

Survey of *Meloidogyne* spp. in Tomato Production Fields of Baix Llobregat County, Spain

F. J. SORRIBAS¹ AND S. VERDEJO-LUCAS²

Abstract: A survey was conducted to determine the frequency and abundance of *Meloidogyne* spp. in tomato production sites located in Baix Llobregat County, Barcelona, Spain. Forty-five sites were sampled before planting and at harvest from February to October, 1991. *Meloidogyne* spp. occurred in 49% of the sites sampled. Preplant population densities ranged from 10 to 220 (\bar{x} = 110) juveniles/250 cm³ soil, and final population densities ranged from 20 to 1,530 (\bar{x} = 410) juveniles/250 cm³ soil. Final population densities were higher in open fields than in field greenhouses, but initial population densities were higher in greenhouses than in fields. *Meloidogyne incognita*, *M. javanica*, and *M. arenaria* were found in this survey. *Meloidogyne* populations that reproduced on *M. incognita*-resistant tomato cultivars in the field sites did not circumvent the *Mi* gene resistance in greenhouse tests.

Key words: greenhouse, *Lycopersicon esculentum*, *Meloidogyne*, nematode, resistance, root-knot nematode, Spain, survey, susceptibility, tomato.

Meloidogyne is the most important genus of plant-parasitic nematodes that affect vegetable crops in Spain. Recent reports have shown an increased concern about the rapid spread and wide distribution of *Meloidogyne* spp. throughout the country (1-3,8,17,19,20,26,28). In the 1960s, *Meloidogyne* was limited to a few localities and infestation levels were low (9,10,15). More recently, however, *Meloidogyne* has become a major nematode problem in some areas of the country, and it has displaced other economically important nematodes (17,29). Information on the distribution and abundance of *Meloidogyne* spp. in vegetable crops in Catalonia (north-east Spain) was lacking, although the nematode had been observed on tomato and other crops in this region (15,18).

Baix Llobregat County is the third-largest regional producer of fresh market vegetables in Catalonia (27). A multiple-cropping system is practiced, with up to 15 different vegetable crops grown in rotation. Tomato (*Lycopersicon esculentum*), cucumber (*Cucumis sativus*), and lettuce (*Lactuca sativa*) are the vegetables produced most frequently in field greenhouses. The

types of vegetables grown in open fields are more numerous and diverse, but tomato and lettuce are the most important annual crops. Tomato is cultivated as an early crop from February to June in field greenhouses, and from April to October as the main crop in open fields. Farms are family-operated, and the average size is 2,500 m².

The objective of this study was to survey tomato production sites for the presence, distribution, and abundance of *Meloidogyne* spp.

MATERIALS AND METHODS

During the 1991 growing season, 45 fields representing 11 ha were sampled for *Meloidogyne*. Sites were located in three municipalities, Gavà, Sant Boi, and Viladecans in Baix Llobregat County, Barcelona, Spain. Sites to be planted to tomato were identified through direct grower contacts. Composite soil and root samples were collected from 16 field greenhouses, and 29 open fields from February to October. Samples were taken from the top 30 cm soil layer with a soil auger (2.5-cm-d × 50 cm deep) preplant (initial population) and at harvest (final population). Samples were sieved to separate roots from soil, and nematodes in 250-cm³ soil subsamples were extracted by the centrifugal-flotation method (14). At harvest, *Meloidogyne* eggs were collected from roots blended in a 0.5% NaOCl solution for 10 minutes (13).

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¹ Escuela Superior de Agricultura de Barcelona, Conde Urgel 187, 08036 Barcelona, Spain.

² Departamento de Patología Vegetal, IRTA. Crta. de Cabriels s/n, 08348 Cabriels, Barcelona, Spain. Author to whom correspondence should be addressed.

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Soil texture (Bouyoucos densimeter and USDA classification) and pH (1:2.5, w/v in water) were determined for each site. Data on previous crops, chemical treatments, and agricultural production practices were obtained from interviews with growers (Table 1).

