *Paratylenchus*. As in the 18S tree, the long-stylet paratylenchids are not monophyletic if *P. straeleni* is considered as a member of *Gracilacus*.

ITS: A 929-bp fragment of ITS1 was sequenced for *G. wuae* n. sp. Rooted with *Hemicriconemoides chitwoodi*, the ITS tree (Fig. 7) placed *Gracilacus* in a 100%-supported monophyletic clade with other long-stylet species, namely *P. guangzhouensis* (KT725625, KT725626), *P. nanjingensis* (KM366102-KM366103), *G. bilineata* (EU247525), *G.* sp. (KR232933), and *G. aculenta* (EU247526). This clade is equivalent to Clade III in the ITS tree by Van Den Berg et al. (2014). Another long-stylet species *P. goldeni* (KJ934186) is together with short stylet paratylenchids. As in 18S and 28S D2/D3, the long-stylet paratylenchids are not monophyletic. Unfortunately, no ITS sequence data are available for *P. straeleni* to compare the result.

Species of paratylenchids are small nematodes with stylet lengths that vary from 10 to 120 µm. This large variation is rare in tylenchids. In this study, the new species G. wuae n. sp. is grouped with other long-stylet species when nucleotide sequences were compared, but with exceptions in all three DNA fragments sequenced. Based on this study, the character "stylet length greater than 41 µm" is not homologous and has evolved more than once within the Paratylenchus lineage. Given this combination of molecular and morphological evidence, we believe it would be taxonomically appropriate to recognize the new species as a member of the genus Gracilacus, although few species are available for comparison. It is clear, however, that considerable taxonomic effort and DNA sequencing on the vast majority paratylenchids will be necessary to clarify relationships and classification within Paratylenchidae.

# LITERATURE CITED

Brown, G. L. 1959. Three new species of the genus *Paratylenchus* from Canada (Nematoda: Criconematidae). Proceedings of the Helminthological Society of Washington 26:1–8.

De Ley, P., Felix, M. A., Frisse, L. M., Nadler, S. A., Sternberg, P. W., and Thomas, W. K. 1999. Molecular and morphological characterisation of two reproductively isolated species with mirror image anatomy (Nematoda: Cephalobidae). Nematology 2:591–612.

Geraert, E. 1965. The genus *Paratylenchus*. Nematologica 11:301–334

Ghaderi, R., Kashi, L., and Karegar, A. 2014. Contribution to the study of the genus *Paratylenchus* Micoletzky, 1922 *sensu lato* (Nematoda: Tylenchulidae). Zootaxa 3841:151–187.

Lopez, M. A. C., Robbins, R., and Szalanski, A. L. 2013. Taxonomic and molecular identification of *Hemicaloosia, Hemicycliophora, Gracilacus* and *Paratylenchus* species (Nematoda: Criconematidae). Journal of Nematology 45:145–171.

Marchand, G., Chen, Y., Berhane, N. A., Wei, L., Lévesque, C. A., and Xue, A. G. 2014. Identification of *Pythium* spp. from the rhizosphere of soybeans in Ontario, Canada. Canadian Journal of Plant Pathology 36:246–251.

Micoletzky, H. 1922. Die Freilebenden Erd-Nematoden. Archiv für Naturgeschichte. Berlin A $87{:}1{-}650.$ 

Misra, S. L., and Edward, J. C. 1971. Two new species of the genus *Paratylenchus* with description of their larval stages and a note on *P. nawadus* a synonym of *P. nainianus*. The Allahabad Farmer 45:345–350

Nunn, G. B. 1992. Nematode molecular evolution. PhD dissertation, University of Nottingham, UK.

Pena, S. R., and Geraert, E. 1990. New data on *Aorolaimus perscitus* (Doucet, 1980) and *Gracilacus teres* Raski, 1976 (Nematoda: Tylenchida) associated with olive (*Olea Europea* L.) in the Province of Jaén, Spain. Nematologica 36:408–416.

