## Parasitism of Rotylenchulus reniformis onSoybean Root Rhizobium Nodules in Venezuela

J. A. Meredith,<sup>1</sup> R. N. Inserra,<sup>2</sup> D. Monzon de Fernandez<sup>1</sup>

Journal of Nematology 15(2):211-214. 1983.

Rotylenchulus reniformis Linford and Oliveira parasitizes soybean Glycine max (L.) Merr. in Venezuela and is often associated with Pratylenchus brachyurus. In the tropical climate of Venezuela, R. reniformis reproduces rapidly on soybean roots; the nematode also infects the Rhizobium root nodules. Since there is no report of R. reniformis infection of soybean Rhizobium root nodules, a histological study was made to determine the effect of parasitism of nodules by a Venezuelan population of R. reniformis.

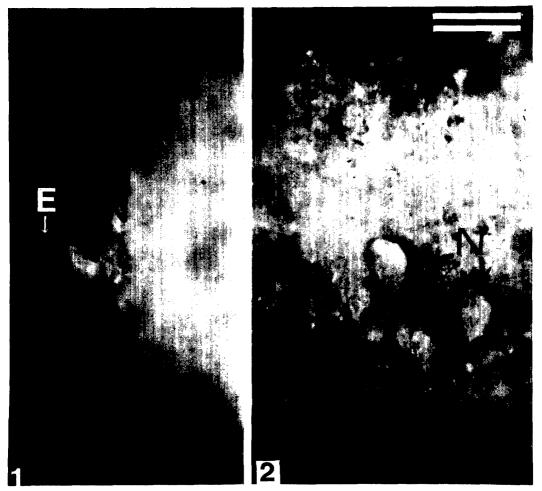
Soybean cv. Jupiter roots infected with R. reniformis were collected from a field in Portuguesa State. Roots were washed, and some were stained with acid fuchsin in hot lactophenol and examined microscopically. Other roots with *Rhizobium* nodules infected by mature nematode females were fixed in FAA (23% formalin, 3% acetic acid, 30% ethyl alcohol, 44% distilled water), dehydrated in TBA (tertiary butyl alcohol), and embedded in paraffin. Cross sections 10–15  $\mu$ m thick were stained with safranin fast-green, mounted in Dummar xylene (3) and observed with a compound microscope.

Rotylenchulus reniformis parasitized both the roots and the Rhizobium nodules of Jupiter soybean. The swollen posterior portion of the females and egg masses protruded from the root nodule surface (Fig. 1). The female nematodes were observed most frequently in the portion of the bacterial nodule in contact with the root, but

Received for publication 26 April 1982.

<sup>&</sup>lt;sup>1</sup>Professor of Nematology and Graduate Student, respectively, Universidad Central de Venezuela, Facultad de Agronomia, Instituto de Zoologia Agricola, Apartado 4579, Maracay, Aragua 2101, Venezuela.

<sup>&</sup>lt;sup>2</sup>Nematologist, Istituto Nematologia Agraria, CNR, Bari, Italy. Present address: Crops Research Laboratory, Utah State University, UMC 63, Logan, UT 84322.



Figs. 1-2. Rotylenchus reniformis on soybean (Glycine max) root nodule. Scale bars = 480  $\mu$ m. 1) Mature female (N) with the body posterior portion protruding from the root nodule surface; E = egg mass. 2) Mature females (N) in the interstitial fissures of ruptured nodule surface.

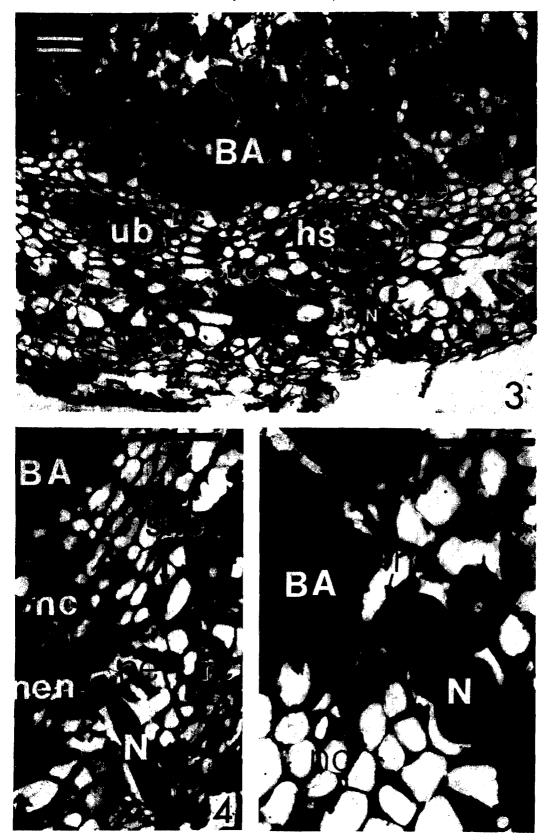
any point of the root nodule surface was susceptible to the nematode penetration. Some females with egg masses were located in the interstitial fissures of the ruptured nodule surface (Fig. 2). The number of mature females with egg masses averaged from one to three per nodule.