For identification of species, soil samples that contained *Meloidogyne* second-stage juveniles (J2) were potted and a susceptible tomato seedling (cv. Precodor) was transplanted into each pot. *Meloidogyne* species were identified based on the perineal patterns of the females (7) and polymerase chain reaction (PCR) analysis (5). The DNA was isolated from J2 cultured monoxenically from single egg masses (30) according to the procedure described by Cenis (5).

The reactions of four *M. incognita*-resistant cultivars and one susceptible tomato cultivar to two isolates of *M. incognita* and one of *M. javanica* were tested under greenhouse conditions. The populations were cultured from single egg masses (30) on potted Precodor tomato plants. The *M. incognita*-resistant tomato cultivars tested were Carmelo, Carpy, Mina, and Rambo. The susceptible cultivar Precodor was also tested for comparison. Seedlings of each cultivar were transplanted 4 weeks after germination to pots (500 cm³ soil capacity) containing steam-sterilized sand. A suspension of 500 eggs of each nematode isolate was pipetted in two holes made in the soil near the plant. Each isolate-cultivar combination was replicated nine times, and plants were maintained in a greenhouse. Plants were watered when needed and fertilized with a slow-release fertilizer (15N + 10P + 12K + 2MgO + microelements). Dry top weight, fresh root weight, and numbers of eggs per plant were determined 7 weeks after infestation. Eggs were extracted from roots in a 0.5% NaOCl solution for 10 minutes (13). Numbers of eggs per plant included egg shells and unhatched eggs. Numbers of eggs per plant were transformed ($\log [x + 1]$) and subjected to analysis of variance. Means were compared by the Tukey test ($P = 0.05$).

RESULTS AND DISCUSSION

Root-knot nematodes were detected in 22 out of 45 sites sampled (12 field greenhouses and 10 open fields). The nematodes were found in preplant soil samples from 11 sites (24%), and in soil or root samples from 11 additional sites at harvest (Table 2). Preplant population densities ranged from 10 to 220 ($\bar{x} = 110 \pm 85$) J2/250 cm³ soil (Table 2). Final population densities in soil ranged from 20 to 1,530 ($\bar{x} = 410 \pm 60$) J2/250 cm³ soil. The number of eggs per g root ranged from 110 to 26,810 ($\bar{x} = 7,570 \pm 460$) (Table 2). In general, final nematode population densities were greater in open fields than in field greenhouses. In contrast, *Meloidogyne* was detected more frequently at the beginning of the season in field greenhouses than in open fields. *Meloidogyne* was found at the end of the season in 67% of the greenhouse sites that had been fumigated just before the initial sampling (Table 2). The use of soil fumigants probably reduced the increase of nematode population densities in these sites.

Numbers of *Meloidogyne* J2 were below detectable level before planting the crop in 76% of the sites, but increased after growing a single crop of tomato. This increase could be explained in some cases by the vertical migration of the nematodes from deeper to upper soil layers in the presence of the host plant (22). Also, the nematodes may have escaped detection in preplant samples as eggs within egg masses (11,12). The efficiency of our extraction method (14) is not satisfactory for separating nematode eggs from the soil.

There was no correlation between the numbers of J2 in the soil and soil texture or pH (range 7.3 to 8.4). Silty loam soils were dominant in this survey, and *Meloidogyne* was present in 45% of the sites with this soil texture. Other soil textures where *Meloidogyne* were present included sandy loam (27%), loamy sand (23%), sand (9%), and loam (5%).

The *Meloidogyne* species identified were *M. incognita* in sites 6, 30, 32, 35; *M. ja-*

TABLE 1. Soil characteristics and history of tomato production sites where *Meloidogyne* spp. were detected in Baix Llobregat County, Barcelona, Spain.

