

Black salt marsh mosquito *Aedes taeniorhynchus* (Wiedemann) (Insecta: Diptera: Culicidae)¹

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Introduction

The black salt marsh mosquito, *Aedes taeniorhynchus* (Wiedemann), is very common in the eastern coastal areas of the Americas, and is responsible for a large part of mosquito insecticide applications in Florida (Connelly and Carlson 2009, Koehler and Castner 2008). Although it is not a primary vector of major concern, it can transmit pathogens to humans and other animals. Its characteristic emergence in large numbers after rains and flooding events as well as its aggressive biting contribute to its notoriety as a pest insect. The black salt marsh mosquito is considered a nuisance in Florida. It is sheltered from large-scale mosquito control as part of the Everglades National Park conservation program to preserve their delicate ecosystem (Day et al. 2004).



Figure 1. Bloodfeeding female *Aedes taeniorhynchus* Credits: Sean McCann, University of Florida



Figure 2. Adult female *Aedes taeniorhynchus*. Credits: Michelle Cutwa, University of Florida



Figure 3. Adult male *Aedes taeniorhynchus*. Credits: Natasha Agramonte, University of Florida

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Synonymy

- Aedes (Ochlerotatus) taeniorhynchus Wiedemann 1821
- Culex taeniorhynchus Wiedemann 1821
- Culex damnosus Say, 1823
- Taeniorhynchus niger Giles, 1904
- Culex portoricensis Ludlow, 1905
- Aedes epinolus Dyar & Knab, 1914
- Aedes pix Martini 1935

(ITIS 2014, WRBU 2014)

Distribution

Aedes taeniorhynchus is found in the brackish, marshy areas of North, Central, and South America including the islands of the Caribbean. Although the larval habitats are typically restricted to the coastal salt marshes, this species has a long flight range and can be found far inland at times, particularly in Florida where it can be found throughout the state (Connelly 2006, Rey and Connelly 2010).



Figure 4. Mosquito control worker sampling larvae in a marsh as part of a surveillance program.

Credits: Florida Keys Mosquito Control District



Figure 5. Aedes taeniorhynchus distribution. Credits: Natasha Agramonte, University of Florida

Description and Life Cycle

Like other true flies, *Aedes taeniorhynchus* has four distinct life stages: adult, egg, larva, and pupa (Borror and White 1970).

Adults

Like other flies (order Diptera), *Aedes taeniorhynchus* possess a pair of wings for flight and a pair of knobby halteres for directional perception and stability. Like other mosquitoes, black salt marsh mosquitoes have long, narrow wings with scales along the wing veins. Female mosquitoes of the subfamily Culicinae, like *Aedes taeniorhynchus*, also possess palps shorter than the proboscis. The mouthparts of mosquitoes are made up of a pair of stylets for piercing and a feeding tube for sucking. Collectively these mouth parts are refered to as a proboscis.

Male and female mosquitoes can be distinguished by their antennae. Males have feather-like or plumose antennae, while females have antennae with only a few hairs (Borror and White 1970). Bands of white scales found in characteristic body locations are useful markings for identification of this species. The white banding on the basal section of abdominal segments, typical of *Aedes* mosquitoes, as well as the white coloration on the tip of the palps and a ring of white scales on the middle of the proboscis, can be used to distinguish *Aedes taeniorhynchus* from similar species (Darsie and Morris 2003).

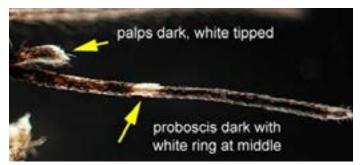


Figure 6. *Aedes taeniorhynchus* adult proboscis. Credits: Michelle Cutwa, University of Florida

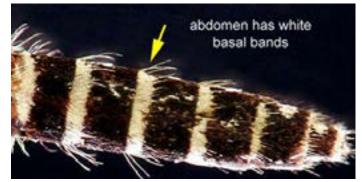


Figure 7. Aedes taeniorhynchus adult abdomen. Credits: Michelle Cutwa, University of Florida

