

# A Flesh Fly *Sarcophaga crassipalpis* Macquart (Insecta: Diptera: Sarcophagidae)<sup>1</sup>

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## Introduction

Flesh flies (Diptera: Sarcophagidae) are commonly found in warm tropical areas, although they do occur in places not suitable for most flies. They are closely associated with humans (synanthropic) and they have been known to enter houses to colonize cadavers (Pohjoismäki 2010). Like their name entails, these flies depend on live or dead tissue to complete their life cycle (Watson and Dallwitz 2003). This family of flies is attracted to many types of dead vertebrate remains, including humans (Nishida 1984). Of the 2,000 known species in this family, 327 are in the United States. *Sarcophaga crassipalpis* Macquart is commonly used in laboratories to study gene expression, diapause processes, and physiological processes (Lee and Denlinger 1985). It also has a significant impact in other areas of science including parasitology and forensic entomology.



Figure 1. Anterior lateral view of an *Sarcophaga crassipalpis* Macquart, a flesh fly. Fly is on glass, which reflects some of the legs.  
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## Synonymy

*Sarcophaga crassipalpis* Macquart, 1839

## Distribution

This species was originally found in temperate climates (Chen 1990) such as the Palearctic ecozone (Iberian peninsula, Arabian peninsula, Japan) (Velasquez et al. 2010), but is now established in the Nearctic ecozone, which includes most of North America and Greenland. Reports have also documented *S. crassipalpis* in the Neotropic ecozone (southern Florida, Central and South America, and the Caribbean). Due to its vast geographical distribution, this species has been described as “nearly cosmopolitan” (Cole 1969). Although the probability of laboratory lines of this species escaping and proliferating in the wild is minimal, fresh wild strains can be collected and incorporated into laboratory colonies due to its cosmopolitan distribution.

## Description

### Adult

The adult ranges in size from 9 to 13 mm. This fly is typically a light grayish color with three black stripes on the thorax (Watson and Dallwitz 2003). Unlike females, males are more hairy, with robust front legs, that assist in their attempts to copulate. Like many flesh fly species, their “tail” or abdominal end is red for both males and females. The front femora are slightly larger than those on the remaining

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legs and often have a light gray coloration. Adults also have a distinctive black strip with golden or yellowish margins between their eyes.



Figure 2. Dorsal view of the three thoracic stripes on *Sarcophaga crassipalpis* Macquart, a flesh fly. The head is at the top of the image.  
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Figure 3. Lateral view of the abdomen of *Sarcophaga crassipalpis* Macquart, a flesh fly. The head is to the left of the image.  
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Figure 4. Frontal view of the head of *Sarcophaga crassipalpis* Macquart, a flesh fly.  
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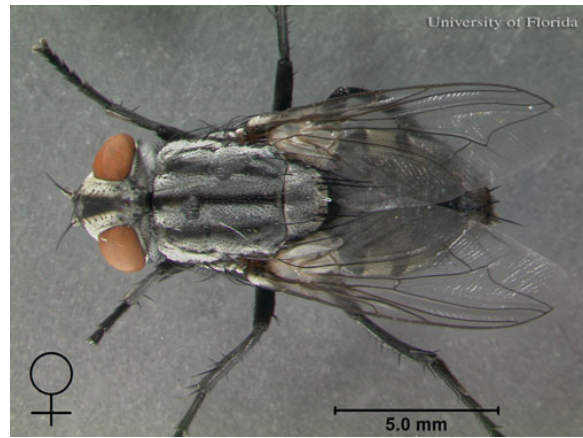


Figure 5. Dorsal view of adult female *Sarcophaga crassipalpis* Macquart, a flesh fly.  
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Figure 6. Dorsal view of adult male *Sarcophaga crassipalpis* Macquart, a flesh fly.  
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Figure 7. Ventral view of adult female *Sarcophaga crassipalpis* Macquart, a flesh fly.  
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## Pupa

The pupa ranges in size from 5 to 10 mm, and color tends to be relative to the age of the pupa. In general the darker the color the more advanced the fly is in pupal development.