Site		Locality	Area (m ²)	Soil texture	pH	Preplant soil treatment and year	Previous crops‡	Tomato cultivar 1991
Number	Cultivation†							
1	Greenhouse	Sant Boi	2,700	Silty loam	7.73	Metham-Na + DD, 1991	T, Cu, F	Mereto
4	Greenhouse	Viladecans	1,750	Loam	7.86	Methyl bromide, 1991	P, Cf	Precodor
6	Greenhouse	Viladecans	3,000	Silty loam	7.55	Methyl bromide, 1990	T, L	Precodor
13	Greenhouse	Sant Boi	630	Silty loam	7.32	Methyl bromide, 1991	T, L	Mereto
12	Greenhouse	Sant Boi	630	Silty loam	7.79	Methyl bromide, 1991	T, L	Mereto
15	Greenhouse	Viladecans	1,500	Silty loam	7.59	Methyl bromide, 1991	T, L	Precodor
2	Greenhouse	Sant Boi	850	Silty loam	7.56	Metham-Na, 1991	T, L	Mereto
10	Greenhouse	Viladecans	1,000	Sandy loam	7.65	Methyl bromide, 1991	P, T	Precodor
14	Greenhouse	Viladecans	4,000	Silty loam	7.77	Methyl bromide, 1991	T, P, L	Luxor
5	Greenhouse	Viladecans	2,500	Silty loam	7.54	Methyl bromide, 1990	T, P, L	Carmelo
16	Greenhouse	Gavà	1,600	Sandy loam	8.14	DD + methyl isothiocyanate, 1991	T, L	Carmelo
3	Greenhouse	Gavà	1,300	Loamy sand	8.04	Untreated	P, R	Carmelo
30	Field	Viladecans	7,000	Sandy loam	8.40	Methyl bromide, 1989	L, S	Royesta
36	Field	Viladecans	2,500	Sand	8.11	Untreated	T, R	Monix
32	Field	Viladecans	2,500	Loamy sand	8.20	Untreated	R	Dario
34	Field	Gavà	2,000	Loamy sand	8.28	Untreated	T, R	Royesta
35	Field	Viladecans	2,000	Sandy loam	8.19	Untreated	T, R, Ce	Monix
39	Field	Gavà	2,500	Sandy loam	8.20	Untreated	L	Riogrande
27	Field	Gavà	4,000	Sand	8.06	Untreated	Po, Cr	Riogrande/ Iberia
28	Field	Gavà	2,500	Loamy sand	7.87	Untreated	Fallow	Iberia
26	Field	Viladecans	5,000	Sandy loam	7.99	Untreated	L, Ce	Cobra
29	Field	Viladecans	2,500	Sandy loam	7.60	Untreated	S, Ca, Cf	Cobra

† "Greenhouse" refers to production in a field site with a temporary covering used to extend the growing season. "Field" refers to production in uncovered sites.

‡ C: cabbage, Ce: celery, Cf: cauliflower, Cr: curly leaf lettuce, Cu: cucumber, F: french bean, E: endive, L: lettuce, P: pepper, Po: potato, R: radish, S: spinach, T: tomato.

TABLE 2. *Meloidogyne* spp. population densities in tomato production sites in Baix Llobregat County, Barcelona, Spain.

Site		Cultivar	Preplant soil treatment	Nematodes/250 cm ³ soil		Number eggs/g root	
Cultivation	Number			Preplant	Harvest		
Greenhouse	1	Mereto (S) [†]	+‡	209	920	276	
	4	Precodor (S)	+	140	29	519	
	6	Precodor (S)	+	196	284	427	
	13	Mereto (S)	+	42	180	0	
	12	Mereto (S)	+	0	54	0	
	15	Precodor (S)	+	0	1,235	0	
	2	Mereto (S)	+	209	0	0	
	10	Precodor (S)	+	144	0	0	
	14	Luxor (R)	+	16	0	0	
	5	Carmelo (R)	+	0	0	108	
	16	Carmelo (R)	+	0	19	0	
	3	Carmelo (R)	-	218	0	0	
	Field	30	Royesta (R)	+	0	44	0
		36	Monix (R)	-	10	0	0
		32	Dario (R)	-	0	504	13,551
		34	Royesta (R)	-	0	42	9,756
35		Monix (R)	-	0	0	1,578	
39		Riogrande (S)	-	0	260	0	
27		Riogrande (S)	-	0	70	15,220	
28		Iberia (S)	-	0	494	7,483	
26		Cobra (S)	-	15	1,530	26,814	
29		Cobra (S)	-	26	510	0	

