# INFLUENCE OF VARIOUS COMMERCIAL FLORAL PRESERVATIVES AND 8-HYDROXYQUINOLINE CITRATE PLUS SUCROSE ON DEVELOPMENT AND LASTING ABILITY OF FLOWER BUDS OF SEVERAL CHRYSANTHEMUM CULTIVARS

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### ABSTRACT

Several commercial floral preservatives and 8-hydroxyquinoline citrate (8-HQC) plus sucrose were evaluated as opening solutions for standard chrysanthemum flower buds. Preservatives which produced the largest increases in stem fresh weight also produced the largest flowers with the greatest longevity. Several commercial preservatives were found to be as effective as 8-HQC plus sucrose for opening the flower buds.

Spray chrysanthemum cultivars "Shasta," "Iceberg," and "Beauregard" were harvested at the "tight" and "opened" bud stage and held in an opening solution of 200 ppm 8-HQC plus 2% sucrose. Flower buds held in the opening solution gained more fresh weight and increased in diameter more than flower buds held in water.

### INTRODUCTION

Marousky (13) recently reported that standard chrysanthemums could be harvested in the bud stage (45-55 mm diameter) and successfully opened off the plant in 8-hydroxyquinoline citrate (8-HQC) and sucrose solutions. The technique appeared sufficiently promising to warrant further exploration for commercial application. The above tests used large flowering incurve and spider cultivars. Information on how to handle spray type (pompon) cultivars would be helpful to Florida flower growers.

Commercial floral preservatives might be used as an adjunct or as a substitute for quinoline salts and sucrose in opening bud-cut chrysanthemums. Floral preservatives have been used successfully to open bud-cut carnations (2, 4, 5) and chrysanthemums (3, 15). Waters (15) showed that 'Iceberg' chrysanthemum flowers harvested at the tight-bud stage increased in diameter and fresh weight when held in Everbloom. He also reported that Everbloom had a deleterious effect on foliage.

Quinoline salts and sucrose solutions have been reported to have beneficial effects on many flowers (7, 8, 9, 11, 14). Possible functions and mechanism of quinoline salt and sucrose have been elucidated elsewhere (1, 10, 12, 6, 14).

This paper reports the influence of commercial floral preservatives on bud-cut flowers of incurved chrysanthemum cultivars and the influence of 8-HQC and sucrose on flower bud opening of spray type (pompon) chrysanthemums.

#### METHODS AND MATERIALS

Experiment 1. On November 15, 1963 commercially grown 'Albatross' standard chrysanthemum flowers were harvested as buds when 50-55 mm in diameter. Flowers were immediately transferred to the laboratory and stems were trimmed to 18 inches in length. Foliage was removed from the lower third of the stem. Stems were placed in quart class jars containing water or various floral preservatives listed in Tables 1 and 2. Distilled water was used for preparing all solutions.

Flower diameter and stem fresh weight were determined initially and at 4 and 7 days. Vaselife was measured from the day flowers were fully opened until the day petals lost turgidity or decorative value or both. Foliage condition was not used as a criterion for vase-life. Six replications were used for each treatment with 1 flower constituting a single replication.

Flowers were held in treatment solutions in a laboratory maintained at  $74^{\circ}$  F  $\pm 2^{\circ}$  and at a continuous light intensity of 200 ft-c supplied by fluorescent tubes. During the experiment relative humidity ranged from 50 to 75%.

Experiment 2. 'Mrs. Roy' standard chrysanthemums were grown in a greenhouse and harvested on December 17, 1968 when flower buds

References to specific commercial products do not constitute an endorscement.

were 50-55 mm in diameter. Stems were placed in water and floral preservative solutions listed in Tables 3 and 4. Handling procedures, laboratory environment, and data collection were similar to those in the previous experiment.

Experiment 3. Chrysanthemum stems of 'Shasta', 'Iceberg', and 'Beauregard', cultivars of the spray category, were harvested from commercial plantings on May 18, 1969. Stems bearing buds that were "tight" or "open," 20-30 mm and 40-50 mm in diameter, respectively, were harvested. Stems were brought to the laboratory, trimmed to 30-inch lengths, and foliage was removed from the lower third of the stem. Stems bearing flowers from each stage of development were placed in plastic containers (15 in. high by 10 in diameter) containing water or 200 ppm 8-HQC + 2% sucrose.