Sections of nematode infected soybean nodules showed that R. reniformis penetrated into the epidermis, root cortex, and nodule cortex and established a permanent feeding site in the nodule endodermis and the bundle endodermis (Figs. 3,4). The most common reaction to the nematode in the nodule vascular bundles was safranin positive stained bundle endodermal cells and accentuated hypertrophy of the bundle sheath cells compared with the bundle

\*\*

Figs. 3-5. Histological alterations caused by *Rotylenchulus reniformis* in soybean root nodules. Scale bars = 71  $\mu$ m. 3) Cross section showing a nematode (N) penetrated into the root cortex (rc) and feeding in the bundle endodermis (be) of a vascular bundle. Note the hypertrophy of the bundle sheath cells (hs) compared to those of uninfected bundle (ub); BA = bacteroid region, nc = nodule cortex, sne = suberized nodule endodermis. 4) Cross section showing a nematode (N) feeding in the nodule endodermis (nen). Note the necrosis (ne) of nodule endodermal (nen) and nodule cortical (nc) cells at the nematode feeding in a nodule cortical (nc) cell adjacent to the bacteroid region (BA). Necrosis (ne) is evident in the nodule cortical cell at the feeding site.

Rotylenchulus on Soybean Nodules: Meredith et al. 213



sheath cells of healthy vascular bundles (Fig. 3). Accentuated necrosis (safranin stained tissue) was also observed in the nodule endodermal cells in direct contact with the nematode head (Fig. 4). In some cases, nematode specimens penetrated the root and nodule cortex and reached the innermost layer of nodule cortical cells free of rhizobial infections and adjacent to the bacteroid region (Fig. 5). Considering that all the sectioned nodules were infected with mature females only, this would indicate that R. reniformis was able to develop by feeding in the nodule cortical cells of soybean. However, there was no evidence of nematode penetration into the bacteroid region.

In addition to *R. reniformis, Meloi*dogyne incognita is able to reproduce in soybean root nodules (2). But soybean root nodules are unfavorable for the development of *Heterodera glycines* (1). *H. glycines* and *M. incognita* both attack the vascular bundles, damaging the vascular elements by inducing syncytium and giant cells formation, respectively (1,2), whereas *R. reni*formis caused alterations of the periphery of vascular bundles, damaging the bundle endodermis and inducing hypertrophy of bundle sheath cells.

The adverse effect of R reniformis on Rhizobium nodule production has been reported in cowpea (4), but from our observations on soybean root nodules there was no evidence of bacteroid disruption by R. reniformis. R. reniformis-infected soybean root nodules did not differ in size and shape from healthy ones. The necrosis induced by the nematode in the endodermal layers and also in the nodule and root cortex may predispose the nodular tissues to the infections of other pathogens and to their consequent premature breakdown.

## LITERATURE CITED

1. Barker, K. R., D. Huisingh, and S. A. Johnston. 1972. Antagonist interaction between Heterodera glycines and Rhizobium japonicum on soybean. Phytopathology 62:1201-1205.

2. Barker, K. R., and R. S. Hussey. 1976. Histopathology of nodular tissues of legumes infected with certain nematodes. Phytopathology 66:851-855.

3. Johansen, D. A. 1940. Plant microtechnique. New York: McGraw-Hill.

4. Taha, A. H. Y., and A. S. Kassab. 1980. Interrelations between Meloidogyne javanica, Rotylenchulus reniformis, and Rhizobium sp. on Vigna sinensis. J. Nematol. 12:57-62.