[†] S = susceptible; R = resistant cultivar to *M. incognita*.

[‡] + = Methyl bromide applied prior to initial sampling in 1991 except for sites 5 and 6, and 30 that were fumigated in 1990 and 1989, respectively. Sites 1, 2, and 16 were treated with DD + Na methylthiocarbamate, Na-methylthiocarbamate and DD + methylisothiocyanate, respectively; - = untreated.

vanica in sites 2, 6, 28, 34; and *M. arenaria* in sites 1, 10, 27, 29. Isolation and identification of *Meloidogyne* species from sites where population densities were low was not achieved.

Although resistance to *M. incognita* is available in commercial tomato cultivars, the results of this survey indicated that different species of *Meloidogyne* can increase to high numbers on *M. incognita*-resistant tomatoes in the field. Thus, *M. incognita*-resistant tomato cultivars, such as Royesta and Monix, supported large population densities of *Meloidogyne* in sites 34 and 35, but not in sites 30 and 36, respectively (Tables 1 and 2). Large numbers of eggs were also found on roots of the *M. incognita*-resistant tomato cv. Dario in site 32. Isolates that reproduced well on *M. incognita*-resistant tomatoes (*M. incognita* in sites 32 and 35, and *M. javanica* in site 34) were selected for the greenhouse test because

they were suspected to be resistance-breaking populations; however, the results of the greenhouse test showed otherwise. The *M. incognita* isolates found in sites 32 and 35 did not reproduce on the *M. Meloidogyne*-resistant cultivars tested, whereas the *M. javanica* isolate from site 34 did reproduce; nonetheless, *M. javanica* reproduction was lower on the resistant cultivars than on the susceptible one (Table 3). Numbers of eggs did not differ ($P = 0.05$) among the three isolates tested on the *M. incognita*-susceptible cv. Precodor, although the *M. javanica* isolate produced 3.4 and 2.6 times more eggs than the *M. incognita* isolates from sites 32 and 35, respectively. The use of *M. incognita*-resistant cultivars may cause a shift in root-knot populations to other species as it has been shown in tobacco fields naturally infested by *M. incognita* and *M. javanica* (6,16). However, the *M. incognita* isolates from

TABLE 3. Reproduction of *Meloidogyne incognita* isolates from two sites and *M. javanica* from one site on four *M. incognita*-resistant cultivars and one susceptible cultivar of tomato 49 days after inoculation with 500 nematode eggs.

Cultivar	Final population/initial population		
	<i>M. incognita</i>		<i>M. javanica</i>
	Site 32	Site 35	Site 34
Resistant			
Carmelo	0.1† b	1.9 b	3.8 b
Carpy	0.1 b	0.1 b	4.8 b
Mina	0.2 b	0.8 b	5.6 b
Rambo	0.6 b	0.8 b	17.8 b
Susceptible			
Precodor	24.7 a	33.0 a	85.2 a
Mean	4.9	7.5	23.4
CV	79.6	72.8	42.1

† Values are means of nine replications. Means followed by the same letter within the same column are not different ($P = 0.05$) according to the Tukey test.

sites 32 and 35 were the predominant species because they were identified from soil and root samples collected at harvest (Table 2). The greenhouse test showed genetic variation in the tomato cultivars (F_1 hybrids) tested. Thus, most plants (6–7 plants) of cvs. Carmelo, Carpy, and Mina were highly resistant to *M. incognita* from site 32, and four of nine plants also reacted as highly resistant to *M. incognita* from site 35. *Meloidogyne javanica* from site 34, however, completed a life cycle and produced eggs on all plants of all cultivars tested. Many *M. incognita*-resistant tomato cultivars support nematode production, and some cultivars have been reported as susceptible to *M. incognita* (31), *M. javanica* (24), and *M. arenaria* (23).