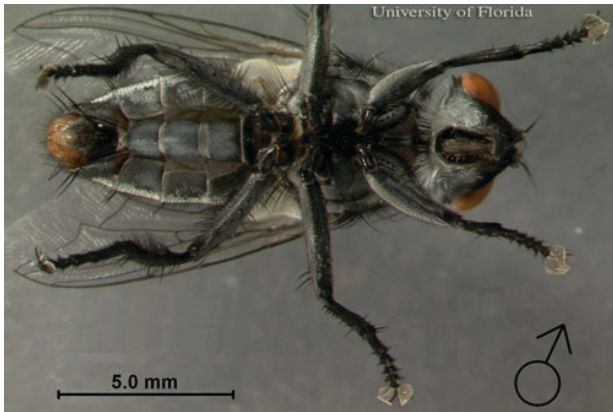


Figure 8. Ventral view of adult male *Sarcophaga crassipalpis* Macquart, a flesh fly.  
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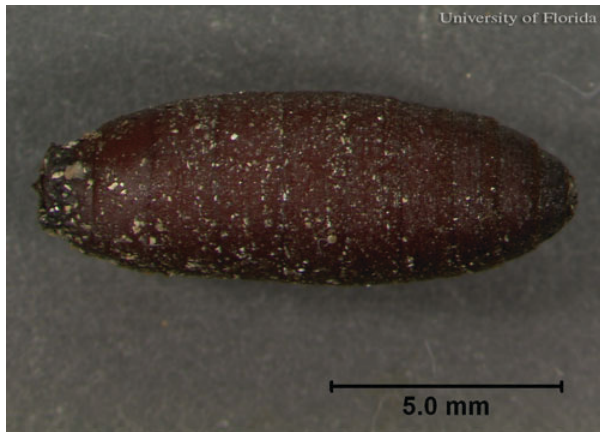


Figure 9. Dorsal view of a pupa of *Sarcophaga crassipalpis* Macquart, a flesh fly, with scale size. The head is to the right.  
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Figure 10. Darkening pupa coloration in *Sarcophaga crassipalpis* Macquart, a flesh fly, indicates advancing development within that stage of the life cycle.  
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## Larva

In the final instar, the larval body ranges from 9 to 13 mm in length. Spiracles are located on plates set inside a cavity or pit on the posterior end.

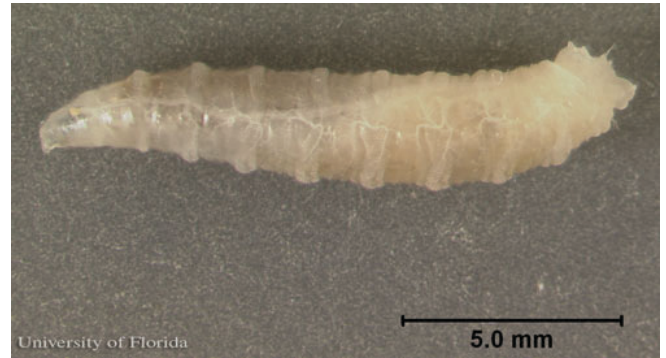


Figure 11. Lateral view of a larva of *Sarcophaga crassipalpis* Macquart, a flesh fly, with scale size. The head is to the left.  
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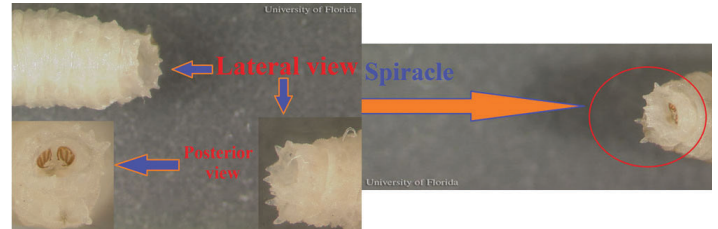


Figure 12. Multiple views of posterior end and spiracles of larvae of *Sarcophaga crassipalpis* Macquart, a flesh fly.  
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## Life Cycle

### Adult

Flesh flies are ovoviviparous, which means that eggs are not deposited upon full development. Instead, the larvae hatch inside of their mother's "uterus" and are held until a proper host is found. The term used to describe the release of the larvae onto the host is 'larviposition.'



Figure 13. Lateral view of adult *Sarcophaga crassipalpis* Macquart, a flesh fly.  
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### Eggs

The eggs undergo full development and hatch within the female. The images below are of eggs dissected from females prior to hatching.



Figure 14. Lateral view of an egg of *Sarcophaga crassipalpis* Macquart, a flesh fly, with scale size.

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Figure 15. Lateral view of an egg cluster of *Sarcophaga crassipalpis* Macquart, a flesh fly.

