South American Cucurbit Fruit Fly, *Anastrepha grandis* (Macquart) (Insecta: Diptera: Tephritidae)

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**Introduction**

The South American cucurbit fruit fly, *Anastrepha grandis* (Macquart), which exists in several South American countries, attacks watermelon and other fruits of the family Cucurbitaceae. Once a pest of minor to moderate importance generally, it has become a rather important pest. The pest status differs in each country and has sometimes changed within a country. This species is potentially of economic importance in Florida and southern Texas should it ever be introduced there. It has been intercepted in the United States in pumpkin from Argentina and Brazil, and in banana debris from Panama.

**Synonymy**

*Acrotoxa grandis* (Macquart)

*Anastrepha schineri* Hendel

*Anastrepha latifasciata* Hering

*Tephritis grandis* Macquart

*Trypeta grandis* (Macquart)

**Distribution**

This species is established in South America: Argentina, Bolivia, Brazil (except north), Colombia, Ecuador, Panama, Paraguay, Peru, and Venezuela (NAPPO 2009).

**Identification**

Rather large, yellow-brown, with yellow and dark-brown markings. Mesonotum is 3.3–4.0 mm long, yellow-brown, with humerus, median stripe widening to include acrostichal bristles but not reaching scutellum, lateral stripes from just before transverse suture to side of scutellum, stripe below notopleuron, metapleuron, and scutellum except extreme base yellow; a sublateral stripe from level of humeral bristle to scutellum, broken at transverse suture, a band along scutoscutellar suture, intensified mediadly, and a spot on pteropleuron dark brown; metanotum blackened laterally. Macrochaetae dark brown; pile yellowish brown. No sternopleural bristle. The wing is 9.0–10.5 mm long, the bands yellow brown, rather diffuse; costal and S bands broadly connected, and no distinct hyaline spot anterior to vein R4+5; distal arm of V band absent, the proximal arm not joining S band.

Female terminalia: The ovipositor sheath is 5.8–6.2 mm long, tapering posteriorly to apical third, which is distinctly depressed and broadened; in profile the sheath is distinctly concave dorsally on median half and concave ventrally on apical third. Rasper well developed, of slender, curved hooks in five or six rows. Ovipositor slightly longer than length of ovipositor sheath, being somewhat curved dorsoventrally to permit fitting into sheath; tip long and slender, without serrations; extreme base slightly widened (Stone 1942).
Steyskal (1977) distinguished adult *A. grandis* from the other four species of the *Grandis* group as follows:

1. Vein R₃ somewhat undulant; metanotum yellow . . . . *A. bezzii* Lima and *A. balloui* Stone

1’ Vein R₃, not undulant; metanotum marked with black . . .

2. Mesonotum not striped with black . . . . *A. atrigona* Hendel

2’ Mesoscutum with black stripes . . . . 3

3. Mesoscutum and pleura largely black, wing with pattern darkened . . . . *A. shannoni* Stone

3’ Mesoscutum and pleura with little black, wing pattern very little darkened . . . . *A. grandis* (Macquart)

**Hosts**

- *Citrullus lanatus*, watermelon
- *Cucumis melo*, “honeydew melon”
- *Cucumis sativus*, cucumber
- *Cucurbita maxima*, autumn and winter squash, pumpkin
- *Cucurbita pepo*, summer and autumn pumpkin and squash, gourd, marrow
- *Cucurbita moschata*, butternut squash and pumpkins
- *Lagenaria siceraria*, bottle gourd or calabash

It has also been known to attack common guava, *Psidium guajava* (Norrbom 2004). The record of rearing from oranges (Greene 1934) is erroneous. Immature fruits of cucurbits apparently are preferred, but mature or nearly mature fruits of some cucurbits are attacked occasionally.
Survey and Detection

Adults are easily recognized by their relatively large size (wing length 10 mm), long aculeus (a sharp spine jutting from beneath the margin of the eight sternite) and rather diffuse wing markings which are not of the typical Anastrepha type.

Larvae can be collected from infested fruit and are very difficult to identify to species except when reared to adults. For best larval preservation, kill in boiling water, place in 50% isopropyl alcohol for two days, then in 75% isopropyl alcohol. Adults may be collected on stickyboard and in baited traps.

Selected References