The *M. javanica* from site 34 had a high Pf/Pi on the *M. incognita*-susceptible cv. Precodor, and also parasitized *M. incognita*-resistant tomato cultivars both under natural and experimental conditions.

The results of this survey confirm other reports on the widespread distribution of *Meloidogyne*, its high frequency of occurrence, and its potential as a problem for vegetable production in Spain (1,2,4,28). Species identification is important and

should be considered when selecting the cultivar or the crop to be planted.

LITERATURE CITED

1. Andreu Lopez, M., T. Salmeron Parra, V. Martinez Sierra, and A. Tobar Jimenez. 1986. Nematodos fitoparásitos de los cultivos hortícolas de Trigueros (Huelva). Boletín de Sanidad Vegetal Plagas 12:319–322.
2. Bello, A., J. L. Cenis, and J. Fresno. 1989. Nematodos formadores de nodulos (*Meloidogyne* spp.) y su relación con el manejo de los suelos en ambientes de clima mediterráneo continental. Resúmenes del V Congreso Nacional de Fitopatología, Badajoz, Spain.
3. Cenis, J. L. 1985. Control del nematodo *Meloidogyne javanica* (Treub) Chit. mediante calor solar (Solarización). Anales del Inia: Servicio Agrícola, Suplemental vol. 28:121–130.
4. Cenis, J. L. 1987. Occurrence of the root-knot nematode, *Meloidogyne* spp. in the southeast of Spain. Proceedings of the 7th Congress of the Mediterranean Phytopathological Union, Granada, Spain.
5. Cenis, J. L. 1993. Identification of four major *Meloidogyne* spp. by random amplified polymorphic DNA (RAPD-PCR). Phytopathology 83:76–80.
6. Csinos, A. S., A. W. Johnson, and A. M. Golden. 1986. Metalaxyl and fenamiphos applied through irrigation water to control black shank-root knot complex on tobacco. Plant Disease 70:210–213.
7. Eisenback, J. D., H. Hirschmann, J. N. Sasser, and A. C. Triantaphyllou. 1981. A guide to the four most common species of root-knot nematodes (*Meloidogyne* spp.), with a pictorial key. Raleigh: North Carolina State University Graphics.
8. García Morato, M. 1987. Principales enfermedades y fisiopatías que en la Comunidad Valenciana presentan las especies hortícolas más cultivadas. Phytoma 30:62–66.
9. Gomez Barcina, A., and F. Jimenez-Millan. 1967. Primeros estudios nematológicos de la zona costera granadino-malagueña. Anales de Edafología y Agrobiología 26:585–600.
10. Guevara Pozo, D., and A. Tobar Jimenez. 1964. Nematodes parásitos de la vega de Granada. Revista Ibérica de Parasitología 24:3–42.
11. Guiran, G. de. 1979. A necessary diapause in root-knot nematodes. Observations on its distribution and inheritance in *Meloidogyne incognita*. Revue de Nématologie 2:223–231.
12. Guiran, G. de, and M. A. Villamin. 1980. Spécificité de la diapause embryonnaire des oeufs de *Meloidogyne incognita* (Nematoda). Revue de Nématologie 3:115–121.
13. Hussey, R. S., and K. R. Barker. 1973. A comparison of methods of collecting inocula of *Meloidogyne* spp., including a new technique. Plant Disease Reporter 57:1025–1028.
14. Jenkins, W. R. 1964. A rapid centrifugal-flotation technique for separating nematodes from soil. Plant Disease Reporter 48:692.
15. Jimenez Millan, F., M. Arias, A. Bello, and