Flower diameter and stem fresh weight were determined initially and at 3, 7, and 10 days. For flower diameter measurements, a single flower was selected randomly from each stem. There were 3 replications per treatment with a single stem considered as 1 replication.

Stems were held in the laboratory as in the previous experiments.

All data were treated by analysis of variance and statistical differences between means determined by Duncan's Multipel Range Test.

#### RESULTS

Experiment 1.—Flowers held in 8-HQC-sucrose or Everbloom were larger than flowers held in other preservatives or water (Table 1). Flowers held in F M Super or Roselife for 7 days were larger than flowers held in water or Bloomlife. Flowers held in water or Bloomlife were flat in appearance while flowers held in other preservatives were fully developed and globular in form after 6 days. All flowers, except those held in water or Bloomlife, would have been commercially acceptable.

Stems held in 8-HQC+sucrose or Everbloom weighed more after 4 and 7 days than stems held in other preservatives or water (Table 2). Stems held in Roselife or F M Super weighed more than stems held in water or Bloomlife. Stems held in Roselife for 7 days were heavier than stems held in F M Super. Stems held in water or Bloomlife did not increase in weight after 3 days; whereas, stems held in 8-HQC+sucrose, Everbloom, Roselife, F M Super continued to increase in weight for 7 days. Table 1. Diameters of 'Albatross' chrysanthemum flowers harvested as buds and held in solutions of various floral preservatives for 7 days.<sup>1</sup>

Floral		Diameter after		
Preservatives	Concentration	4 days	7 days	
		99.	inter inter	
Control (water)		93 c <sup>2</sup>	96 c <sup>3</sup>	
Bloomlife	27,	96 c	90 c	
F H Super	8 ml/liter	96 c	107 Б	
Roselife	32	105 b	114 þ	
8-HQC+sucrose	200 ppm+27,	116 a	121 8	
Everbloom	37.	116 a	124 a	

<sup>1</sup>Flower buds harvested when 50-55 mm in diameter

<sup>2</sup>Means within a column followed by different letters differ significantly at the 12 level.

<sup>3</sup>Heans within a column followed by different letters differ significantly at the 32 level.

Vase-life paralleled the changes that occurred in fresh weight and flower size (Table 2). The largest flowers with the heaviest stems (8-HQC +sucrose and Everbloom treatments) lasted the longest, whereas, the smallest flowers (control and Bloomlife treatments) had short vase-life. Foliage on stems held in all preservatives became chlorotic after 15 days but the most severe chlorosis appeared on foliage from stems held in Roselife. Foliage on stems held in water did not become chlorotic.

*Experiment 2.*—'Mrs. Roy' chrysanthemum flowers responded to various floral preservatives in a manner similar to that of 'Albatross' flowers in the previous experiments. Flowers held in preservatives were larger than flowers held in

<u>Table 2</u>. Fresh weight and vasc-life of 'Albatross' chrysanthemmum flowers harvested as buds and held in solutions of various floral preservatives.<sup>1</sup>

Floral		Changes in weight after <sup>2</sup>		Vase-life
preservatives	Concentration	4 days	7 days	
		7.	7	days
Control (water)	-	123.1 c <sup>3</sup>	120.7 d <sup>3</sup>	5.2 d <sup>4</sup>
Bloomlife	22	125.2 c	117.7 d	2.2 e
F M Super	8 ml/liter	135.6 Б	140.4 c	9.0 c
Roselife	37.	135.2 Ь	156.6 b	15.0 b
8-HQC+sucrose	200ppn+2%	144.4 a	168.9 4	18.6 a
Everbloom	3%	148.4 a	179.2 a	16.2 ab

<sup>1</sup>Flower buds harvested when 50-55 mm in diameter

<sup>2</sup>Weights are expressed as percentage values of initial weight. Initial fresh weight equals 100%.

<sup>3</sup>Means within a column followed by different letters differ significantly at the 5% level.

<sup>4</sup>Means within a column followed by different letters differ significantly at the 1% level. Vase-life was recorded from the day flowers were determined to be fully open (6 days).