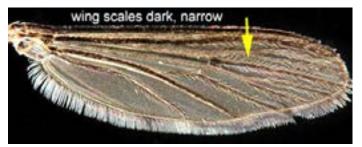


Figure 8. *Aedes taeniorhynchus* adult wing. Credits: Michelle Cutwa, University of Florida

Mating takes place soon after emergence, often within a few days (Foster and Walker 2009). Adults of this species can feed during the day if resting sites are disturbed (Rey and Connelly 2010), although feeding primarily occurs at night (Connelly 2006). While both males and females may nectar-feed, only females are haematophagous, or blood feeders. Mammals and birds are the preferred hosts for this species (Connelly 2006), but females will readily bite humans. Females generally take a bloodmeal before ovipositing; however, this species is partially autogenous, whereby it can lay an initial clutch of eggs without a bloodmeal, unlike most mosquitoes (Lea 1964).

Eggs

Eggs are laid in moist soil, not directly into water, in areas that are sheltered and high enough above the water line to be affected only by sufficient flooding, which will trigger eggs to hatch (Ritchie and Johnson 1991). Additionally, eggs may be deposited in, and hatched from, container sites



Figure 9. *Aedes mosquito* egg. Credits: Centers for Disease Control, #688

similar to *Aedes aegypti* (Linnaeus), although this is likely an infrequent occurrence with this species (Rey and Connelly 2010). Egg hatching depends on flooding by rainfall or tides; however, once this occurs, larval development proceeds rapidly.

Larvae



Figure 10. *Aedes taeniorhynchus* 4th instar larva. Credits: Natasha Agramonte, University of Florida

The larval stage consists of four instars that may take anywhere from 5 to 15 days to complete, based on the ambient temperature. Larval development can occur in any salinity, from fresh water to ocean water (Nielsen and Nielsen 1953).

Pupae

The pupal stage follows the fourth instar, and lasts for only a couple of days, after which the adults emerge. Pupae are active swimmers but do not feed during this life stage.



Figure 11. *Aedes taeniorhynchus* pupa. Credits: Natasha Agramonte, University of Florida



Figure 12. *Aedes taeniorhynchus* adult female emerging from pupa. Credits: James M. Newman, University of Florida

EEE virus is closely related to both Venezuelan equine encephalitis (VEE) virus and Western equine encephalitis (WEE) virus, which occasionally occur in Florida. As the name suggests, EEE acutely afflicts horses in the eastern United States, but it can also strike humans during seasonal outbreaks (Drew 2001). Both humans and horses are dead-end hosts for this virus because infected individuals do not produce sufficient viremia to infect mosquitoes. The EEE virus is primarily maintained in a cycle between the mosquito Culiseta melanura (Coquillett) and bird hosts in swamp habitats (Buckner et al. 2013). The transmission of EEE to humans requires different mosquito species, such as the black salt marsh mosquito, which can act as bridge vectors between infected birds and uninfected mammals, although other mosquitoes are more competent vectors (CDC 2009). Similarly, although Aedes taeniorhynchus is able to transmit VEE virus, Aedes sollicitans (Walker) is a better vector due to higher transmission rates and propensity for infection (Turell et al. 1992).



Medical and Veterinary Significance

Aedes taeniorhynchus is medically relevant, primarily as a vector of two alphaviruses from the family Togaviridae, Eastern equine encephalitis (EEE) and Venezuelan equine encephalitis (VEE) (Drew 2001). Additionally, black salt marsh mosquitoes have also been known to transmit the filarial worm *Dirofilaria immitis*, commonly known as the dog heartworm.