Powers, T. O., Harris, T., Higgins, R., Sutton, L., and Powers, K. S. 2010. Morphological and molecular characterization of *Discocriconemella inarata*, an endemic nematode from North American native tallgrass prairies. Journal of Nematology 42:35–45.

Raski, D. J. 1962. Paratylenchidae n. fam. with descriptions of five new species of *Gracilacus* n.g. and an emendation of *Cacopaurus* Thorne, 1943, *Paratylenchus* Micoletzky, 1922 and Criconematidae Thorne, 1943. Proceedings of the Helminthological Society of Washington 29:189–207.

Raski, D. J. 1976. Revision of the genus *Paratylenchus* Micoletzky, 1922 and descriptions of new species. Part III of three parts—*Gracilacus*. Journal of Nematology 8:97–115.

Ronquist, F. and Huelsenbeck, J. P. 2003. MRBAYES 3: Bayesian phylogenetic inference under mixed models. Bioinformatics 19: 1572–1574.

Siddiqi, M. R. 1986. Tylenchida: Parasites of Plants and Insects. 1st ed. Farnham Royal, UK: Commonwealth Agricultural Bureau.

Siddiqi, M. R. 2000. Tylenchida: Parasites of Plants and Insects. 2nd ed. Wallingford, UK: CABI Publishing.

Siddiqi, M. R., and Goodey, J. B. 1964. The status of the genera and subfamilies of the Criconematidae (Nematoda); with a comment on the position of *Fergusobia*. Nematologica 9:363–377.

Thorne, G., and Malek, R. B. 1968. Nematodes of the Northern Great Plains part I. Tylenchida [Nemata: Secernentea]. Brookings, SD: South Dakota University.

Van Den Berg, E., Tiedt, L. R., and Subbotin, S. A. 2014. Morphological and molecular characterisation of several *Paratylenchus* Micoletzky, 1922 (Tylenchida: Paratylenchidae) species from South Africa and USA, together with some taxonomic notes. Nematology 16:323–358.

Vrain, T. C., Wakarchuk, D. A., Lévesque, A. C., and Hamilton, R. I. 1992. Intraspecific rDNA restriction fragment length polymorphism in the *Xiphinema americanum* group. Fundamental and Applied Nematology 15:563–573.

Wang, K., Li, Y., Xie, H., Xu, C. L., and Wu, W. J. 2015. Morphology and molecular analysis of *Paratylenchus guangzhouensis* n. sp.

(Nematoda: Paratylenchinae) from the soil associated with *Bambusa multiplex* in China. European Journal of Plant Pathology DOI 10.1007/s10658-015-0836-z.

Wang, K., Xie, H., Li, Y., Wu, W. J., and Xu, C. L. 2016. Morphology and molecular analysis of *Paratylenchus nanjingensis* n. sp. (Nematoda: Paratylenchinae) from the rhizosphere soil of *Pinus massoniana* in China. Journal of Helminthology 90:166–173.

Wouts, W. M. 2006. Criconematina (Nematoda: Tylenchida). Fauna of New Zealand 55. Lincoln, Canterbury, New Zealand.

Wu, L. Y. 1974. *Paratylenchus robustus* n. sp. (Paratylenchinae: Nematoda) from forest soil in Ontario. Canadian Journal of Zoology 52:1423–1452.

Yu, Q., Popovic, V., and Gu, J. 2014. *Deladenus valveus* n. sp. (Nematoda: Neotylenchidae) in dunnage wood from South Korea intercepted in Ningbo, China. Journal of Nematology 46:119–124.

Zeng, Y. S., Ye, W., Wang, X., Du, Z., and Giblin-Davis, R. M. 2013. *Schistonchus fistulosus* n. sp. (Nematoda: Aphelenchoididae) associated with *Ficus fistulosa* in China. Nematology 15:879–890.