<u>Table 3.</u> Diameters of 'Mrs. Roy' chrysanthesum flowers harvested as buds and hold in solutions of various floral preservatives.<sup>1</sup>

Floral		Diameter after		
preservatives	Concentration	3 days	7 days_	
		<b>8</b>		
Control (water)	•	80 c <sup>2</sup>	- 2,3	
Roselife	21.	102 b	115 cd	
Roselife	47.	105 6	117 bcd	
7 H Super	2 m1/1	108 5	. 3	
9 M Super + sucrose	2 m1/1+27.	111 ab	114 cd	
F M Super	4 m1/1	101 Ъ	112 d	
Y M Super + sucross	4 m1/1+27.	112 ab	116 bcd	
Everbloom	27,	111 ab	123 ab	
Evesbloom	47.	111 ab	120 abo	
8-HQC+sucress	200ppm+27	120 a	126 a	

<sup>1</sup>Flower buds harvested when 50-55 mm in diameter.

<sup>2</sup>Neans within a column followed by different letters differ significantly at the 1% level.

<sup>3</sup>Flowers from control and F H Super (2 ml/1) wilted after 6-7 days and were not included in analysis.

water (Table 3). Flowers held in water or F M Super (2 ml/1) wilted before they were fully open. No differences in diameter were evident for flowers held in 2 and 4% Roselife, 2 ml F M Super plus 2% sucrose, and 4 ml F M Super with and without 2% sucrose. Flowers held in 8+HQC-sucrose were larger than flowers held in other preservatives except 2 and 4% Everbloom. All stems held in solutions continued to in-

All stems held in solutions continued to h

<u>Table 4.</u> Fresh weight and vase-life of 'Mrs. Roy' chrysanthemum flowers harvested as buds and held in solutions of various floral preservatives.<sup>1</sup>

Floral		Fresh weight after		Vase
preservatives	Concentration	3 days	7 days	life
		7.	7.	days
Control (water)	•	115.9 cd <sup>3</sup>	83.5 e <sup>3</sup>	o <sup>4</sup>
Rosclife	2%	118.3 bcd	145.6 b	14.0+
Roselife	42	110.0 d	132.3 bc	14.0+
7 H Super	2 ml/1	127.4 a	114.3 d	1.0
7 H Super + sucrose	2 m1/1+27	126.2 ab	139.0 b	4.0
7 M Super	4 m1/1	128.2 a	117.5 cd	7.0
F M Super + sucrose	4 ml/1+27,	122.1 abc	144.0 b	4.5
Everbloom	2%	129.3 a	147.9 b	14.04
Everbloom	47.	114.3 cd	124.1 cd	14.0+
8-HQC+sucrose	200ppm+2%	129.5 a	165.4 a	14.0+

<sup>1</sup> Flower buds harvested when 50-55 mm in diameter. Initial stem fresh weights were 27-37 gmms.

<sup>2</sup>Weights are expressed as percentage values of initial weight. Initial weight equals 100%.

<sup>3</sup>Means within a solumn followed by different letters are significant at the 17 level.

<sup>4</sup>Data allow for 6 days to open flowers. Experiment terminated after 20 days of vase life,

crease in fresh weight for 7 days except those held in 2 or 4 ml F M Super or water which reached their maximum weight in 3 days (Table 4). Stems held in the latter 3 solutions lost weight after the third day. Stems held in 8+ HQC-sucrose gained more weight after 7 days than stems held in other preservatives. Two percent Everbloom, 2 and 4% Roselife, and 2 and 4 ml FM Super plus 2% sucrose were equally effective in increasing fresh weight.

Everbloom, Roselife, and 8-HQC+sucrose were more effective in prolonging vase-life than F M Super (Table 4). Flowers held in F M Super, with or without sucrose, lasted 1 week or less but flowers held in other preservatives lasted a minimum of 2 weeks.

Experiment 3 .--- Generally, flower diameter of all spray cultivars was greater when stems were held in 8-HQC+sucrose than in water (Fig. 1). Flowers that developed from "tight" buds and were held in 8-HQC+sucrose had similar diameter and form as flowers that developed from "open" buds. Maximum flower diameter of "tight" and "open" buds occurred at 7 days, but the rate of flower development was greater for "tight" buds. 'Shasta' had the largest flowers followed respectively by 'Iceberg' and 'Beauregard' with smaller flowers. Flowers from 'open' buds of 'Beauregard' decreased slightly in diameter after 3 days. Decrease in diameter was due to outer petals reflexing and was not due to premature wilting. Although flowers from all cultivars held in water increased in diameter, flower form was not typical of flowers which open on the plant. Flowers of each cultivar held in 8-HQC+sucrose were similar in form and appearance to those which develop on the plant.