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## Larvae

Female flesh flies deposit their 1st instar larvae directly on the host, and the larvae commence feeding immediately. These larvae eat and develop rapidly. Approximately five days after larviposition, the larvae are already in their 3rd instar and are almost ready to pupate. When the larvae are ready to pupate, they leave the host and wander until they find a suitable location.

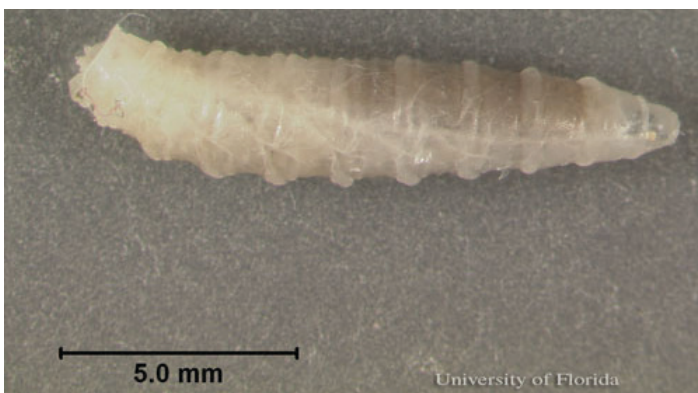


Figure 16. Lateral view of 3rd instar larva of *Sarcophaga crassipalpis* Macquart, a flesh fly, with scale size. The head is to the right.

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Figure 17. Dorsal view of the pre-pupa larval stage of *Sarcophaga crassipalpis* Macquart, a flesh fly. The head is to the left.

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## Pupae

Pupation starts approximately one week after larviposition at 25°C. Adult flies will emerge about 10 days after pupation has occurred. *Sarcophaga crassipalpis* have facultative diapause, meaning that they are able to enter, if necessary, a “hibernation”-like state where metabolic processes are reduced almost to a complete stop until appropriate environmental conditions return (Henrich and Denlinger 1982). In *S. crassipalpis*, diapause occurs during the pupal stage. Diapause can be pre-determined by environmental cues such as change in temperature, humidity, and/or light availability experienced by the mother prior to larviposition. The larva's environmental perception after larviposition also can induce diapause.



Figure 18. Dorsal view of a dark (more mature) pupa of *Sarcophaga crassipalpis* Macquart, a flesh fly. The head is to the right.

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## Host

The diversity of hosts used by sarcophagids is almost as great as the number of species within the family. Some flesh fly species are parasitoids, attacking and developing within other arthropods (Coppel 1959), while most have been found on varying types of carrion: pig carcasses, human cadavers, and/or wildlife (Goff 1991). Although *S. crassipalpis* is known as a carrion-inhabiting species, they have been reported to cause myiasis in humans (Morris 1987). Myiasis is a parasitic condition in which the maggots infest a living

host, in this case a human being, and feed on the living or dead tissue present.

## Importance

Flesh flies commonly colonize human remains early in the decay process (Nishida 1984). Although they are not the predominant type of fly found in situations of forensic importance, they can be valuable evidence when present (Nishida 1984, Goff 1991). Sarcophagids are a very useful tool for the forensic entomologist; like many other insects, they can be used to estimate the time of colonization, which can be used to infer a minimum postmortem interval (m-PMI) (Nishida 1984, Oliveira-Costa and Mello-Patiu 2004). Weather conditions and geographic location are key factors to consider when using insect evidence to estimate the m-PMI of a corpse as the variation associated with time of colonization after death is not completely understood at this time (Catts 1992).

This particular species of flesh fly is commonly thought of as the “lab rat,” and therefore it is usually overlooked when dealing with issues of forensic entomology. Over the years scientists have conducted and documented countless experiments on *S. crassipalpis*. Most of these experiments were studies regarding diapause and physiological responses to environmental changes (temperature, light, and humidity changes) (Nishida 1984, Velasquez et al. 2010). The data accumulated regarding sarcophagid development under different environmental conditions are vast (Grassberger 2002), making flesh flies an effective tool to accurately calculate a postmortem interval range (Nishida 1984, Goff 1991).

## Management

As *S. crassipalpis* and other sarcophagid species are usually not abundant, a flyswatter typically will suffice to eliminate a few flies. This species is not known to be a major pest. Should this fly become a problem, the proper removal of carcasses will disrupt their life cycle by inhibiting larval development. The removal of vegetative refuse, a resting site for the adults, will reduce the annoyance of resident adult populations and risk of recontamination (Koehler and Oi 2003). For more information on management and prevention techniques see the:

[Florida Insect Management Guide for filth-breeding flies.](#)

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