- J. M. Lopez Pedregal. 1965. Catálogo de los nematodos fitoparásitos y peri-radicales encontrados en España. Boletín Real Sociedad Española de Historia Natural (Biología) 63:47–104.
16. Johnson, A. W., A. S. Csinos, A. M. Golden, and N. C. Glaze. 1992. Chemigation for control of black shank-root-knot complex and weeds in tobacco. Supplement to the *Journal of Nematology* 24:648–655.
17. Lopez Robles, J. 1991. Los problemas planteados por nematodos y su control en el cultivo de la remolacha. *Phytoma* 8:73–77.
18. Marull, J., J. Pinochet, S. Verdejo, and A. Soler. 1991. Reaction of *Prunus* rootstocks to *Meloidogyne incognita* and *M. arenaria* in Spain. Supplement to the *Journal of Nematology* 23:564–569.
19. Millan de Aguirre, J. R. 1989. Especies del género *Meloidogyne* presentes en los cultivos de la C.A. Vasca. Pp. 164–167 in J. Del Moral de la Vega, ed. *Estudios de Fitopatología*. Badajoz. Consejería de Agricultura, Industria y Comercio. Junta de Extremadura.
20. Moral de la Vega, J., and F. Martínez Aljama. 1985. Estudio sobre la biología y control de diversas especies de nematodos fitoparásitos en cultivo de maíz. *Anales del Inia: Servicio Agrícola*, Suplemental vol. 28:73–89.
21. Noe, J. P. 1992. Variability among populations of *Meloidogyne arenaria*. *Journal of Nematology* 24:404–414.
22. Prot, J. C. 1980. Migration of plant-parasitic nematodes towards plant roots. *Revue de Nématologie* 3:305–318.
23. Prot, J. C. 1984. A naturally occurring resistance breaking biotype of *Meloidogyne arenaria* on tomato. Reproduction and pathogenicity on tomato cultivars Roma and Rossol. *Revue de Nématologie* 7:23–28.
24. Roberts, P. A., and I. J. Thomason. 1986. Variability in reproduction of isolates of *Meloidogyne incognita* and *M. javanica* on resistant tomato genotypes. *Plant Disease* 70:547–551.
25. Roberts, P. A., and I. J. Thomason. 1989. A review of variability in four *Meloidogyne* spp. measured by reproduction on several hosts including *Lycopersicon*. *Agricultural Zoology Reviews* 3:225–252.
26. Rodríguez Rodríguez, R. 1984. El género *Meloidogyne* en Canarias. I. Rastreo geográfico preliminar y especies encontradas. *Xoba* 4:41–51.
27. Rosès, J. 1989. L'economia del Baix Llobregat. Creixement i desequilibris. Caixa d'Estalvis de Catalunya. Barcelona, Spain.
28. Salmeron, T., and T. Cabello. 1989. Incidencia de *Meloidogyne incognita* en cultivos de tabaco de la Vega de Granada (SE. de España). *Boletín de Sanidad Vegetal Plagas* 4:307–314.
29. Tobar Jiménez, A., T. Salmerón Parra, and V. Martínez Sierra. 1984. La utilización de suelos de regadío en Granada, valorada por sus niveles de población nematológica. *Boletín de Sanidad Vegetal Plagas* 10:69–76.
30. Verdejo, S., R. Mankau, and B. A. Jaffee. 1988. Reproduction of *Meloidogyne javanica* on plant roots genetically transformed by *Agrobacterium rhizogenes*. *Journal of Nematology* 20:599–604.
31. Viglierchio, D. R. 1978. Resistant host responses to ten California populations of *Meloidogyne incognita*. *Journal of Nematology* 10:224–227.