Stems of all cultivars held in 8-HQC+sucrose increased in fresh weight for 10 days (Fig. 2). Stems held in water reached their maximum fresh weight in 3 days. Stage of flower bud development did not influence changes in fresh weight as did 8-HQC+sucrose. Eight-hydroxyquinoline citrate plus sucrose not only produced the largest flowers but also produced the heaviest stems.

Flowers held in 8-HQC+sucrose were fully turgid and of excellent quality after 2 weeks but foliage showed incipient chlorosis. Flowers held in water for 2 weeks were beginning to wilt but foliage did not show signs of chlorosis.



Figure 1.—Flower diameter of several chrysanthemum cultivars harvested at 2 stages of development and held in water or 200 ppm 8-hydroxyquinoline citrate plus 2% sucrose. 1) tight bud held in water, 2) tight bud held in 8-HQC+sucrose 3) open bud held in water, 4) open bud held in 8-HQC+sucrose.

#### DISCUSSION

These experiments indicate that chrysanthemum flowers could be harvested as buds and opened in commercial floral preservatives. Considering flower diameter, fresh weight, and vase life, the best floral preservatives were 8-HQC+ sucrose and Everbloom. Flowers opened in Roselife were commercially acceptable but did not attain the weight or size as flowers held in 8-HQC+sucrose. Flowers opened in F M Super had much shorter vase-life than those opened in 8-HQC+sucrose or Everbloom. Possibly different concentrations of F M Super or Roselife might increase flower longevity or size. Flowers held in Bloomlife or water opened poorly and did not have commercially acceptable symmetry. Chlorosis was evident on foliage of stems held in preservatives. Chlorosis appeared 17-20 days after harvest. This agrees with Waters' report (15) that Everbloom induced chlorosis in chrysanthemum foliage. Marousky (13) reported that 8-HQC+sucrose induced chrysanthemum foliar



Figure 2.—Fresh weight of several chrysanthemum cultivars harvested at 2 stages of development and held in water or 200 ppm 8-hydroxyquinoline citrate plus 2% sucrose. 1) tight bud held in water, 2) tight bud held in 8-HQC+sucrose, 3) open bud held in water, 4) open bud held in 8-HQC+sucrose.

chlorosis after 10-18 days and indicated that sucrose was responsible for foliar chlorosis.

Flower buds of spray chrysanthemum cultivars held in 8-HQC+sucrose followed similar patterns of development reported for large incurved flowers (13). Flower buds of spray cultivars held in 8+HQC-sucrose opened and weighed more than flowers held in water. Flowers on stems held in 8-HQC+sucrose were similar in semmetry to flowers which developed on the intact plant.

These data suggest that flowers of spray cul-

tivars could be harvested, handled, and opened in a manner similar to that used with the large incurved chrysanthemum cultivars. Commercially available floral preservatives might be used for opening bud-cut chrysanthemums in a possible new marketing procedure. Commercial application of this procedure could reduce cost of transportation, facilitate harvest, and reduce damage. Because of the range of bud size that can be opened successfully, mechanically harvesting might be considered.

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#### LITERATURE CITED

Aarts, J. F. T. 1957. Over de houbaarheid van snij-blocmen (On the keepability of cut flowers). Mededelingen van de Landbouwhogeschool, Wagenigen, Holland 57: 1-62.
 Anonymous. 1969. Bud-cut carnations nip shipping costs. U. S. Dept. Agr. Res. 17(9): 12.
 Besemer, S. T. 1961. Shipping methods used for fresh cut-flowers. Calif. Agr. 15(3): 6-7.
 Holley, W. C. and Cheng, L. 1967. Harvesting and handling of immature carnations. Proc. Amer. Soc. Hort. Sci. 90: 377-383.
 Kohl, H. C., Jr. and Smith, D. E. 1960. Develop-

Sci. 90: 377-383.
5. Kohl, H. C., Jr. and Smith, D. E. 1960. Development of carnation flowers cut before fully open. Carnation Craft. 53: 7-8.