Figure 13. EEE and VEE disease distribution in the Americas. Credits: Natasha Agramonte, University of Florida

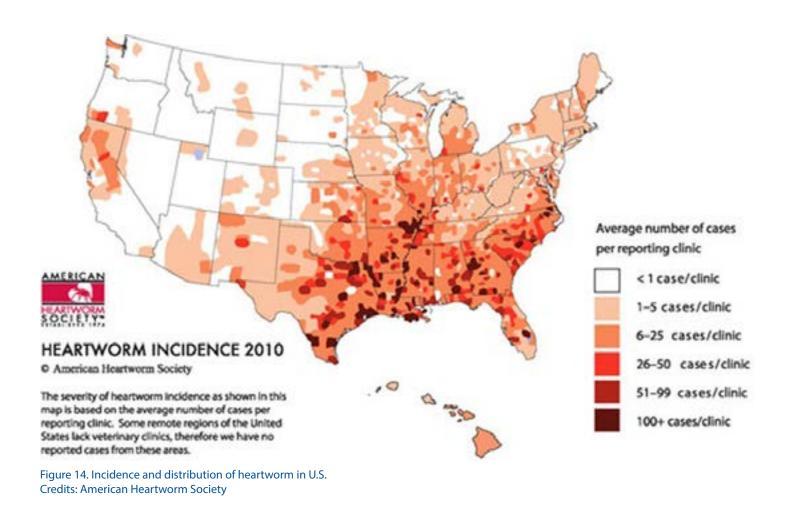
Signs and symptoms in encephalitic patients start within 3-7 days of infection and include fever, headache, irritability, restlessness, drowsiness, anorexia, vomiting, diarrhea, cyanosis, convulsions, and coma (Drew 2001, CDC 2009). Approximately one in three people who develop encephalitic infections die (VEE and EEE), and those who survive will often have mild to severe permanent neurologic damage (CDC 2009); by contrast, the systemic infection of EEE displays mild symptoms for 1-2 weeks and has no lasting effects. Despite only causing about 6 deaths annually in the United States, control of EEE vectors is of particular concern to public health officials (CDC 2009).

In the U.S., black salt marsh mosquitoes are one of 16 species identified as competent hosts of *Dirofilaria immitis*, 11 of which are found in Florida (Nayar and Rutledge 2008). In the United States, transmission of heartworms to dogs most often occurs during the late spring and summer months when mosquito production is high. In the southern states, such as Florida, higher temperatures facilitate year-round transmission of this pathogen and present a risk to both wild and domestic animals.

Management

Control strategies for management of salt marsh mosquito populations are complex and often involve integrated mosquito management strategies that utilize source reduction (the elimination of mosquito breeding sites), surveillance (a measurement of the mosquito population size and species present), larviciding (the use of chemical or biological control on mosquito larvae), and adulticiding (the use of chemical control on mosquito adults). Source reduction can be accomplished through water management strategies which include impounding, ditching, and open marsh water management (OMWM) techniques, all of which aim to reduce larval mosquito populations by reducing the available larval habitat or by encouraging consumption of larvae by native larvivorous (larvae-eating) fish (Meredith and Lesser 2007, Rey and Connelly 2001).

Chemical control can also be used and may involve the application of insecticides in the aquatic habitats where



mosquito larvae are found or the aerial applications of insecticides to reduce adult populations of mosquitoes over marshes or in areas adjacent to human activity (Meredith and Lesser 2007, Rey and Connelly 2001).

Personal protection techniques, such as the use of insect repellents containing US-EPA registered active ingredients, are encouraged for individuals to protect exposed skin in areas that may have mosquitoes (Xue et al. 2007). For individuals that spend a lot of time outdoors, long-sleeved clothing pre-treated with insecticide is commercially available as an added measure of protection for individuals, but should be used in combination with a repellent to be maximally effective (Xue et al. 2007). Screened windows and doors can also provide added protection to a home by acting as a physical barrier to the invasion of biting insects. Removal of mosquito larval habitats by emptying standing water from planters, buckets, kiddie pools, bromeliads and other containers is also a good general exercise in mosquito control. Although destroying container-breeding sites will have a minimal effect on Aedes taeniorhynchus, this practice



Figure 15. Mosquito control professionals digging a drainage ditch for mosquito source reduction. Credits: Alameda Mosquito Control Abatement District



Figure 16. Two planes applying an aerial insecticide treatment to a marsh. Credits: Florida Keys Mosquito Control District

can reduce the numbers of other mosquito species found around one's home.

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