6. Larsen, F. E. and Cromarty, R. W. 1967. Micro-organism inhibited by 8-hydroxyquiniline citrate as related to cut flower senescence. Proc. Amer. Soc. Hort. Sci. 90: 546-549.

546-549. 7. Larsen, F. E. and Scholes, J. F. 1965. Effects of succinamic acid on vase-life and quality of cut carnations. Proc. Amer. Soc. Hort. Sci. 87: 458-463. 8. Larsen, F. E. and Scholes, J. F. 1966. Effects of 8-hydroxyquinoline citrate, N-dimethyl amino succinamic acid and sucrose on vase life and spike characteristics of cut snapdragons. Proc. Amer. Soc. Hort. Sci. 89: 694-701. 9. Larsen, F. E. and Scholes, J. F. 1966. Effect of 8-hydroxyquinoline citrate, sucrose, and Alar on vase life and quality of cut stocks. Flor. Rev. 139 (3608): 46-47, 10. Marousky. F. J. 1969. Var.

10. Marousky, F. J. 1968. Vascular blockage, water ab-sorption, stomatal opening and respiration of cut 'Better Times' roses treated with 8-hydroxyquinoline citrate and

Lines' roses treated with 8-hydroxyquinoline citrate and sucrose. Jour. Amer. Soc. Hort. Sci. 94(8): 223-226. 11. Marousky, F. J. 1968. Physiological role of 8-hy-droxyquinoline citrate and sucrose in extending vase-life and improving quality of cut gladiolus. Proc. Fla. State Hort. Soc. 81: 409-414. 12. Marousky, F. J. 1968. Influence of 8-hydroxyquino-line citrate and sucrose on vase-life and curvity of cut sucression.

MATOUSKY, F. J. 1905. Initiate of o-hydroxyduno-line citrate and sucrose on vase-life and quality of cut gladiolus. Proc. Fla. State Hort. Soc. 81: 415-419.
 Marousky, F. J. 1969. Handling and opening bud-cut chrysanthemum flowers with 8-hydroxyquinoline citrate and sucrose. U. S. Dept. Agr., Mkt. Res. Rpt. Submitted for multication.

and sucrose. U. S. Dept. Agr., Mkt. Res. Rpt. Submitted for publication.
14. Scholes, J. F. 1963. Some effects of various chemi-cals on the post-harvest physiology of Velvet Times roses.
M. S. Thesis. Cornell University, Ithaca, New York.
15. Waters, W. E. 1964. Influence of chemical pre-servative on keeping quality of asters, carnations, chrysan-themums and gerbera daisies. Proc. Fla. State Hort. Soc. 77: 466-470.

# EFFECT OF RATIOS OF NH+ TO NO3 AND LEVELS OF N AND K ON CHEMICAL CONTENT OF CHRYSANTHEMUM MORIFOLIUM. "BRIGHT GOLDEN ANN"

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#### ABSTRACT

Three 5x3 factorial experiments in randomized block design with 4 replications were initiated under soil culture conditions to test effects of 5 ratios of NO3 versus NH4 nitrogen at 3 N-K levels on growth and chemical composition of Chrysanthemum morifolium 'Bright Golden Ann'. Only 8 of the 15 treatments survived. Samples of leaf tissue from plant mid-section were taken for chemical analyses 6 and 8 weeks after potting and at experiment termination. Termination samples also were analyzed for total oxalate content.

Plants receiving higher ratio of  $NO_3$  to  $NH_4$ at high N-K level and those receiving higher ratio of NH<sub>4</sub> to NO<sub>3</sub> at low N-K were tallest.

Plants receiving NH<sub>4</sub>-N had higher N contents than those receiving NO<sub>3</sub>-N. Increasing NH<sub>4</sub> ions depressed uptake of Ca. Increasing No3 ions depressed P uptake. K was least effected by form of N. NO3-N promoted oxalate formation.

#### INTRODUCTION

Effects of different levels of N fertilization had been studied and reported for a large variety of plants, but the form of N supplied to plants has received less consideration. A few investigators have reported differences in growth responses and chemical composition of plants given NO<sub>3</sub>- versus NH<sub>4</sub>-N.

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